

1 **SUPPLEMENTARY MATERIAL**

2 **Study of the electrochemical oxidation of 4,6-**
3 **dimethyldibenzothiophene on a BDD electrode employing**
4 **different techniques**

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17 Text S1

18 *HPLC-UV analysis*

19 The electrolyzed solutions were collected at different electrolysis times, and the HPLC-
20 UV analysis to monitor the abatement of 4,6-DMDBT concentration during its oxidation
21 upon bulk electrolysis was performed using a 1260 Infinity Series Instrument LC-DAD
22 chromatograph coupled to a 1260 Infinity Multiple Detector (MWD) from Agilent
23 Technologies, (Santa Clara, CA). A Kinetex Reversed Phase C18 100 Å (30 mm × 2.1 mm)
24 column at 28 °C was utilized with a gradient elution at 0.5 mL min⁻¹ programmed as follows:
25 acetonitrile-water (10% A-90% B) for 0-14 min, 90% B at 14-20 min and (90% A-10% B)
26 at 20-25 min. Both solvents contained 0.1% formic acid (reagent grade purchased from
27 Sigma-Aldrich). The injection volume was 5 µL and the detection wavelengths were 208,
28 232, 244 and 254 nm.

29 Text S2

30 *GC-MS analysis*

31 After evaporation, the solid phase was dissolved in methanol to analyze the oxidized
32 products using a Varian CP-3800 gas chromatograph coupled to a mass spectrometry detector
33 (Saturn 4000, Varian). The compounds were separated on an HP-5 (30 m × 0.25 mm, 0.25
34 μm film thickness) capillary column, using helium as carrier gas at a constant flow rate of 1
35 mL min⁻¹. The temperature program was: 70 °C for 2 min, up to 150 °C at 30 °C min⁻¹ and
36 then, up to 250 °C at 5 °C min⁻¹, with a final isothermal stage for 25.33 min. A 100 μL sample
37 was injected in splitless mode (injection port temperature of 220 °C) using a Varian 8400
38 autosampler equipped with a 10 μL syringe. Ion trap MS determination was carried out in
39 full scan mode (*m/z* scan range of 40-600 Da) using EI ionization at 70 eV in positive mode
40 and external ionization configuration. GC-MS interface, ion trap and manifold temperatures
41 were 275, 190 and 60 °C, respectively.

42 Text S3

43 *UHPLC-ESI-Q-TOF-MS analysis*

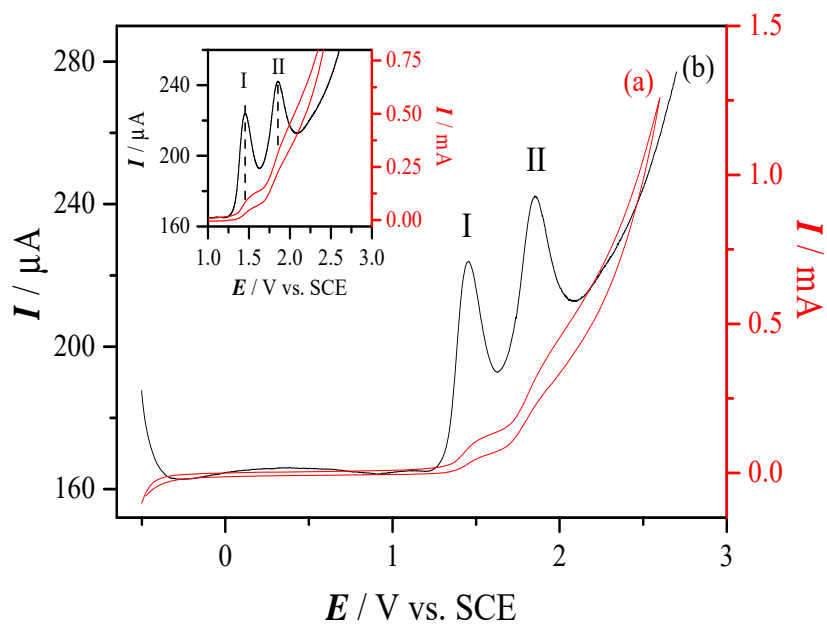
44 The electrolyzed solutions were concentrated by evaporation. Then, they were diluted
45 (1:100) for the analysis of the products by UHPLC-ESI-Q-TOF-MS. The modern Q-TOF-
46 MS instruments allow the simultaneous acquisition of two full spectra with different collision
47 energies in a single injection (MSE mode). The use of the low energy (LE) function with a
48 collision energy of 4 eV informs about non-fragmented ions related to the parent protonated
49 molecule $[M+H]^+$ in positive ionization mode. The high energy (HE) function, with a
50 collision energy ramp ranging from 15 to 40 eV, provides a wide range of fragmented ions.

51 A Waters Acquity ultra-performance liquid chromatography (UPLC) system (Waters,
52 Milford) was employed. The separation was performed using an Acquity UPLC BEH C18
53 (2.1 mm \times 100 mm, 1.7 μ m particle size) column from Waters. The mobile phases contained
54 water (A) and/or methanol (B), both with 0.01% formic acid. The percentage of B changed
55 as follows: 10% at 0 min, 90% at 14 min, and 10% again at 16.10 min. The flow rate was
56 300 μ L min^{-1} and the analysis run time was 18 min. The sample injection volume was 20 μ L.
57 The UPLC system was interfaced to a hybrid quadrupole-TOF high resolution mass
58 spectrometer (HRMS) (Xevo G2 Q-TOF, Waters Micromass), using an orthogonal Z-spray-
59 ESI interface operating in both positive and negative ion mode. TOF-MS resolution was
60 approximately 25000 at full width-half maximum at m/z 556. Nitrogen was used as drying
61 and nebulizing gas at a flow rate of 800 $\text{L}\cdot\text{h}^{-1}$. The MS data were acquired over an m/z range
62 of 50-1200 at a scan time of 0.4 s. A capillary voltage of 0.7 kV and cone voltage of 20 V
63 were used in positive ionization mode. The collision gas was argon (99.995%, Praxair). The
64 interface, source and column temperatures were 450, 150 and 40 $^{\circ}\text{C}$, respectively.

