

**ANALYTICAL STRATEGY BASED ON THE USE OF LIQUID
CHROMATOGRAPHY AND GAS CHROMATOGRAPHY WITH TRIPLE
QUADRUPOLE AND TIME-OF-FLIGHT MS ANALYZERS FOR
INVESTIGATING ORGANIC CONTAMINANTS IN WASTEWATER**

E. Pitarch¹, T. Portolés¹, J.M. Marín¹, M. Ibáñez¹, F. Albarrán², F. Hernández^{1*}

¹Research Institute for Pesticides and Water, University Jaume I, 12071 Castellón,
Spain

Tel: +34 964 387366, Fax: +34 964 387368, E-mail: felix.hernandez@qfa.uji.es

²RECIPLASA, Reciclados de Residuos La Plana, S.A., 12200 Onda, Castellón, Spain

ABSTRACT

The presence of a wide variety of organic pollutants with different physico-chemical characteristics has been investigated in wastewater samples from a municipal solid waste treatment plant placed at Castellón, Spain. An advanced analytical strategy has been applied, consisting on the combined use of two powerful and complementary techniques, GC and LC, both hyphenated with tandem mass spectrometry with triple quadrupole analyzers. The GC-MS/MS method was based on sample extraction using C₁₈ SPE cartridges and allowed the determination of around 60 compounds from different chemical families, such as PAHs, octyl/nonyl phenols, PCBs, organochlorine compounds, insecticides, herbicides and PBDEs. Most of compounds selected are included as priority contaminants in the European Union (EU) Water Directive. The UHPLC-MS/MS method, which provided high chromatographic resolution and sensitivity and short analysis time, used a sample extraction with OASIS HLB SPE cartridges and allowed the determination of 37 (more polar) pesticides.

The methodology developed has been applied to the analysis of 41 water samples (20 non-treated, raw leachates, and 21 treated) collected between March 2007 and February 2009. Treated (reverse osmosis) water samples analyzed rarely exceeded 0.5 µg/L for the contaminants investigated. As expected, in non-treated leachates the number of detections and the concentration levels found were notably higher than in treated waters. The most commonly detected pollutants were herbicides (simazine, terbuthylazine, terbutryn, terbumeton, terbacil and diuron), together with fungicides (thiabendazole and carbendazim) and 4-t-octylphenol. In the light of data obtained, it has been proven that reverse osmosis process used for water treatment was efficient and notably reduced the levels of organic contaminants found in raw leachate samples.

In order to investigate the presence of other non-target contaminants, water samples were also analyzed by using GC-TOF MS and LC-QTOF MS. Several organic pollutants that did not form a part of the previous list of target contaminants were identified in the samples, thanks to the good sensitivity of TOF MS in full spectrum acquisition mode and the valuable accurate mass information provided by these instruments. The insecticide diazinon, the fungicide diphenylamide, the UV filter benzophenone, N-butyl benzenesulfonamide (N-BBSA), the insect repellent diethyltoluamide, caffeine or pharmaceuticals like erythromycin, benzenesulfonamide, ibuprofen, atenolol or paracetamol, were some of the compounds identified in the water samples analyzed.

Keywords

Organic pollutants, wastewater, UHPLC, GC, tandem MS, TOF MS

INTRODUCTION

Nowadays, many organic contaminants can be present in environmental water, normally at the $\mu\text{g/L}$ level or below [1]. One of the routes for the contaminants to enter into the aquatic environment is from municipal solid waste landfill leachates. These leachates frequently contain a variety of hazardous chemicals which may cause severe biological effects in the aquatic environment, as many of them are highly toxic or even carcinogenic [2-3]. Therefore, efficient treatment of landfill leachates is required and monitoring of organic pollutants is compulsory to assure the quality of treated water. When possible, the treatment process should be performed in the Municipal Solid Waste (MSW) treatment plants, generally by the application of membrane technology, which is free of any chemical addition and uses relatively low energy. Membrane filtration, such as micro and ultra filtration or nano filtration, and reverse osmosis could be a choice for the treatment of landfill leachates, depending of the type of particles or salts to be removed.

The selection of the analytical methodology to be applied for water quality control is of outstanding relevance to obtain realistic results, especially in the analysis of treated water that are discharged into the aquatic environment. The development of sensitive and multi-class methods for determination of organic contaminants in wastewater has become a major issue, due to the presence of many different compounds in this type of samples and to the strict legal European Union requirements for water quality [4-6]. General reviews relating to water analysis and emerging environmental contaminants [7-10] have been published over the last 2 years reporting different analytical methodologies and new developments in this field.

Due to the complexity of the wastewater matrices, their high organic matter content, the low analyte levels typically found, and the high variety of organic contaminants with

quite different physico-chemical characteristics, the complementary use of gas chromatography (GC) and liquid chromatography (LC), both coupled to mass spectrometry (MS), is required to have a realistic and more complete overview of the organic pollution present in these waters. GC-MS has been the major adopted analytical technique to perform multi-residue analysis of volatile and semi-volatile organic pollutants [11]. Nowadays, enrichment via solid-phase extraction (SPE) using relatively low sample volumes followed by GC-MS, or even better, GC coupled to tandem mass spectrometry (MS/MS) is the preferred approach for GC-amenable micropollutants. Ion trap (IT) and triple quadrupole (QqQ) analyzers offer the possibility of adequate precursor and product ion selection, which allows improving sensitivity (reducing the chemical noise in the chromatograms) and selectivity. The use of two stages of mass analysis in MS/MS systems based on QqQ allows working on selected reaction monitoring (SRM) mode, one of the most selective and sensitive approaches at present for quantification and confirmation, especially in trace water analysis. Our own research group has recently reported the determination of more than 50 priority organic pollutants in water by GC-MS/MS with QqQ [12].

For more polar, less or non GC-amenable contaminants, LC-MS/MS is surely the most appropriate analytical technique [13-15], leading to satisfactory results from both the quantification and confirmation point of view. Recently, ultra high pressure liquid chromatography (UHPLC) has been developed as an innovative and powerful separation technique based on the use of stationary phases of particle size ($<2 \mu\text{m}$) smaller than in conventional HPLC. UHPLC coupled to MS/MS has been shown as an excellent analytical tool for multi-class analysis of water for compounds like pharmaceuticals and drugs [16-19], toxins [20] or pesticides [21, 22], due to its improved selectivity and sensitivity. With the modern QqQ analyzers even more than

two SRM transitions can be acquired for a safe identification without losing sensitivity. Despite the improved sensitivity when using UHPLC-MS/MS, the application of a pre-concentration step (e.g. based on SPE) is typically required in multi-class methods where a large number of contaminants are determined [16-22].

Despite the excellent performance of LC-MS/MS and GC-MS/MS methods, qualitative information that supports the recognition and structural elucidation of compounds other than target is still needed to obtain more information on actual water sample composition. Time-of-flight mass spectrometry (TOF MS) is an excellent technique for this purpose. TOF MS provides the selectivity and sensitivity required for an efficient and wide-scope screening, as it combines high full-spectral sensitivity with high mass resolution, allowing any LC-ionizable components in the sample (LC-TOF MS) or GC-amenable (GC-TOF MS) to be accurately mass-measured. TOF MS gives a notable amount of chemical information in a single analysis that allows searching for a high number of compounds after MS acquisition. Our own research group has recently reported several applications of both GC-TOF and LC-TOF for investigation of organic contaminants in water samples [23-27].

TOF MS is also a powerful technique for the investigation of non-target compounds, making feasible the elucidation of unknowns without any previous information or analyte selection. On the basis of these improved characteristics, GC has been combined with high-resolution TOF-MS (GC-HR-TOFMS) for non-target screening of GC-amenable organic (micro) pollutants in water [25, 26, 28, 29]. With regard to LC, very few applications using UHPLC-(Q)TOF MS have been reported in non-target field analysis [27].

The aim of this paper is to investigate the presence of a large number of organic pollutants in treated and raw non-treated leachates from a MSW treatment plant. Information on the quality of leachates after the treatment process is required to estimate the feasibility of discharging them into the aquatic environment. For this purpose, an analytical strategy consisting on the combined use of GC-MS/MS and UHPLC-MS/MS, both with triple quadrupole, has been applied in order to detect and quantify 94 target contaminants. Although most of selected analytes are considered as priority pollutants in water, their determination does not surely offer a realistic overview on the samples quality, as only a limited number of contaminants are determined. For this reason, all water samples were also analyzed by GC-TOF MS and LC-QTOF MS in order to widen the searching to other non-target contaminants, giving in this way useful information that could be applied to improve future monitoring programs.

EXPERIMENTAL

Reagents and chemicals

Reference standards of organic contaminants (see **Table 1**) were purchased from Dr. Ehrenstorfer (Augsburg, Germany), Wellington Laboratories (Guelph, Ontario, Canada), Fluka (Buchs, Switzerland), Riedel de Haën (Seelze, Germany) or Sigma (St. Louis, MO, USA).

Isotopically labelled surrogates used for GC-MS/MS were *p,p'*-DDE-d₈, lindane-d₆, benzo(a)anthracene-d₁₂ and terbuthylazine-d₅ (Dr. Ehrenstorfer) and hexachlorobenzene (HCB)-¹³C₆ (Cambridge Isotope Labs, Inc. Andover, MA, USA). Isotopically labeled surrogates used for UHPLC-MS/MS were dimethoate-d₆, 2-methyl-4-chlorophenoxy acetic acid (MCPA)-d₃, carbofuran-d₃, diuron-d₆, terbuthylazine-d₅, imazalil-d₅ and thiabendazole-d₆ (Dr. Ehrenstorfer).