65 Text S4

66 *ICP-MS analysis*

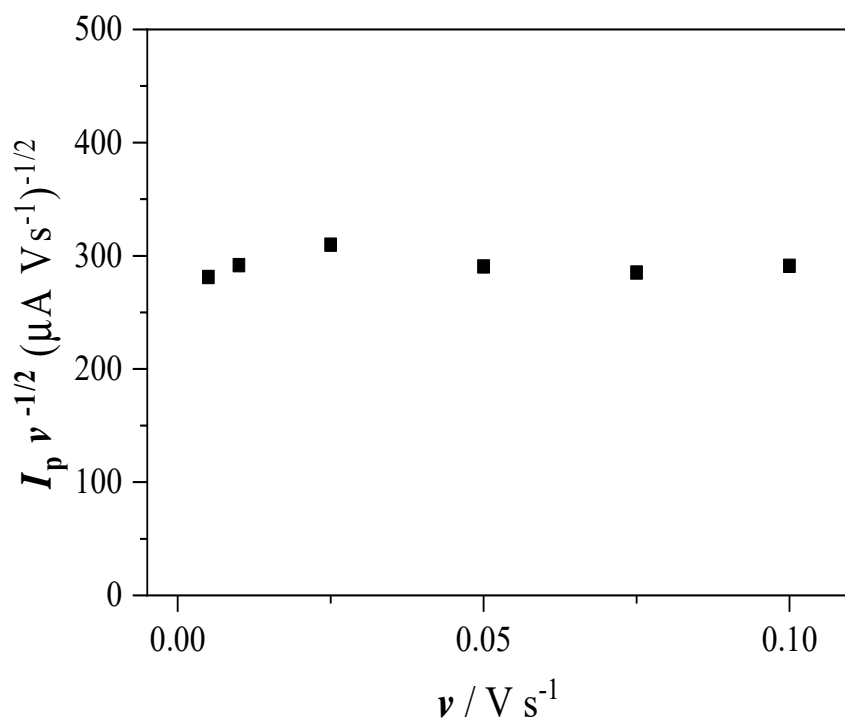
67 Qualitative determination of niobium in electrolyzed solutions was conducted with an
68 Agilent 7500cx ICP-MS instrument. For this purpose, the selected samples were evaporated
69 to dryness and redissolved with 2 mL of 1% HNO₃. Afterwards, the resulting samples were
70 nebulized to the ICP-MS and the signal at m/z 93 was monitored in qualitative mode (20
71 points per peak). Reagents free of Nb were checked from the analysis of blank solutions with
72 1% HNO₃.



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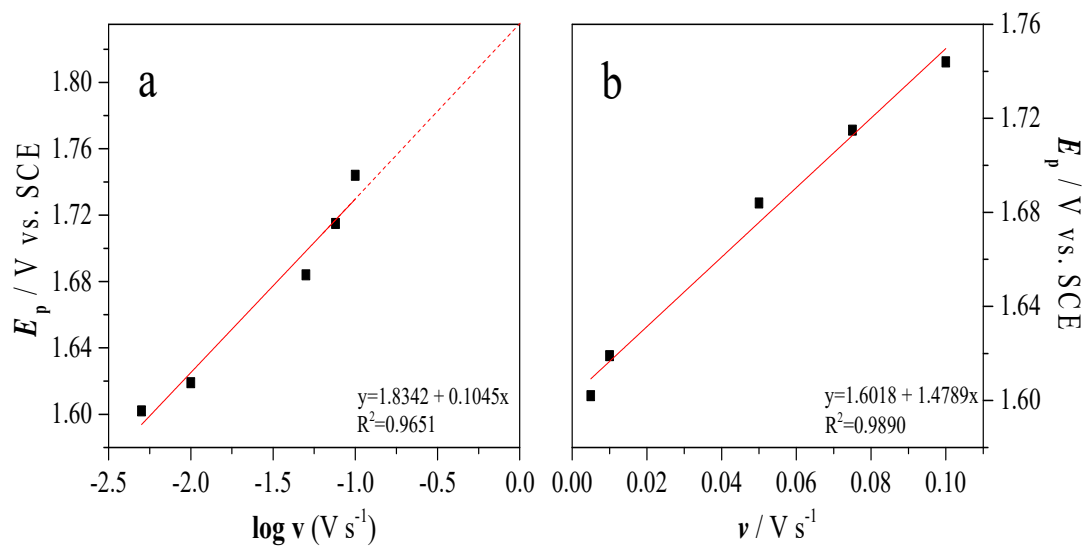
74 **Fig. S1.** (a) CV and (b) DPV curves obtained with 14 mg L^{-1} 4,6-DMDBT in ACN (93.5%

75 v/v)–water (6.5% v/v, 0.010 M LiClO_4) using a BDD electrode at $\nu = 0.005 \text{ V s}^{-1}$.



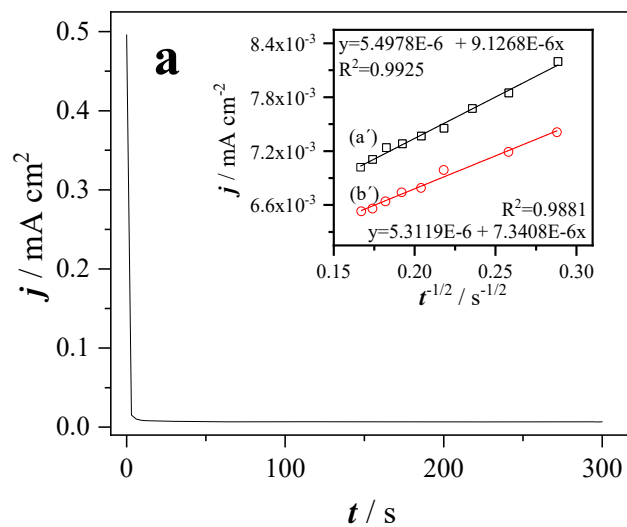
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77 **Fig. S2.** Variation of $(I_p v^{-1/2})$ vs. scan rate for peak I recorded by CV, as shown in Fig. 3a.



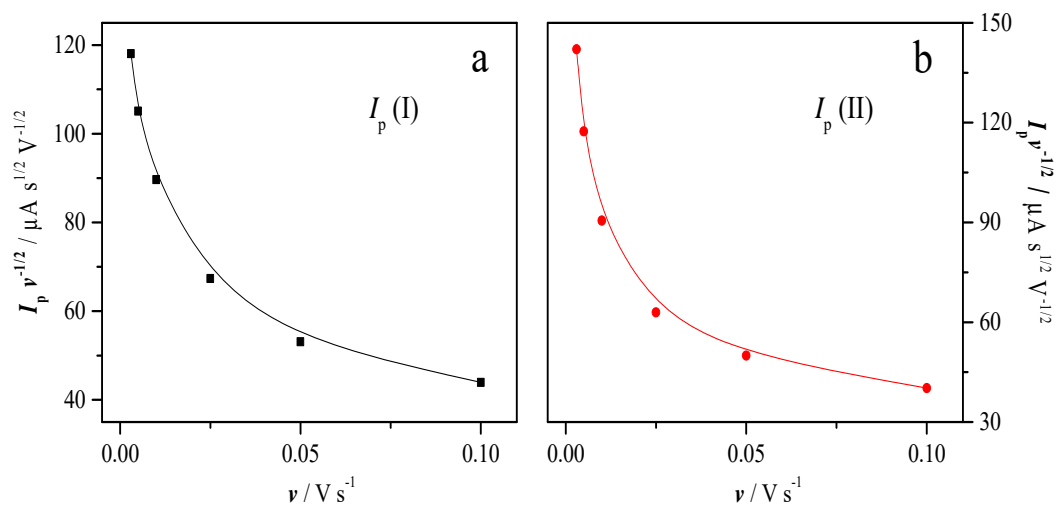
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79 **Fig. S3.** Variation of the peak potential with the (a) logarithm of scan rate and (b) scan rate,
 80 corresponding to the oxidation peak found in the cyclic voltammograms of Fig. 3a.



81

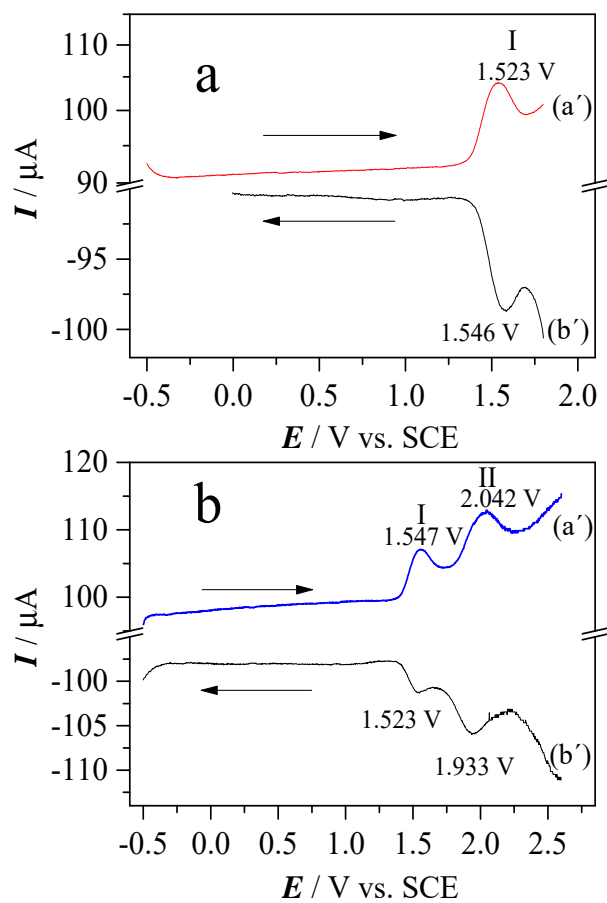
82 Fig. S4. (a) Chronoamperogram obtained with BDD in the presence of 27 mg L⁻¹ 4,6-
 83 DMDBT in ACN (93.5% v/v)–water (6.5% v/v, 0.010 M LiClO₄) solution at 1.5 V. Inset:
 84 Cottrell plot obtained from the chronoamperogram in the presence of (a') 27 mg L⁻¹ and (b')
 85 14 mg L⁻¹ of 4,6-DMDBT.



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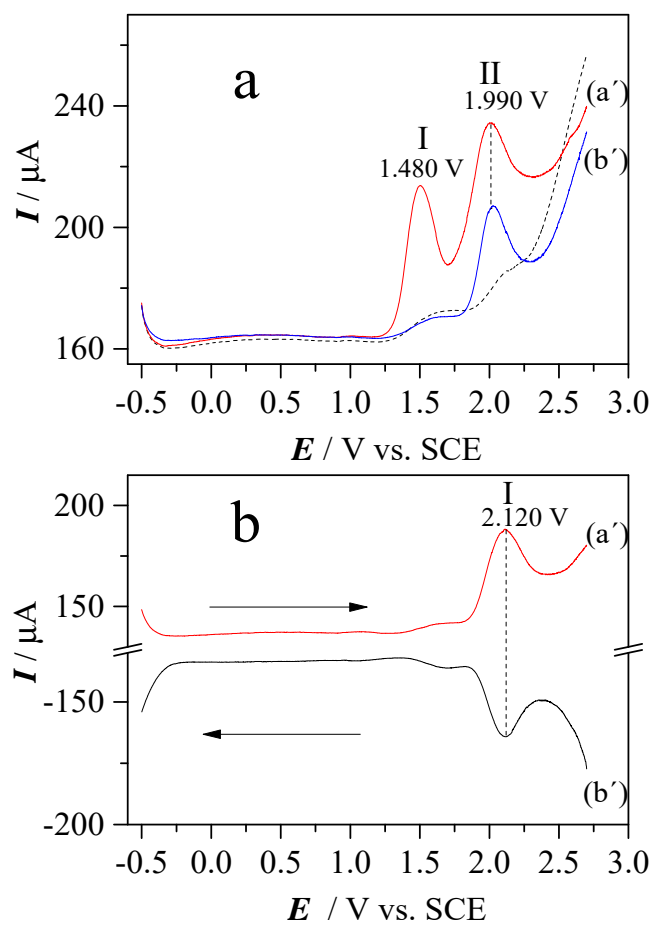
87 **Fig. S5.** Change of $(I_p v^{-1/2})$ with scan rate for peaks (a) I and (b) II recorded by DPV, as

88 shown in Fig. 4a.