To prepare calibration curves, working mix solutions of organic contaminants and isotopically labeled compounds were prepared in hexane or acetonitrile:water (10:90, v/v) for GC-MS/MS or UHPLC-MS/MS, respectively.

Acetone (residue analysis), acetonitrile (HPLC grade), ethyl acetate, dichloromethane and hexane (ultra-trace quality) were purchased from Scharlab (Barcelona, Spain). HPLC-grade water was obtained by purifying demineralized water in a Milli-Q Gradient A10 (Millipore, Bedford, MA, USA). Formic acid (HCOOH, content > 98%) and ammonium acetate (NH₄Ac, reagent grade) were supplied by Scharlab.

Cartridges used for solid phase extraction were 500 mg Bond Elut C₁₈ (Varian, Harbor City, CA, USA) and 200 mg Oasis HLB (Waters, Milford, MA, USA).

Sampling

Treated (21 samples) and raw leachate (20 samples) water samples were collected monthly from RECIPLASA, a Municipal Solid Waste (MSW) treatment plant sited at the Castellón province (Spain) along the period between March 2007 and February 2009. Treated water was submitted to a reversed osmosis process.

Raw leachate samples were 50-fold diluted with HPLC water before analysis due to their high organic matter content. All samples were stored in the dark at a temperature below -18°C. Before analysis, water samples were previously centrifuged at 3500 rpm for 10 min if suspended particulate matter was present.

LC-MS Instrumentation

UHPLC-MS/MS

UHPLC analysis was carried out using an Acquity UPLC system (Waters, Milford, MS, USA), equipped with a binary solvent. The chromatographic separation was achieved using an Acquity UPLC HSS T3 column, 1.8 μm , 100 mm x 2.1 mm I.D (Waters) at a flow rate of 0.3 mL/min. The mobile phase consisted of water/methanol gradient (both 0.1 mM NH_4Ac). The LC system was interfaced to a TQDTM (quadrupole T-wave quadrupole) mass spectrometer with an orthogonal electrospray ionization source Z-spray (Waters). For operation in the MS/MS mode, collision gas was Argon 99.995% (Carbueros Metálicos, Valencia, Spain) with a pressure of 2×10^{-3} mbar in the T-wave cell. The additional experimental setups can be found elsewhere [22].

UHPLC-QTOF MS

An ultra performance Acquity liquid chromatography (UPLCTM) system (Waters) was interfaced to a QTOF mass spectrometer (QTOF Premier, Waters) using an orthogonal Z-spray electrospray interface. LC separation was performed using an Acquity UPLC

HSS T3 column, 1.8 μm , 100 mm x 2.1 mm I.D at a flow rate of 300 $\mu\text{L}/\text{min}$. The mobile phase consisted of water/methanol gradient (both 0.1 mM NH_4Ac) where the methanol percentage was changed linearly as follows: 0 min, 5%; 7 min, 90%; 8 min, 90%; 8.1 min, 5%. The injection volume was 20 μL . TOF-MS resolution was $\sim 10,000$ FWHM (V-mode) at m/z 556. The MCP detector potential was set to 1750 V in positive ionization mode. A cone voltage of 25 V and a capillary voltage of 3 kV were used. The interface temperature was set to 350 $^\circ\text{C}$ and the source temperature to 120 $^\circ\text{C}$. A scan time of 0.05 s was chosen. The automated attenuated function (dynamic range enhancement, DRE) was selected to correct possible mass peak saturations, making it feasible to achieve quantification and accurate mass measurements over a wide concentration range. Calibration experiments from 50 to 1000 m/z were performed monthly using a mixture of NaOH 0.05 M: HCOOH 10% (50:50). A 2 mg/L standard solution of leucine enkephalin was introduced via the lock spray needle (cone, voltage, 90 V) at a flow rate of 30 $\mu\text{L}/\text{min}$.

GC-MS Instrumentation

GC-MS/MS

A GC system (Agilent 6890N, Palo Alto, USA) equipped with an autosampler (Agilent 7683) was coupled to a triple quadrupole (QqQ) mass spectrometer, Quattro Micro GC (Waters), operating in electron ionization (EI) mode. The GC separation was performed using a fused silica HP-5MS capillary column with a length of 30 m x 0.25 mm i.d. and a film thickness of 0.25 μm (J&W Scientific, Folsom, CA, USA). Splitless injections of 1 μL sample were carried out. The system operated in MS/MS (SRM) mode using Argon 99,995% (Carbueros Metálicos) as collision gas at a pressure of 2.8×10^{-3} mbar in the collision cell. More detailed information can be found elsewhere [12].

GC-TOF MS

An Agilent 6890N GC system (Paloalto, CA, USA) equipped with an Agilent 7683 autosampler was coupled to a time-of-flight mass spectrometer, GCT (Waters), operating in EI mode. The GC separation was performed using the same characteristics as in the above GC-MS/MS system. The interface and source temperatures were both set to 250°C and a solvent delay of 3 minutes was selected. TOF MS was operated at 1 spectrum/s acquiring the mass range m/z 50-650 and using a multi-channel plate voltage of 2850V. TOF MS resolution was about 8500 (FWHM) at m/z 612 and heptacosane was used for the daily mass calibration as well as lock mass (m/z ion monitored was 218.9856). The application manager TargetLynx, a module of MassLynx software, was used to process the qualitative and quantitative data obtained from standards and samples for target compounds. The application manager ChromaLynx, also a module of MassLynx software, was used to investigate the presence of non-target compounds in samples. Library searching was performed using the commercial NIST library

Analytical procedure

The UHPLC-MS/MS procedure was based on the previous work developed in our laboratory for the determination of multi-class pesticides in environmental and wastewater samples [22]. Briefly, 100 mL of water sample acidified with HCOOH and containing the surrogate internal standards (IS's) was passed throughout an Oasis HLB cartridge, previously conditioned. After elution with 5 mL acetone, the extract was evaporated and reconstructed with 1 mL acetonitrile:water (10:90, v/v). 20 μ L of the final extract was injected in the UHPLC-MS/MS system. Three SRM transitions were acquired for each compound.

The GC-MS/MS procedure was based on our previous work dealing with the determination of priority of organic contaminants in water [12]. Several of the target compounds are relevant in the field of water policy of the European Union, and in fact are included in Annex X of the Directive 2000/60/EC [5]. Briefly, 100 mL of water sample containing the surrogate IS's was passed throughout a C₁₈ cartridge previously conditioned. After elution with 5 mL ethyl acetate:dichloromethane (50:50), the extract was evaporated and redissolved in 1 mL hexane. 1 µL of the final extract was injected in the GC-MS/MS system. Two SRM transitions were acquired for each compound.

All methods applied were previously validated fulfilling the analytical characteristics required in the field of residue analysis.

RESULTS AND DISCUSSION

GC-MS/MS and LC-MS/MS target analysis

The study presented here was a part of a project, which required the determination of around 100 organic pollutants (see **Table 1**) in treated and raw leachate water samples collected from a MSW treatment plant sited at the Castellón province. The main objective was to investigate the quality of the leachates after treatment with a reverse osmosis process in order to know the feasibility of their dumping into the aquatic environment. Moreover, the analysis of both types of water (treated and non-treated) allowed to evaluate the efficiency of the reverse osmosis process. According to the polarity of target analytes and to our previous experience [13], a modern and efficient analytical strategy, consisting of the combined use of two complementary techniques, GC-MS/MS and UHPLC-MS/MS, was applied. In this way, we were able of widening the scope of the method, covering around 100 target analytes.

This study was carried out between March 2007 to February 2009, and a total of 41 water samples (21 treated and 20 raw leachate) were analyzed. Both methods, GC-MS/MS and UHPLC-MS/MS, were applied for the analysis of all samples collected. The acquisition of, at least, two transitions per compound -one for quantification (Q) and one (or two) additional for confirmation (q_i)- allowed simultaneous quantification and reliable identification of positive findings. Thus, all findings were confirmed by the compliance of both retention time and Q/q_i ratio when compared with a reference standard. Maximum Q/q_i deviations accepted were based on the European Commission Decision [30].

Data obtained from the analysis of samples (**Figure 1**) showed that pesticides were by far the most commonly detected compounds in both treated and raw leachate samples, specially herbicides (phenylurea, triazine, uracil and carbamate), fungicides

(benzimidazole, conazole and anilide) and insecticides (carbamate, organophosphorus (OP) and phenylthiadiazinone). Other contaminants also frequently found were octyl nonyl phenols (ONP) and polycyclic aromatic hydrocarbons (PAH), detected in almost all raw, and in 86 and 57%, respectively, of treated water samples. Among pesticides, phenylurea herbicides exceeded the concentration level of 0.1 µg/L in more than 50% of treated water samples analyzed, followed by triazine herbicides (around 40%) and benzimidazole fungicides (around 30%).

Table 2 shows the detection frequency for the specific organic pollutants detected. It can be seen that most of the positive findings in raw leachate exceeded the 0.1 µg/L level. However, in treated water only ten contaminants surpassed this value in one or more occasions (six herbicides: terbutometon, terbutryn, terbuthylazine, terbacil, simazine and diuron; two fungicides: thiabendazole and imazalil; one insecticide: carbaryl; one PAH: naphthalene). Among them, the phenylurea herbicide diuron was present at concentrations higher than 0.1 µg/L in more than 50% of treated water samples analyzed (57% of samples), due to its wide use in the Castellón province.