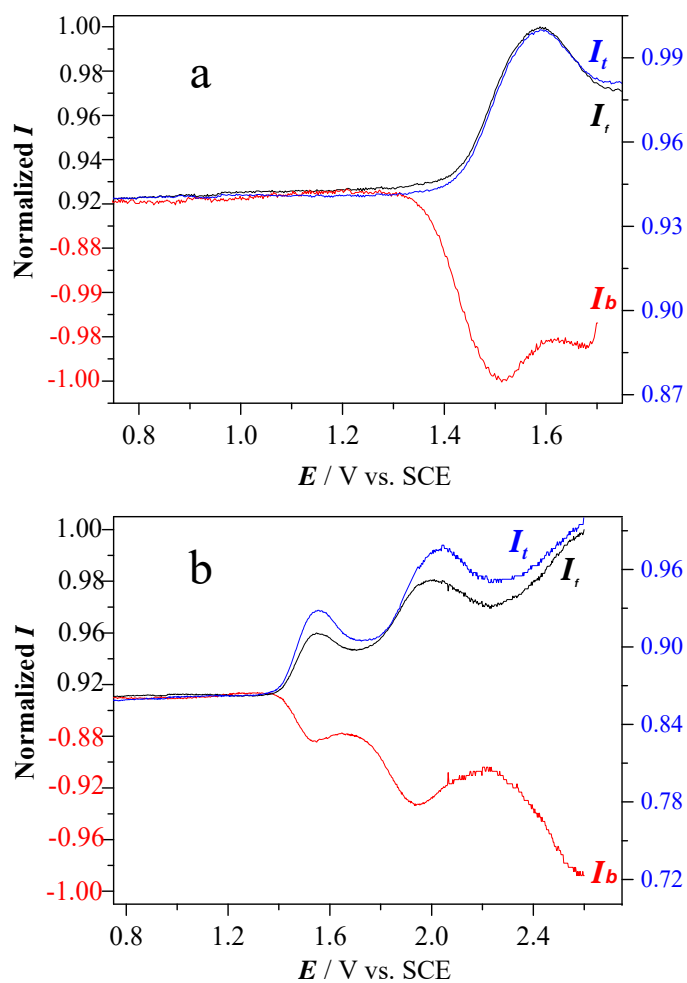
89

90 **Fig. S6.** DPV curves obtained with 14 mg L^{-1} 4,6-DMDBT in ACN (93.5% v/v)–water (6.5%
 91 v/v, 0.010 M LiClO_4) using a BDD electrode. (a) Potential range from -0.50 to $+1.75 \text{ V}$ for
 92 the (a') anodic and (b') cathodic scans at $\nu = 0.003 \text{ V s}^{-1}$. (b) Potential range from -0.50 to
 93 $+2.60 \text{ V}$ for the (a') anodic and (b') cathodic scans at $\nu = 0.003 \text{ V s}^{-1}$.



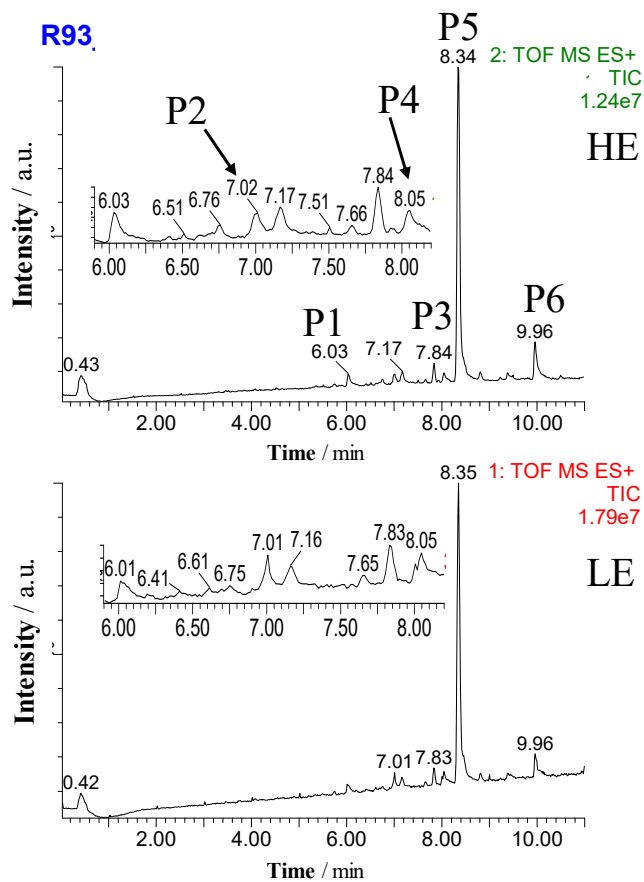
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95 **Fig. S7.** (a) DPV profiles recorded during oxidation scan in (---) ACN (93.5% v/v)–water
 96 (6.5% v/v, 0.010 M LiClO₄) using a BDD electrode at $\nu = 0.003 \text{ V s}^{-1}$, and in the presence of
 97 27 mg L⁻¹ of (a') DBT and (b') DBTO₂. (b) DPV curves obtained with 27 mg L⁻¹ DBTO₂ in
 98 the potential range from -0.50 to +2.75 V for the (a') anodic and (b') cathodic scans.