Summarizing data obtained in treated water, among 349 positive findings in the two-years of monitoring, only 34 corresponded to concentrations higher than 0.1µg/L, although rarely exceeded 0.5 µg/L. The only exceptions were carbaryl and diuron, with maximum concentrations of 1.5 µg/L (sample of January 2008) and 0.61 µg/L (sample of February 2008), respectively. In the case of 4-t-octylphenol, chlorphenvinphos, chlorpyrifos, diuron and simazine, priority substances in the Environmental Quality Standards (EQS) in the field of water [6], all of them were detected in some occasion in treated water but never exceeding the admissible maximum concentration referenced (0.1, 0.3, 0.1, 1.8 and 4 µg/L, respectively).

As expected, the percentage of detections and concentration levels was notably higher in raw leachates in comparison to treated samples. Thus, among 477 positive findings in raw leachate, 373 exceeded 0.1 µg/L. Several compounds were detected in all samples analyzed. Within the group of pesticides, the herbicides diuron, simazine, terbacil and terbutryn were detected in all the 20 water samples analyzed, reaching concentration levels as high as 21 µg/L (terbacil, August 2008). Four insecticides (carbaryl, carbofuran, dimethoate and pirimicarb) were also found in all non-treated samples. The highest concentration level was reported for dimethoate in the sample of October 2008 (82 µg/L). In the case of fungicides, imazalil and triadimenol were detected in all samples as well, reaching maximum concentrations of 2.3 µg/L in both cases. Other compound, 4-t-octylphenol, used as precursor in the manufacture of non-ionic surfactants, was also found in all the samples analyzed, with a maximum concentration level of 5.6 µg/L.

As illustrative examples, **Figures 2** and **3** show UHPLC-MS/MS and GC-MS/MS chromatograms for treated and raw leachate water samples collected on March 2008.

From all the results obtained, it seems that the treatment process applied (reverse osmosis) in the MSW treatment plant was rather efficient, as it notably reduced the concentration levels of organic contaminants found in raw leachate.

Analysis of water samples by TOF

As illustrated in the previous section, the combined use of GC-MS/MS and LC-MS/MS with triple quadrupole analyzers was a satisfactory approach for quantitative determination of around 100 selected contaminants in treated and raw leachate water samples. However, the list of target analytes was limited to a number of contaminants that, although relevant from environmental point of view, are not the only ones present in the samples. So, to have more realistic information about the pollution degree of

these samples, the investigation of other non-selected contaminants would be necessary. TOF MS was chosen to this aim due to its great potential for wide-scope screening, as stated in the introduction section. Consequently, all water samples were also analyzed by GC-TOF MS and by LC-QTOF MS in order to investigate the presence of other contaminants not included in the list of target analytes. The sample treatment was the same as used for GC-MS/MS (GC-TOF MS analysis) and for UHPLC-MS/MS (UHPLC-QTOF MS analysis). The objective was to identify other pollutants present in the samples that could be added to the list of target analytes in future monitoring programs.

Analysis by GC-TOF MS

The use of GC-TOF MS allowed us to investigate other selected compounds thanks to the full spectrum acquisition at satisfactory sensitivity. In addition, the elucidation of several unknown compounds (non-target analytes) was tested. The methodological approach previously developed for screening and confirmation of organic micropollutants in water [25, 26] was applied in this project for searching target and non-target contaminants in wastewater samples.

The investigation of other selected compounds was carried out in a post-target way, as searching of the compound was performed after MS acquisition [23]. Up to 5 narrow-window eXtracted Ion Chromatograms (nw-XIC), with a mass window of 0.02 Da, at selected m/z ions were obtained for every compound. The application manager TargetLynx was employed to automatically process data and to confirm the identity of compounds detected in samples. Analyte confirmation was performed by comparing the experimental Q/q intensity ratios in samples with the theoretical ones, calculated from standards in solvent. The presence of at least two ions measured at their accurate mass and the compliance of their Q/q ratio within specified tolerances [30] was required for a

reliable confirmation. In the present work, a list of around 150 compounds (see **Supplement Table 1**), where many target analytes investigated by GC-MS/MS QqQ were also included, was investigated in treated and non-treated water samples. Calibration curves were included in every sequence of analysis; so, a semi-quantitative estimation of positive findings could be performed. **Table 3** shows four pesticides that were detected in the samples analyzed. These compounds were not included in the target list of neither the GC-MS/MS nor LC-MS/MS method. Three of them were OP insecticides (diazinon, dichlorvos and fenthion) and were detected in several raw leachates, reaching concentration levels as high as 79 µg/L (fenthion, sample of October 2008). The fungicide diphenylamide was detected in only one raw sample (June 2007), although at high concentration (152 µg/L). In treated water, only diazinon was detected (6 out of 21 samples analyzed), always at concentrations higher than 0.1 µg/L.

Investigation of non-target compounds in the samples was carried out by applying the ChromaLynx Application Manager. This software automatically detected peaks with a response over user-defined parameters, displayed their deconvoluted mass spectra to be searched in the library, and produced a hit list with positive matches (library match > 700 was used as criterion). The formulas from the library hit were submitted to the elemental composition calculator and the five most intense ions were scored by exact mass measurement for the confirmation/rejection of finding [25]. Using this approach, several contaminants were discovered. These compounds were not included in the target list of QqQ-based methods nor the list of post-target GC-TOF MS. **Table 3** shows the non-target compounds detected using this approach. Some of these compounds had been already detected by our group in environmental and biological samples using GC-TOF MS [26, 31]. N-butyl benzenesulfonamide (N-BBSA) used in polyamide and copolyamide plastics and in the manufacturing of sulfonyl carbamate herbicides was the

compound more frequently detected (100% treated and 90% non-treated water). Diethyltoluamide, an insect repellent, was found in all raw water samples and in 8 treated water samples. Benzophenone, a UV filter used primarily as photoinitiator, fragrance enhancer, and also used in the manufacture of insecticides, agricultural chemicals and pharmaceuticals, was identified in around 50% of both treated and non-treated water. Other compounds frequently detected in non-treated waters were caffeine and the pharmaceuticals ibuprofen and benzenesulfonamide.

As an illustrative example, **Figure 4** shows a positive finding of ibuprofen in non-treated water using the GC-TOF MS non-target approach. Accurate mass confirmation automatically performed by the software for four representative ions led to the confirmation of the identity of ibuprofen with mass errors below 1 mDa for three of them.

Analysis by UHPLC-QTOF MS

The analysis of samples by UHPLC-QTOF MS was carried out in a post-target way searching for around 500 compounds that were included in a home-made database. The list contained 377 pesticides and 40 transformation products (TP), as well as 47 antibiotics, 20 pharmaceuticals, and other emerging contaminants reported to have been detected in aquatic environment, such as cocaine or caffeine [27]. For investigation of these compounds, ChromaLynx XS software was used.

Briefly, this application manager automatically processes data, and obtains nw-XICs (mass window of 0.02 Da) at selected m/z ions, usually those corresponding to protonated or deprotonated molecules, based on a selected list of accurate masses and retention times, if available. Besides, this software allows visualizing the complete spectrum of positive findings at accurate masses, which can be compared with a library, if available. This facilitates a rapid and simple review by cataloguing pollutants on

colors, as a function of mass errors. In our case, a theoretical home-made library was built without the need of injecting reference standards. It showed the theoretical spectrum with information on molecular ion mass (typically $(M+H)^+$ in ESI positive) and the theoretical isotopic pattern.

Following this methodology, most of analytes detected by triple quadrupole were also confirmed by TOF MS. In addition, other contaminants not included in the target list of the QqQ methods were identified. **Table 4** shows compounds not investigated by QqQ that were discovered in several samples analyzed. Antibiotics, such as erythromycin and clarythromycin, were detected by UHPLC-QTOF MS in around 5% of treated water samples and around 50% of raw leachates. The analgesic paracetamol was found in more than 70% of samples analyzed. Atenolol, a beta-antagonist used primarily in cardiovascular diseases in the treatment of hypertension, was detected in 85% of samples. Metamizole, an anti-inflammatory drug commonly used as powerful analgesic and antipyretic, was found in 75% of raw samples and in 50% of treated water. Caffeine and the insecticide diazinon, which were already identified in some samples by using GC-TOF MS, were also found in around 40% of raw leachate and a few treated water samples. Paraxanthine, a caffeine metabolite, was detected in both raw and treated water (around 30% samples). Cocaine was detected in only one sample out of the 20 raw leachates analyzed. However, benzoylecgonine, one of the main metabolites of cocaine, was detected in 95% of raw leachate and 60% of treated water samples. Finally, some pesticide TPs, concretely deethyl-terbumeton, 2-hydroxy-terbuthylazine and deethyl-2-hydroxy-terbuthylazine, were also found in several samples.

As an example, **Figure 5** shows a positive finding of diazinon in raw leachate using the UHPLC-QTOF MS post-target approach. The nw-XIC at m/z corresponding to the exact mass of diazinon $[M+H]^+$ is shown together with its accurate mass spectrum and the

theoretical one. Accurate mass confirmation automatically performed for the $[M+H]^+$ ion showed a mass error of 0.7 mDa.

In addition, the availability of a QTOF instrument made it feasible to perform MS/MS experiments to go further in the identification process of the compounds detected, thanks to the useful information given by product ion spectra at accurate mass. Full-acquisition accurate mass data were also processed in a non-target way [27], trying to elucidate “unknown” components detected in samples. The difference with respect to the GC-TOF MS approach was the use of only two ions (softer ionization in ESI in comparison to EI) and the use of the theoretical home-made library previously described, due to the non-availability of extensive and reproducible LC-MS commercial libraries. After processing MS data in the non-target approach, only five contaminants were discovered: paracetamol, cocaine, benzoilecgonine, caffeine and diazinon. All of them had been previously detected in the post-target screening, and corresponded to those analytes with sensitive response in TOF MS. The rest of compounds (see **Table 4**) could not be elucidated using this non-target approach due to either their low sensitivity and/or their low concentration level in the samples. It is important to remark that the LC-TOF MS screening of organic contaminants using a non-target approach may not be fully satisfactory at the moment, as the success of this approach gets notably worse at low analyte responses, being therefore less efficient than the post-target analysis. The non-availability of wide commercial reproducible libraries, as in GC with EI, is an important limitation at present. In fact, using non-target analysis only five compounds were elucidated, as stated above.

As an example of the non-target UHPLC-QTOF MS approach, **Figure 6** shows a positive finding of paracetamol in non-treated leachate. Accurate mass confirmation

automatically performed for two representative ions led to the confirmation of the identity of paracetamol in this sample with mass errors of 0 and -0.7 mDa.

CONCLUSIONS

Investigation of organic pollutants of wide polarity ranges in water requires the use of two complementary techniques: GC-MS, for the determination of non-polar/semi volatile analytes, and LC-MS, for more polar analytes. In this paper, an analytical strategy consisting on the combined use of GC-MS/MS and UHPLC-MS/MS, both with triple quadrupole analyzers, has been applied in order to investigate the presence of around 100 organic contaminants in treated and raw leachate samples from a Municipal Solid Waste treatment plant.

Pesticides have been the most commonly detected compounds in both type of samples, especially herbicides (phenylurea, triazine, uracil and carbamate), fungicides (benzimidazole, conazole and anilide) and insecticides (carbamate, organophosphorus and phenylthiadiazinone). Other contaminants widely detected were 4-t-octylphenol and several PAHs such as naphthalene, phenanthrene and pyrene.

As expected, the frequency of detections and pollutant concentrations in raw leachates was notably higher than in treated samples. Most of positive findings in non-treated samples were at concentrations above 0.1 $\mu\text{g/L}$. However, in treated water very few compounds were detected at levels higher than 0.1 $\mu\text{g/L}$, and rarely exceeded 0.5 $\mu\text{g/L}$. From data obtained, it seems that the reverse osmosis treatment applied in the MSW treatment plant was rather efficient, as it notably reduced the concentration levels of organic contaminants found in raw leachates.

MS/MS techniques using triple quadrupole analyzer have shown a great potential in environmental analysis due to its high sensitivity and selectivity. However, tandem MS methods are developed on purpose for a limited list of target contaminants (around 100 organic contaminants in this work); so, other relevant pollutants that might be present in the samples would be ignored in these analyses. For this reason, all the samples were

also analyzed by GC-TOF MS and UHPLC-QTOF MS in order to investigate the presence of many other contaminants, either in a post-target way (searching for selected pollutants after MS acquisition data) or in a non-target way (searching for unknowns without any previous selection nor information on the compounds to be investigated). This was feasible thanks to the full MS spectra acquisition by TOF analyzers, which offered the possibility of searching for a large number of contaminants with the help of the accurate mass information of the molecules and of the fragment ions. This has allowed discovering several compounds not included in the initial target list of organic contaminants. Other pesticides (diazinon, dichlorvos, diphenylamine and fenthion) and some TPs (deethyl-terbutometon, 2-hydroxy-terbutylazine and deethyl-2-hydroxy-terbutylazine), pharmaceuticals (erythromycin, clarythromycin, atenolol, metamizole, benzenesulfonamide, ibuprofen and paracetamol), drugs of abuse (cocaine and its metabolite benzoylecgonine), the UV filter benzophenone, N-BBSA, the insect repellent diethyltoluamide, or caffeine and its metabolite paraxanthine, are examples of compounds identified in additional analyses performed by TOF instruments. These discovered analytes could be included in the target quantitative methods applied in futures monitoring programs. This paper shows the potential of TOF MS for screening purposes, as this analyzer in combination with GC and LC is able to detect and identify a huge number of pollutants, making of it an excellent analytical tool for wide-scope screening in the environmental field.

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FIGURE CAPTIONS

Figure 1. Frequency of detection (%) of different families of organic contaminants in treated and raw leachate samples collected from the MSW treatment plant between March 2007 and February 2009.

INS: insecticide, FG: fungicide, HB: herbicide, OC: organochlorine, ONP: octyl nonyl phenols, OP: organophosphorus, PAH: polycyclic aromatic hydrocarbons

Figure 2. UHPLC-MS/MS chromatograms for treated and raw leachate samples, both collected on 3rd March 2008. Only the quantification transition (Q) is shown for every analyte (n.d., not detected).

Figure 3. GC-MS/MS chromatograms for treated and raw leachate samples, both collected on 3rd March 2008. Only the quantification transition (Q) is shown for every analyte (n.d., not detected).

Figure 4. Identification of non-target ibuprofen by GC-TOF MS in a raw leachate sample collected on 28th August 2008. (A) Extracted-ion chromatogram for four m/z ions. (B) Commercial library mass spectrum of ibuprofen at nominal mass. (C) Deconvoluted accurate mass spectrum of ibuprofen in the sample (mass errors shown in mDa).

Figure 5. Identification of diazinon by UHPLC-QTOF MS in a raw leachate sample collected on 31th October 2008. (A) Extracted-ion chromatogram for $[M+H]^+$ ion, m/z 305.1089. (B) Theoretical library mass spectrum of diazinon at nominal mass. (C) Accurate mass spectrum of diazinon in the sample.

Figure 6. Identification of non-target paracetamol by UHPLC-QTOF MS in a raw leachate sample collected on 10th October 2008. (A) Extracted-ion chromatogram for two m/z ions. (B) Theoretical library mass spectrum of

paracetamol at nominal mass. (C) Deconvoluted accurate mass spectrum of paracetamol in the sample (mass errors shown in mDa).

SUPPLEMENT

Table 1. List of compounds investigated by GC-TOF MS in a post-target way

4-n-Nonylphenol	Cyfluthrin	Hexythiazox	PCB 180
4-n-Octylphenol	Cyfluthrin_1	Imazalil	PCB 189
4-t-Octylphenol	Cyfluthrin_2	Indeno(1,2,3,cd)pyrene	PCB 28
Acenaphthene	Cyfluthrin_3	Iprodione	PCB 52
Acenaphthylene	Cyfluthrin_4	Isodrin	PCB 77
Alachlor	Cypermethrin_1	Isofenfos	PCB 81
Aldrin	Cypermethrin_2	lambda-Cyhalothrin	Penconazole
Anthracene	Cypermethrin_3	Lindane	Pentachlorobenzene
Atrazine	Cypermethrin_4	Malathion	Permethrin_1
Atrazine desethyl	Cyprodinil	Metalaxyl	Phenanthrene
Atrazine desisopropyl	Deltamethrin	Metamidophos	Phosmet
Azinphos methyl	Diazinon	Methacrifos	Pirimicarb
BDE 100	Dibenzo(a,h)anthracene	Methidathion	Pirimiphos ethyl
BDE 138	Dichlofluanide	Methiocarb	Pirimiphos methyl
BDE 153	Diclorvos	Methiocarb sulfone	Procymidone
BDE 154	Dieldrin	Metolachlor	Profenofos
BDE 183	Diflufenican	Metoxychlor	Propyzamide
BDE 28	Dimethoate	Mevinfos	Pyrazofos
BDE 47	Diphenylamine	Mirex	Pyrene
BDE 66	Endosulfan ether	Molinate	Quinalfos
BDE 71	Endosulfan sulfate	Naphthalene	Simazine
BDE 85	Ethion	Omethoate	tau-Fluvalinate_1
BDE 99	Etrimfos	Oxadixyl	tau-Fluvalinate_2
Benzo(a)anthracene	Fenarimol	<i>p,p'</i> -DDD	Tebuconazole
Benzo(a)pyrene	Fenchlorfos	<i>p,p'</i> -DDE	Tecnazen
Benzo(b)fluoranthene	Fenitrothion	<i>p,p'</i> -DDT	Terbacil
Benzo(g,h,l)perylene	Fenoxycarb	Parathion ethyl	Terbumeton
Benzo(k)fluoranthene	Fenthion	Parathion methyl	Terbumetona desethyl
Bifentrin	Fenvalerate_1	PCB 101	Terbutylazine
Bupimirate	Fluoranthene	PCB 105	Terbutylazine desethyl
Buprofezin	Fluorene	PCB 114	Terbutryn
Carbaryl_1	Fonofos	PCB 118	Tetradifon
Carbaryl_2	Forate	PCB 123	Thiabendazole
Chlorfenvinphos	Fosalone	PCB 126	Trifluraline
Chlorpropham	Fosfamidon	PCB 138	α-Endosulfan
Chlorpyrifos	Heptachlor	PCB 153	β-Endosulfan
Chlozolinate	Heptachlor epoxide A	PCB 156	
Chlopyriphos methyl	Heptachlor epoxide B	PCB 157	
Chrysene	Heptenofos	PCB 167	
Coumafos	Hexachlorobenzene	PCB 169	