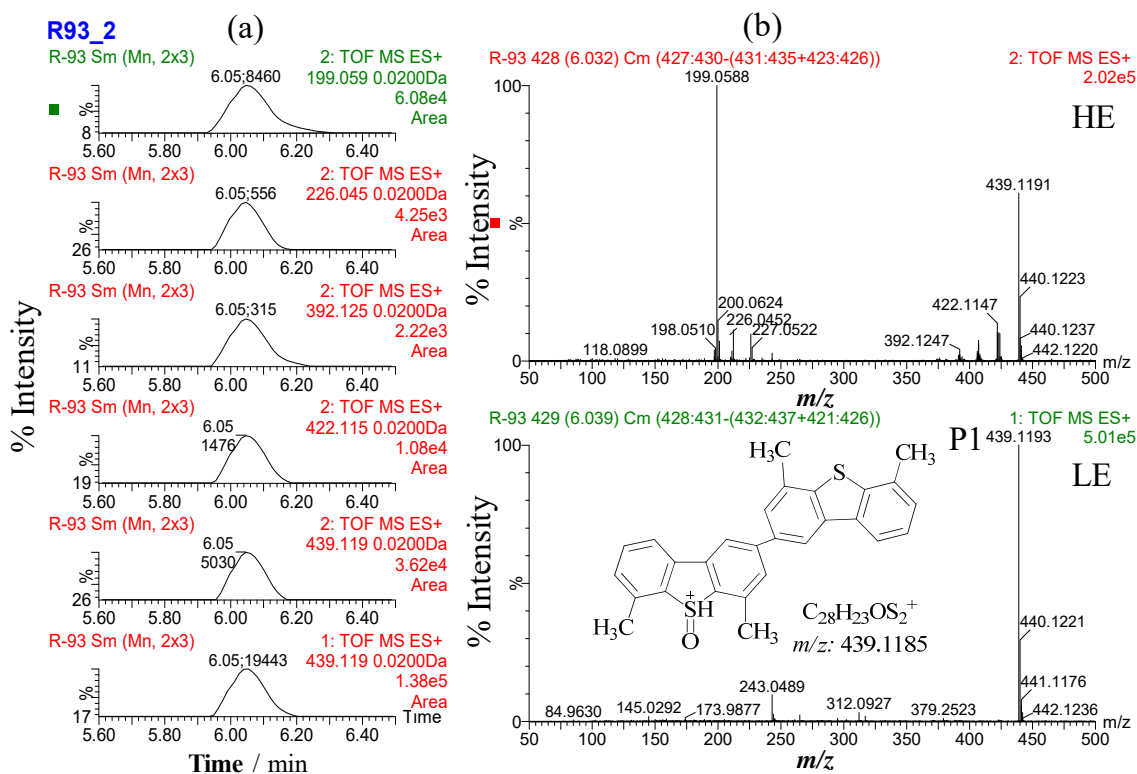
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100 **Fig. S8.** SWV curves determined with 14 mg L^{-1} 4,6-DMDBT in ACN (93.5% v/v)–water
 101 (6.5% v/v, 0.010 M LiClO_4) using a BDD electrode at a frequency of 8 Hz and $\nu = 0.020 \text{ V}$
 102 s^{-1} . Potential range: (a) from -0.50 to +1.80 V and (b) from -0.50 to +2.60 V. Current in the
 103 anodic (forward) scan: I_f ; current in the cathodic (backward) scan: I_b ; total current: I_t .



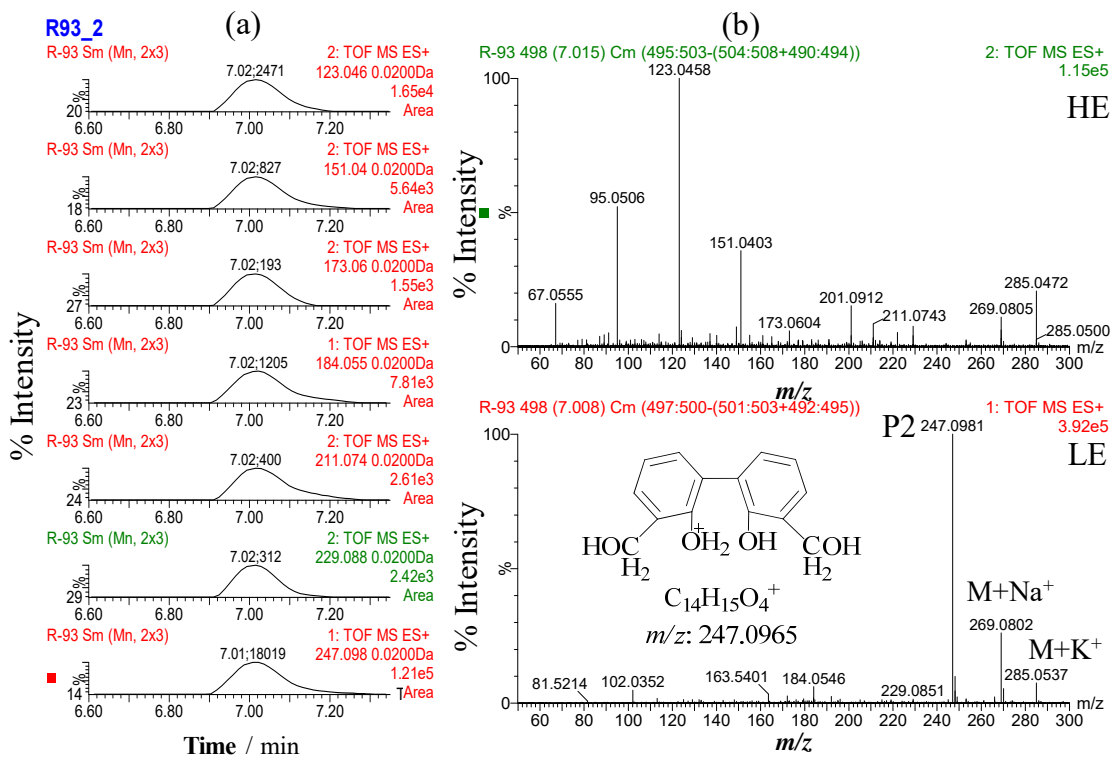
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105 **Fig. S9.** Total ion chromatograms obtained from the UHPLC-ESI-Q-TOF-MS analysis of
 106 samples collected after electrolysis of 27 mg L⁻¹ 4,6-DMDBT in ACN (93.5% v/v)-water
 107 (6.5% v/v, 0.010 M LiClO₄) for 90 min in a BDD/BDD cell at 1.50 V. High energy (HE) and
 108 low energy (LE).



109

110 **Fig. S10.** Detection and identification of compounds with possible chemical structure:
 111 $C_{28}H_{22}OS_2$ in the extracts obtained upon electrolysis of 27 mg L^{-1} 4,6-DMDBT at 1.5 V for
 112 90 min. LE and HE TOF mass spectra for the sample (b) and extracted ion chromatograms
 113 (XICs) at 20 mDa mass window (a) for $[M+H]^+$ in LE function and main fragments in HE
 114 function. Possible structure assigned by Mass Fragment software.