Table 1. List of target compounds included in the analyses (method 1: GC-MS/MS; method 2: UHPLC-MS/MS)

Compound	Family	Method	Compound	Family	Method
4-n-Nonylphenol	ONP	1	Heptachlor	INS OC	1
4-n-Octylphenol	ONP	1	Heptachlor epoxide A	Heptachlor TP	1
4-t-Octylphenol	ONP	1	Heptachlor epoxide B	Heptachlor TP	1
Acenaphthene	PAH	1	Imazalil	FG Conazole	2
Acenaphthylene	PAH	1	Imidacloprid	INS Nitroguanidine	2
Acetamiprid	INS Pyridylmethylamine	2	Indeno(1,2,3,cd)Pyrene	PAH	1
Alachlor	HB Chloroacetanilide	1, 2	Isodrin	INS OC	1
Aldrin	INS OC	1	Isoproturon	HB Phenylurea	2
Anthracene	PAH	1	Lindane	INS OC	1
Atrazine	HB Triazine	1, 2	Malathion	INS OP	2
Azinphos-methyl	INS OP	2	MCPA	HB Phenoxyacetic	2
Azoxystrobin	FG Strobilurin	2	Metalaxyl	FG Anilide	2
B(a)Anthracene	PAH	1	Methidation	INS OP	2
B(a)Pyrene	PAH	1	Methiocarb	INS Carbamate	2
B(b)Fluoranthene	PAH	1	Methomyl	INS Carbamate	2
B(g,h,i)Perylene	PAH	1	Methoxychlor	INS OC	1
B(k)Fluoranthene	PAH	1	Metolachlor	HB Chloroacetanilide	1, 2
BDE 100	PBDE	1	Mirex	INS OC	1
BDE 138	PBDE	1	Naphthalene	PAH	1
BDE 153	PBDE	1	<i>p,p'</i> -DDD	DDT TP	1
BDE 154	PBDE	1	<i>p,p'</i> -DDE	DDT TP	1
BDE 28	PBDE	1	<i>p,p'</i> -DDT	INS OC	1
BDE 47	PBDE	1	PCB 101	PCB	1
BDE 66	PBDE	1	PCB 118	PCB	1
BDE 71	PBDE	1	PCB 138	PCB	1
BDE 85	PBDE	1	PCB 153	PCB	1
BDE 99	PBDE	1	PCB 180	PCB	1
Bentazone	HB Carbamate	2	PCB 28	PCB	1
Bromacil	HB Uracil	2	PCB 52	PCB	1
Buprofezin	INS Phenylthiadiazinone	2	Pentachlorobenzene	Chlorobenzene	1
Carbaryl	INS Carbamate	2	Phenanthrene	PAH	1
Carbendazim	FG Benzimidazole	2	Pirimicarb	INS Carbamate	2
Carbofuran	INS Carbamate	2	Pirimiphos-methyl	INS OP	2
Chlorfenvinphos	INS OP	1	Propanil	HB Anilide	2
Chlorpyrifos	INS OP	1	Pyrene	PAH	1
Chrysene	PAH	1	Pyridaphenthion	INS OP	2
Cyprodinil	FG Anilinopyrimidine	2	Simazine	HB Triazine	1, 2
Dibenzo(a,h)Anthracene	PAH	1	Terbacil	HB Uracil	2
Dieldrin	INS OC	1	Terbumeton	HB Triazine	2
Dimethoate	INS OP	2	Terbutylazine	HB Triazine	1, 2
Diuron	HB Phenylurea	2	Terbutryn	HB Triazine	2
Endosulfan ether	Endosulfan TP	1	Thiabendazole	FG Benzimidazole	2
Endosulfan sulphate	Endosulfan TP	1	Thiobencarb	HB Carbamate	2
Fenarimol	FG Pyrimidine	2	Triadimenol	FG Conazole	2
Fluoranthene	PAH	1	Trifluralin	HB Dinitroaniline	1
Fluorene	PAH	1	α -Endosulfan	INS OC	1
HCB	INS OC	1	β -Endosulfan	INS OC	1

FG: fungicide; HB: herbicide; INS: insecticide; OC: organochlorine; ONP: octyl nonyl phenols; OP: organophosphorus; PAH: polycyclic aromatic hydrocarbons; PBDE: polybrominated diphenyl ether; TP: transformation product

Table 2. Results obtained from UHPLC-MS/MS and GC-MS/MS target analysis of water samples from the MSW treatment plant between March 2007–February 2009 (total number of samples: 21 treated and 20 non-treated).

Compound	% positive samples		% samples > 0.1 µg/L		Maximum level (µg/L)	
	Non-treated	Treated	Non-treated	Treated	Non-treated	Treated
Acetamiprid	0	9.5	0	0	nd	<0.025
Atrazine	5	5	0	0	<0.025	<0.025
Azinphos-methyl	15	0	10	0	4.3	nd
Azoxystrobin	30	29	30	0	0.41	<0.025
B(a)Pyrene	0	5	0	0	nd	<0.025
Bromacil	75	43	50	0	14	0.03
Buprofezin	65	57	35	0	1.0	<0.025
Carbaryl	100	48	90	14	40	1.5
Carbendazim	95	81	75	0	41	0.08
Carbofuran	100	71	95	0	43	0.10
Chlorfenvinphos	75	57	75	0	3.6	0.082
Chlorpyrifos	55	38	50	0	7.5	<0.025
Dieldrin	5	0	5	0	1.3	nd
Dimethoate	100	48	85	0	82	0.10
Diuron	100	95	100	57	19	0.61
Fluorene	5	0	0	0	<0.025	nd
Imazalil	100	48	65	5	2.3	0.31
Imidacloprid	5	5	0	0	<0.025	<0.025
Isoproturon	5	5	0	0	<0.025	<0.025
Lindane	5	0	0	0	<0.025	nd
Malathion	75	24	70	0	64	0.04
MCPA	55	29	50	0	10	0.03
Metalaxyl	95	62	40	0	4.4	<0.025
Methidation	40	5	15	0	13	<0.025
Methiocarb	55	9.5	30	0	1.6	0.030
Naphthalene	55	48	55	14	15	0.31
4-t-Octylphenol	100	86	85	0	5.6	0.044
Phenanthrene	85	57	60	0	1.3	<0.025
Pirimicarb	100	71	50	0	13	<0.025
Pirimiphos-methyl	15	0	5	0	0.13	nd
Pyrene	60	43	50	0	0.42	<0.025
Simazine	100	81	65	9.5	17	0.23
Terbacil	100	81	100	5	21	0.14
Terbumeton	85	67	75	5	29	0.16
Terbuthylazine	95	81	95	14	40	0.48
Terbutryn	100	71	95	9.5	14	0.20
Thiabendazole	95	90	85	29	14	0.37
Thiobencarb	35	43	15	0	1.4	<0.025
Triadimenol	100	71	60	0	2.3	0.06

nd: not detected

Table 3. Results obtained from GC-TOF MS analysis of water samples from the MSW treatment plant between March 2007 – February 2009 (total number of samples: 21 treated and 20 non-treated).

Compound	% positive samples		Maximum level (µg/L)	
	Treated	Non-treated	Treated	Non-treated
<i>Post-Target approach</i>				
Diazinon	28	55	7.7	38
Dichlorvos	0	5	nd	11
Diphenylamine	0	5	nd	152
Fenthion	0	5	nd	79
<i>Non-target approach</i>				
Benzenesulfonilide	5	70		
Benzoguanamine	0	5		
Benzophenone	38	55		
BHT	10	5		
BHT-CHO	5	10		
n-Butylbenzenesulfonamide	100	90		
Caffeine	5	35		
Cotinine	5	0		
3,4-Dichloroaniline	5	10		
Diethyltoluamide	38	100		
8-Hydroxyquinoline	5	0		
Ibuprofen	10	30		
Ibuprofen, trimethylsilylester	5	0		
Methylparaben	5	20		
Nicotine	0	5		
Triacetin	0	5		
3,4,5-Trichlorobenzeneamine	0	5		

nd: not detected

Table 4. Results obtained from UHPLC-QTOF MS analysis of water samples from MSW treatment plant between March 2007–February 2009 (total number of samples: 21 treated and 20 non-treated).

Compound	% positive samples	
	Treated	Non-treated
<i>Post-target approach</i>		
Atenolol	85	85
Benzoylecgonine*	60	95
Caffeine*	40	43
Clarithromycin	5	45
Cocaine*	0	5
Diazinon*	5	42
Erythromycin	5	65
Metamizole*	50	75
Paracetamol	70	84
Paraxanthine	25	35
deethyl-Terbutometon	10	15
deethyl-2-hydroxy-Terbuthylazine	35	55
2-hydroxy-Terbuthylazine	50	70

*Compounds that were also discovered performing the non-target approach

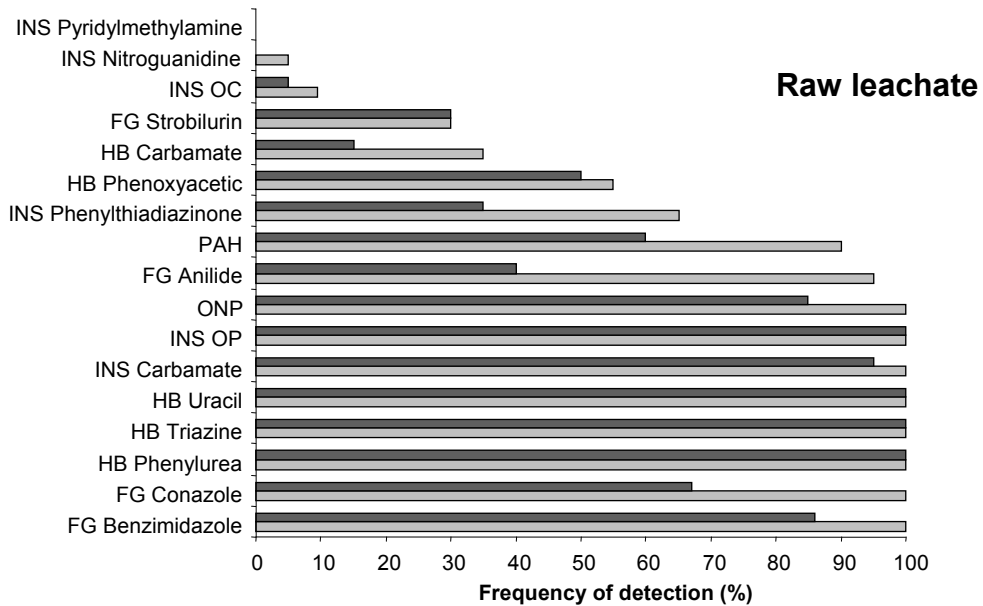
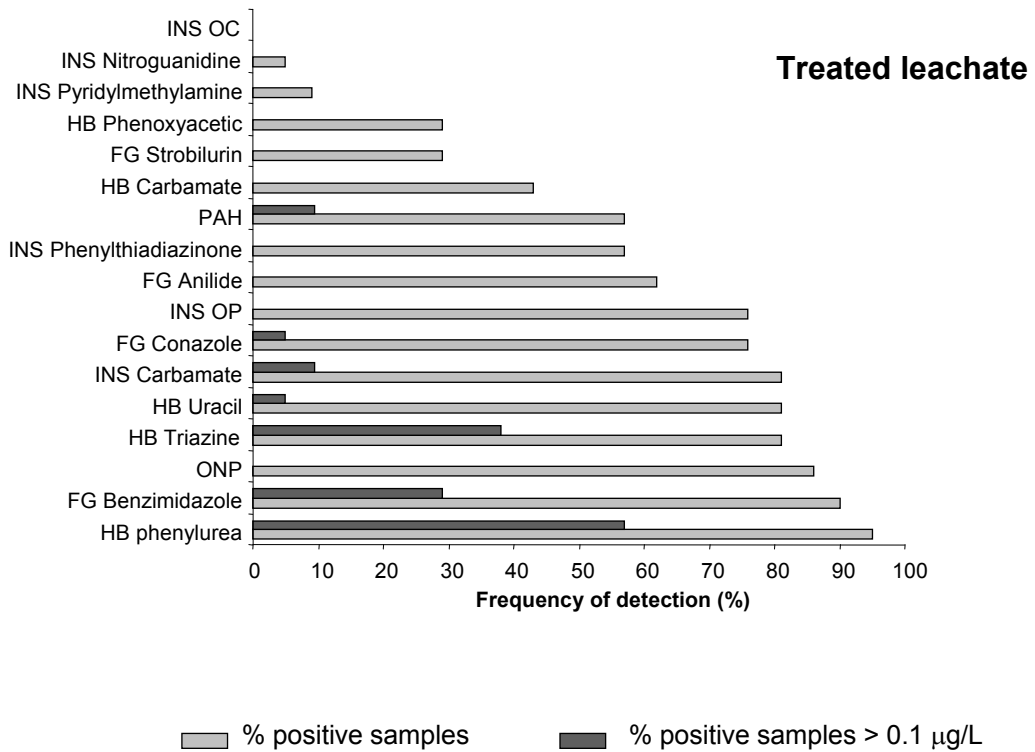