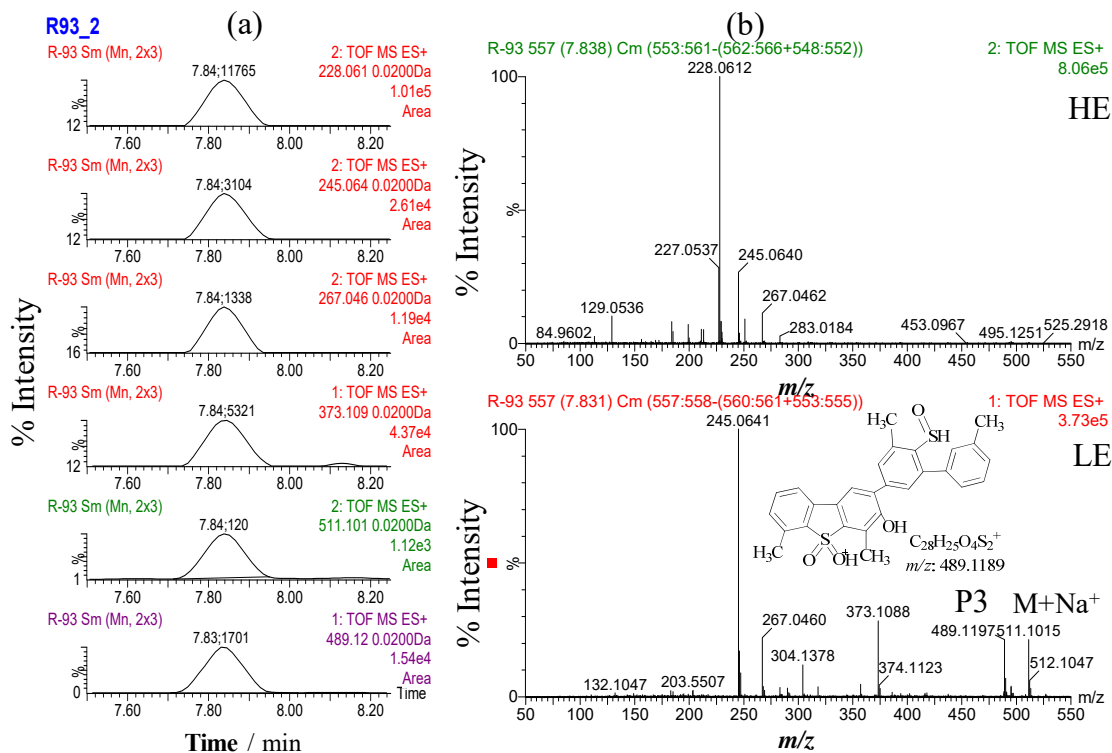
115

116 **Fig. S11.** Detection and identification of compounds with possible chemical structure:

117 $C_{14}H_{14}O_4$ in extracts obtained upon electrolysis of 27 mg L^{-1} 4,6-DMDBT at 1.5 V for 90

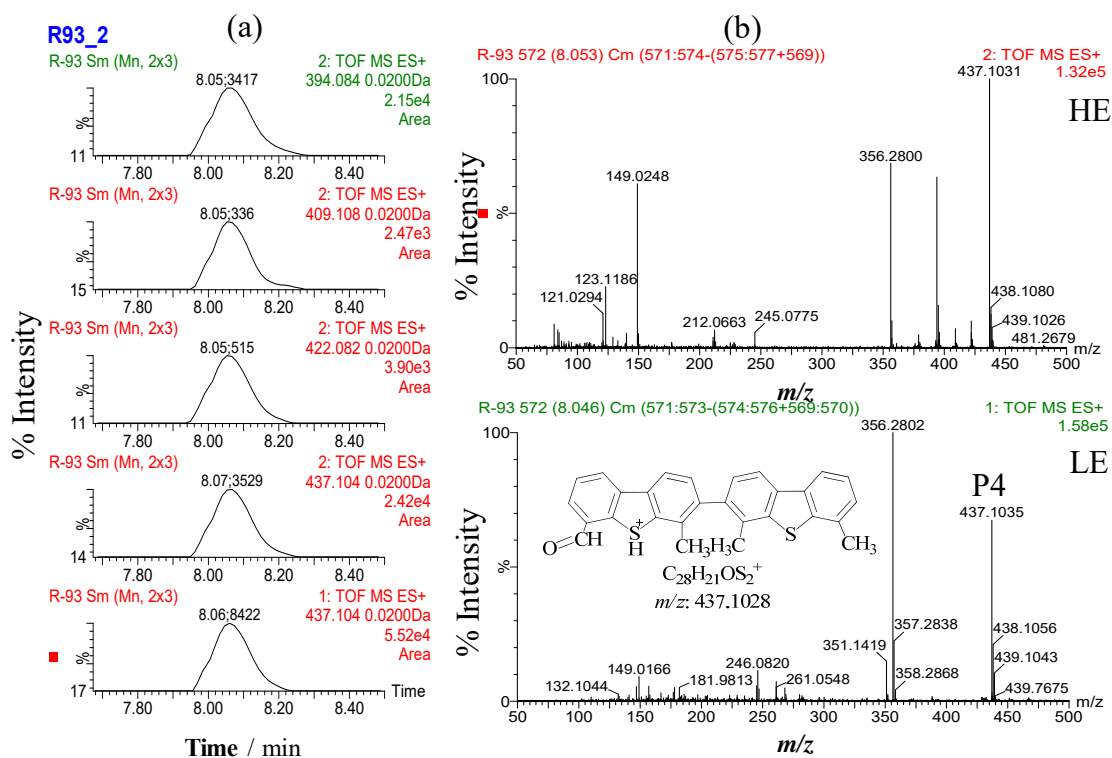
118 min. The LE and HE TOF mass spectra for (a) and (b), as well as the possible structure

119 assigned, were obtained upon the same conditions of Fig. S10.



120

121 **Fig. S12.** Detection and identification of compounds with possible chemical structure:
 122 $C_{28}H_{24}O_4S_2$ in extracts obtained upon electrolysis of 27 mg L^{-1} 4,6-DMDBT at 1.5 V for 90
 123 min. The LE and HE TOF mass spectra for (a) and (b), as well as the possible structure
 124 assigned, were obtained upon the same conditions of Fig. S10.