Figure 1

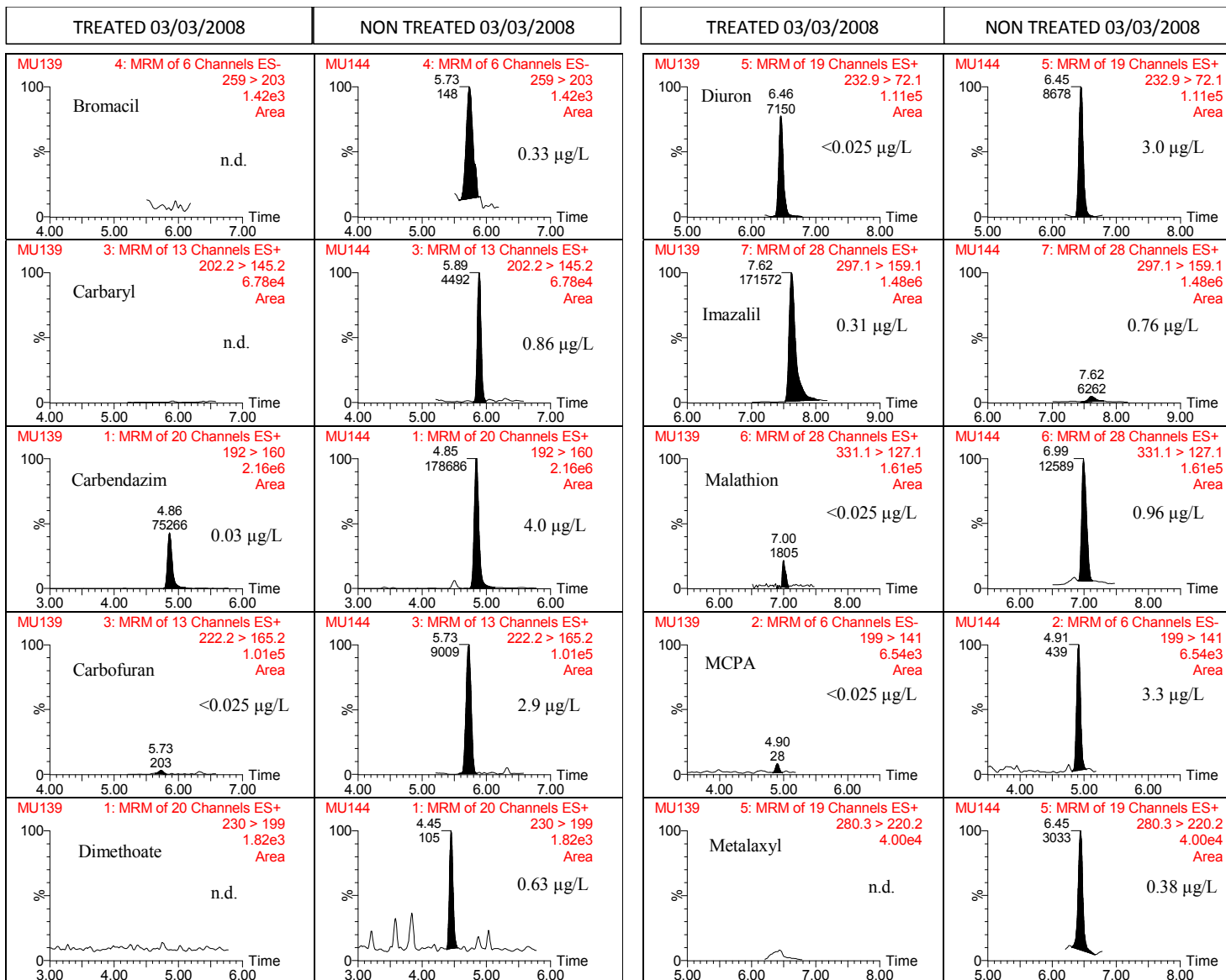


Figure 2

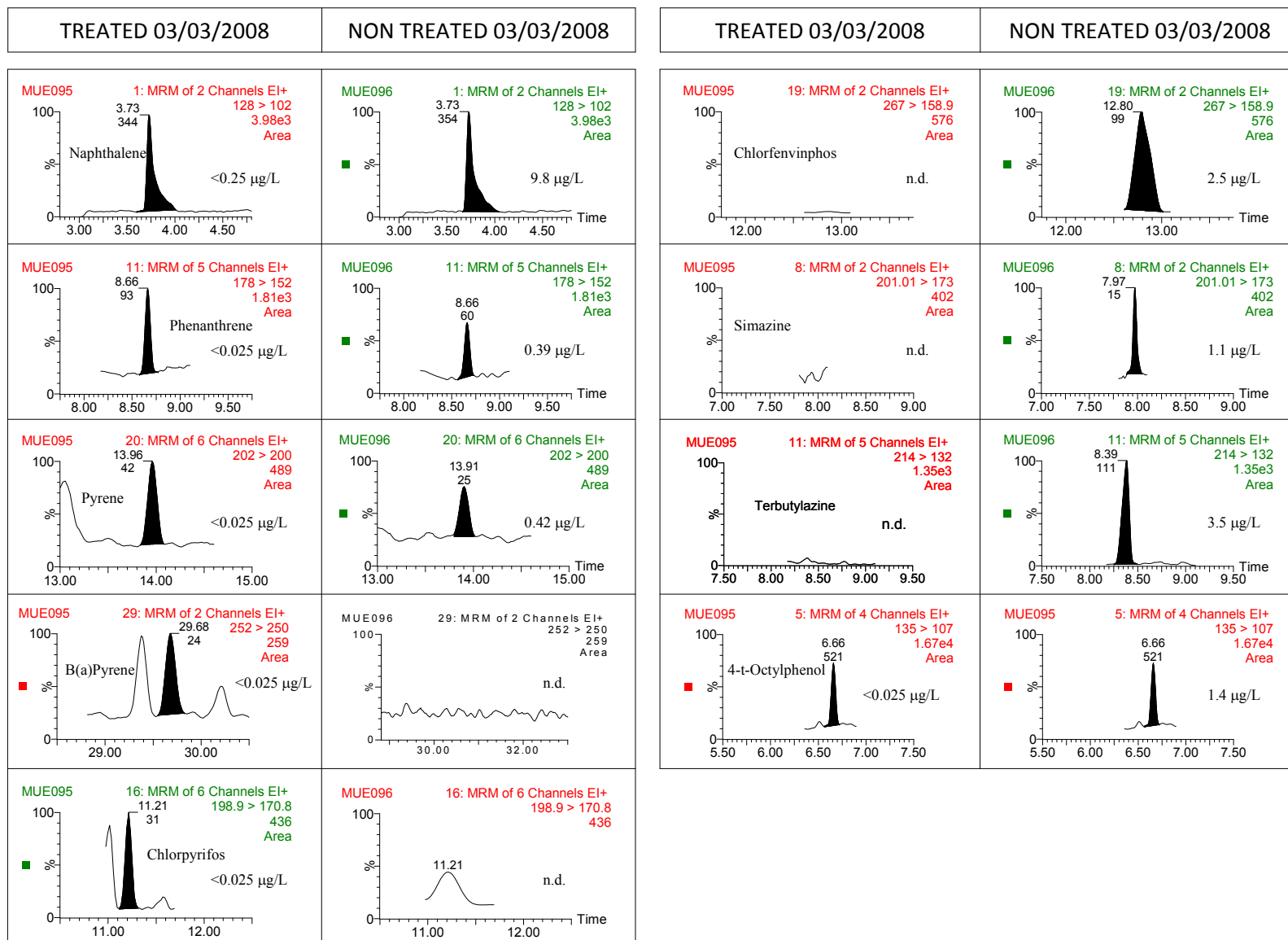
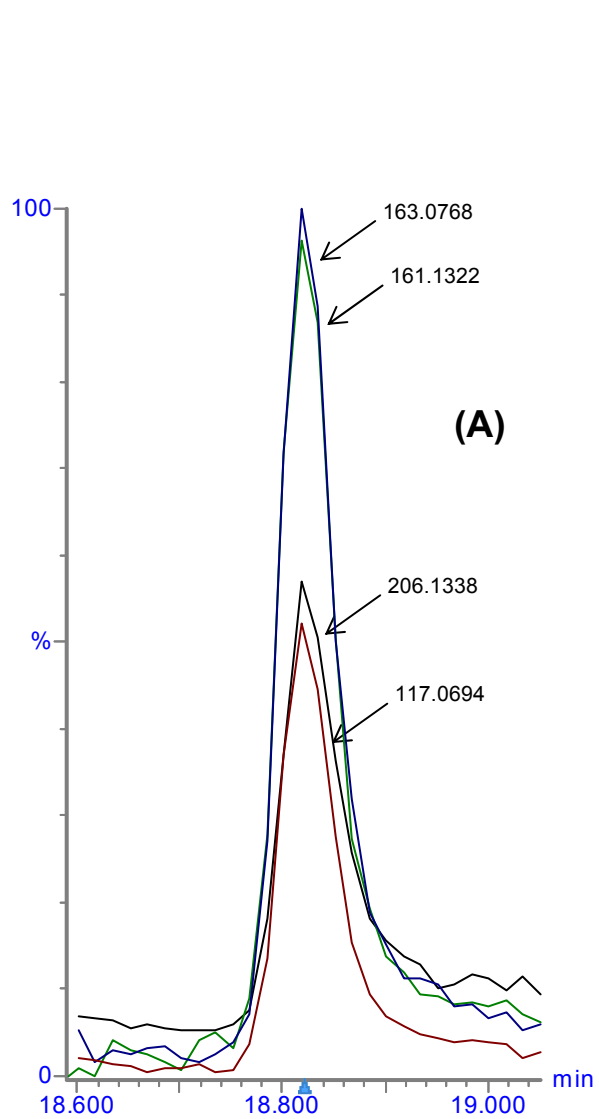
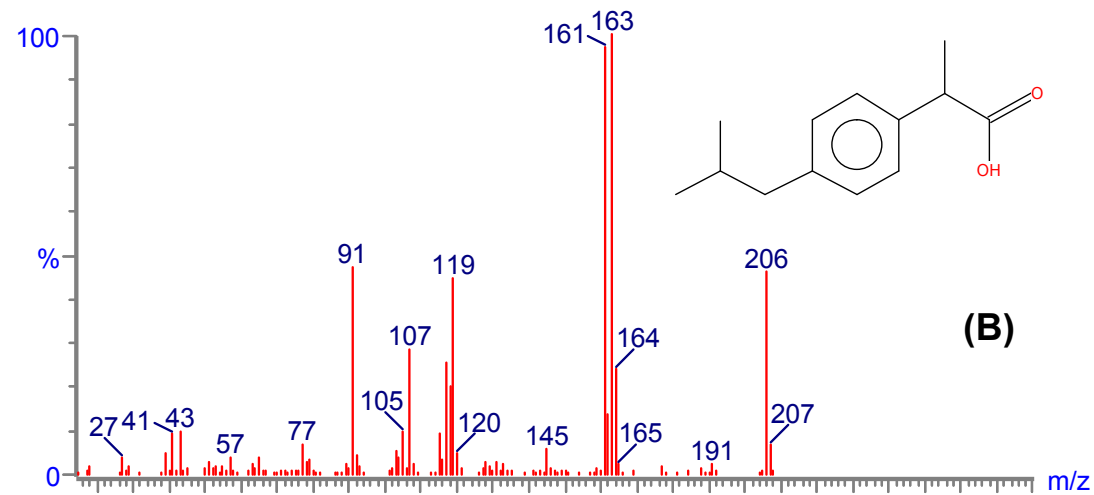


Figure 3



Library: Ibuprofen



Suggested compound: Ibuprofen

2.702e+003

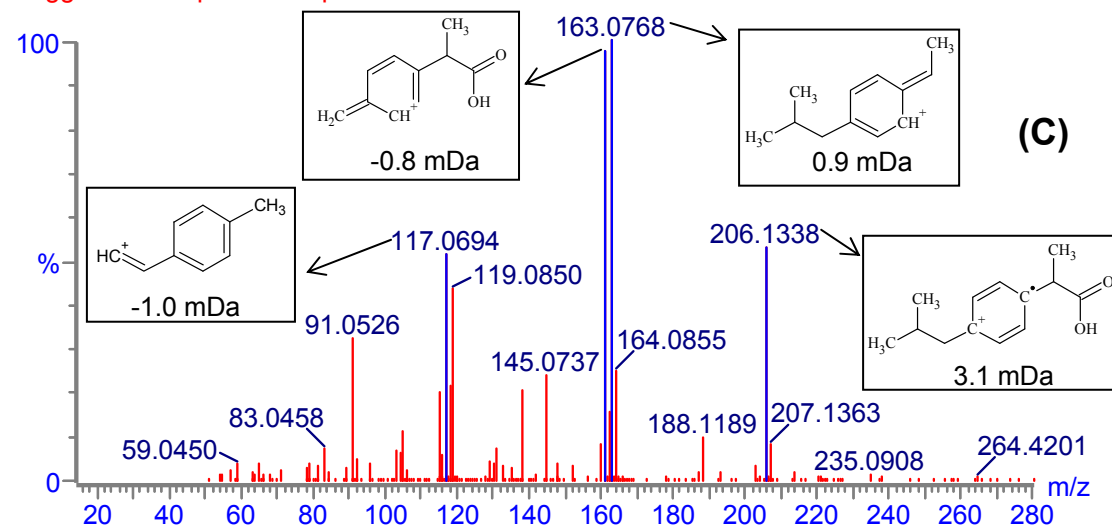
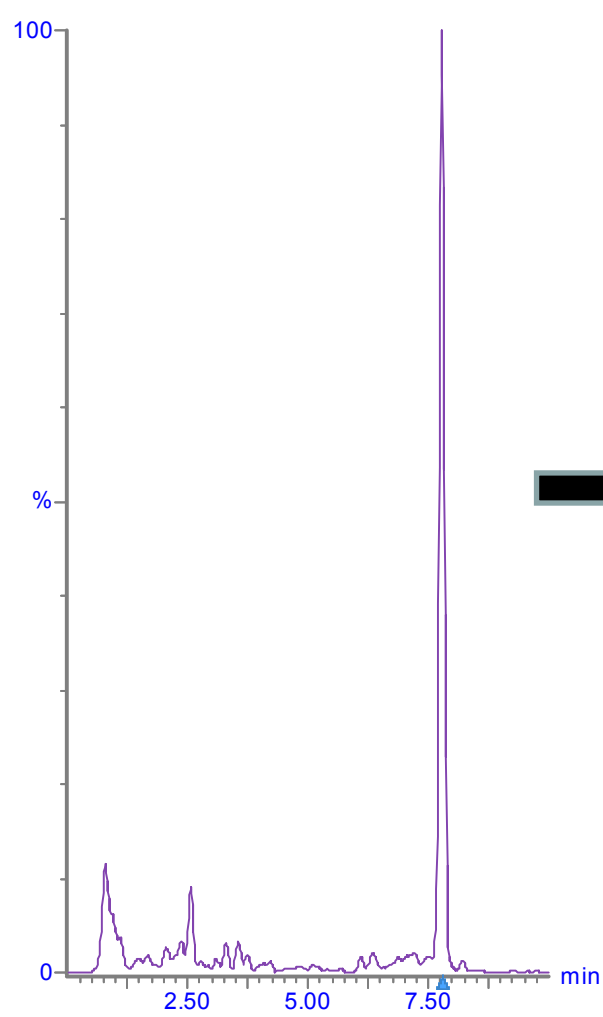
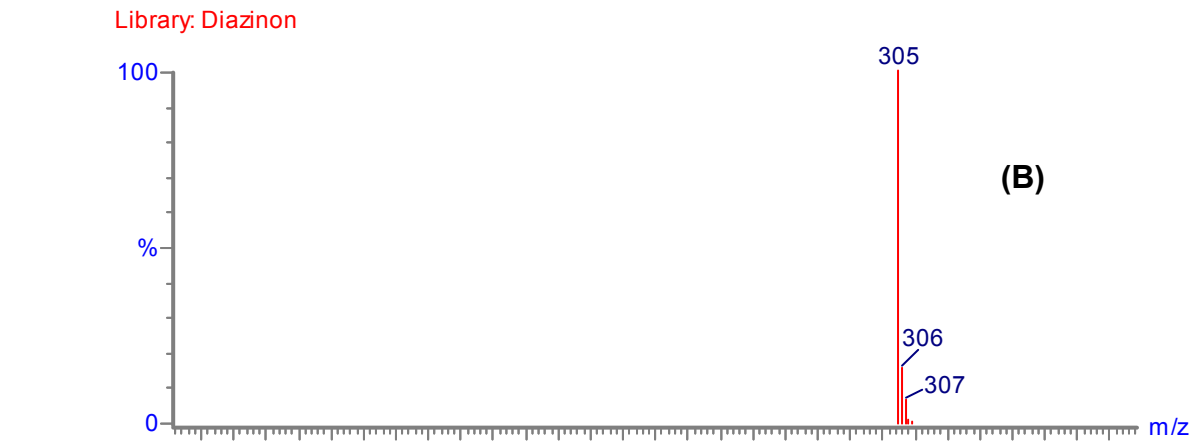


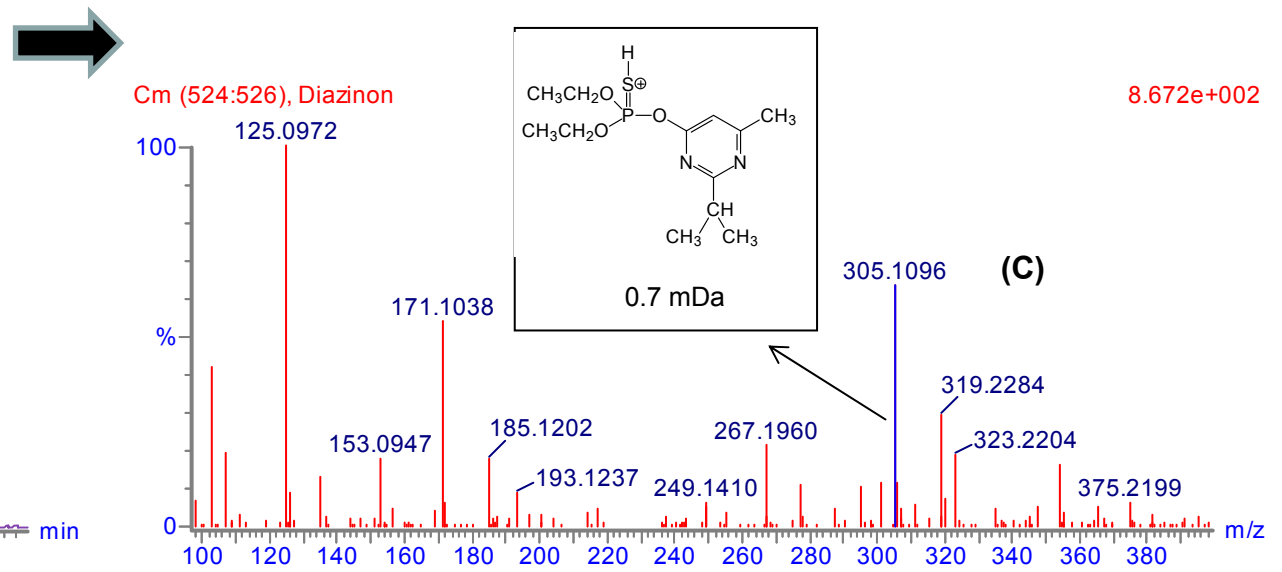
Figure 4



(A)

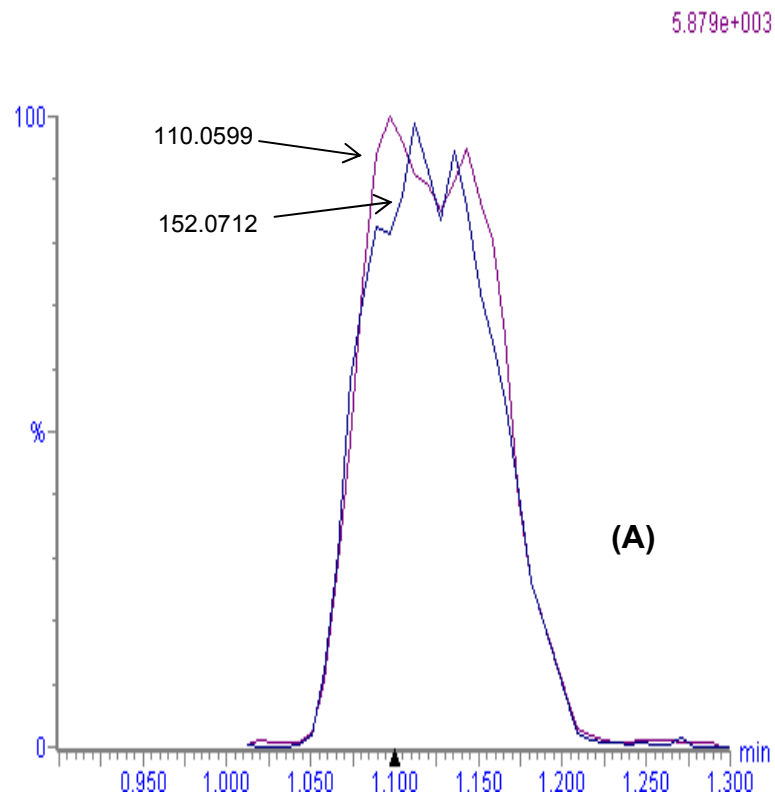


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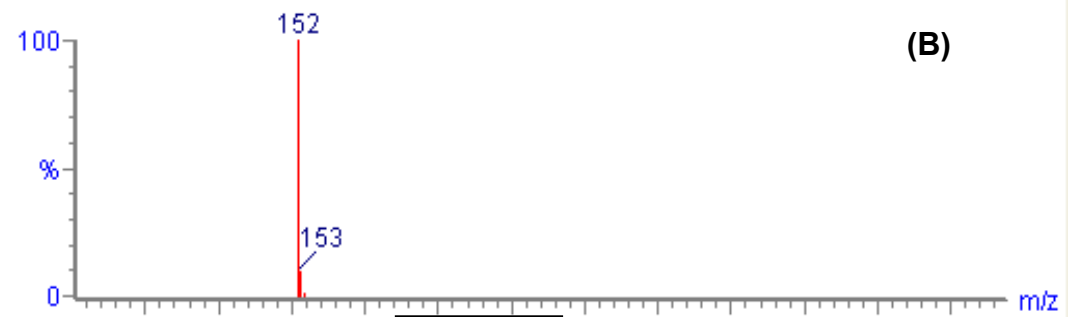


(C)

Figure 5



Library: Paracetamol



Component: Paracetamol

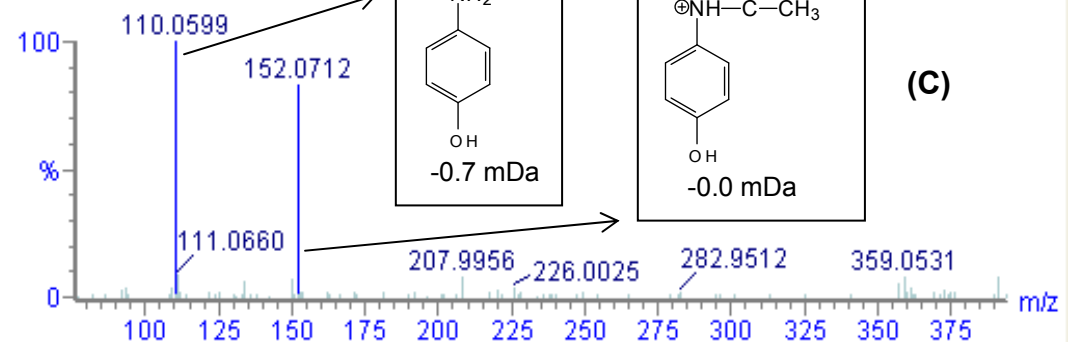


Figure 6