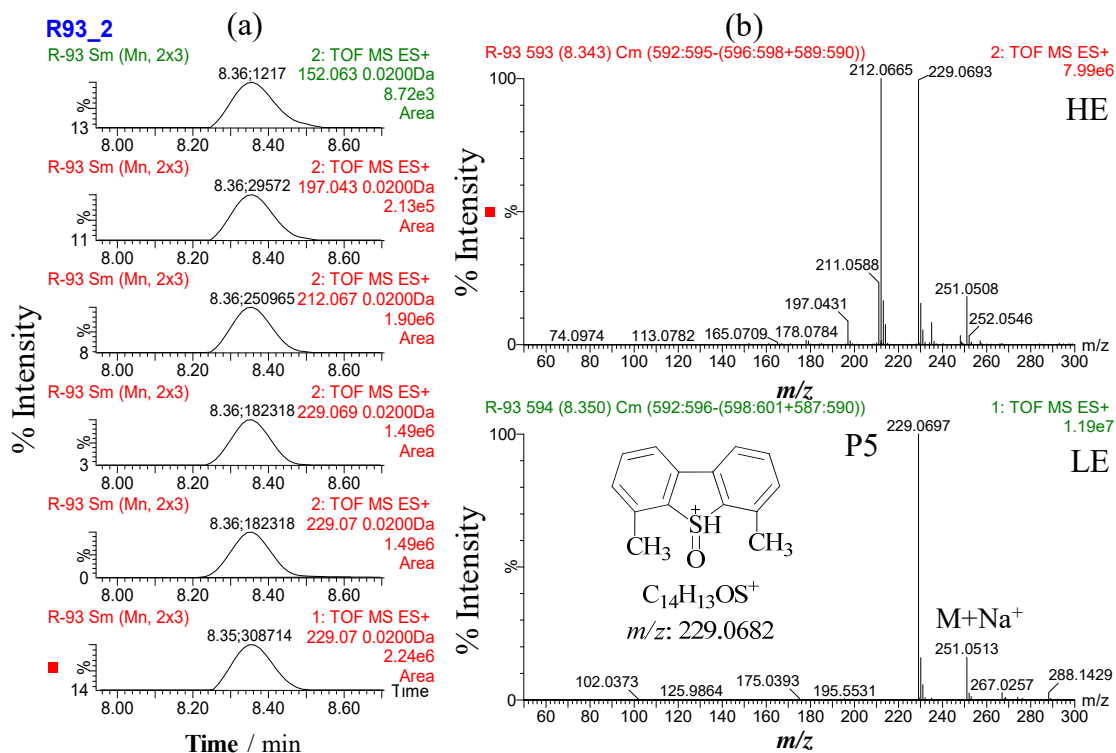
125

126 **Fig. S13.** Detection and identification of compounds with possible chemical structure:

127 C₂₈H₂₀OS₂ in extracts obtained upon electrolysis of 27 mg L⁻¹ 4,6-DMDBT at 1.5 V for 90

128 min. The LE and HE TOF mass spectra for (a) and (b), as well as the possible structure

129 assigned, were obtained upon the same conditions of Fig. S10.



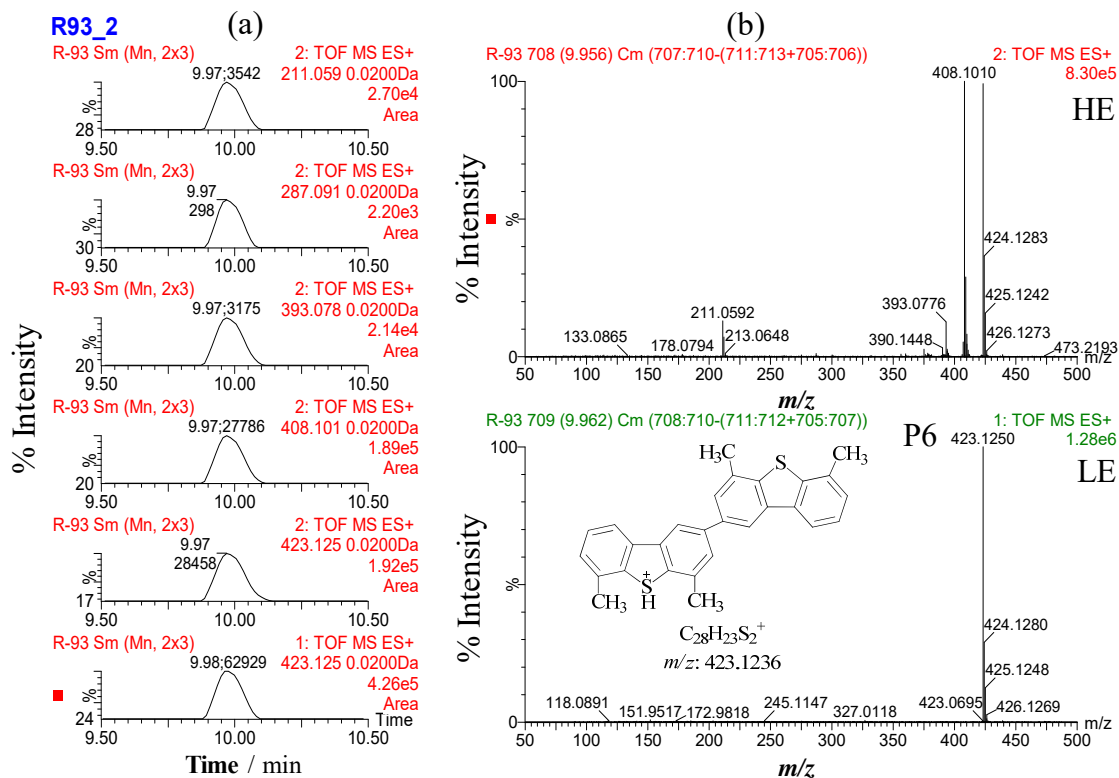
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131 **Fig. S14.** Detection and identification of compounds with possible chemical structure:

132 $C_{14}H_{12}OS$ in extracts obtained upon electrolysis of 27 mg L^{-1} 4,6-DMDBT at 1.5 V for 90

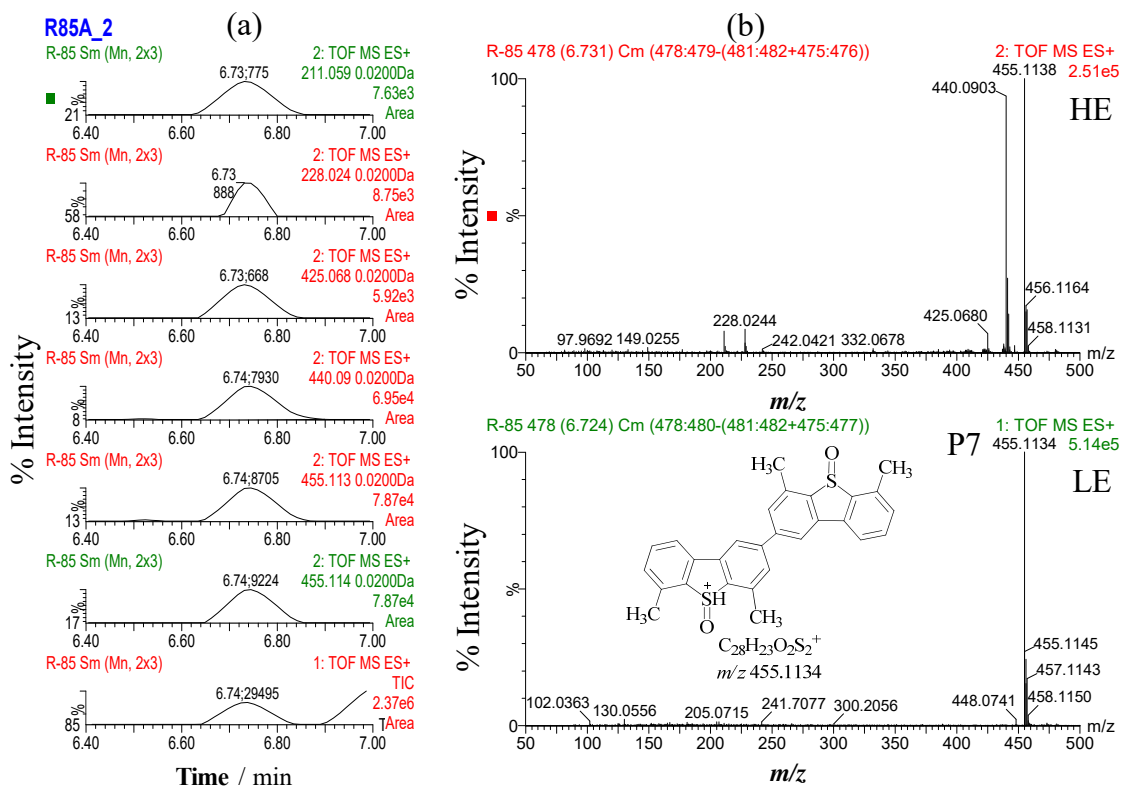
133 min. The LE and HE TOF mass spectra for (a) and (b), as well as the possible structure

134 assigned, were obtained upon the same conditions of Fig. S10.



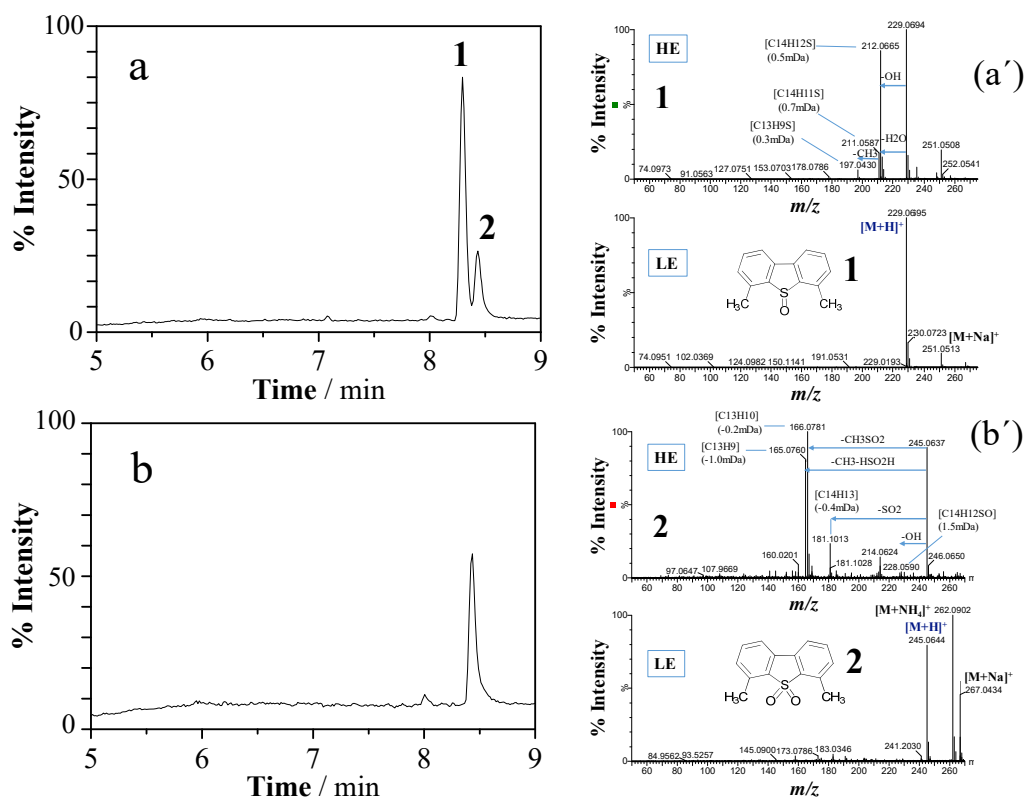
135

136 **Fig. S15.** Detection and identification of compounds with possible chemical structure:
 137 $C_{28}H_{22}S_2$ in extracts obtained upon electrolysis of 27 mg L^{-1} 4,6-DMDBT at 1.5 V for 90
 138 min. The LE and HE TOF mass spectra for (a) and (b), as well as the possible structure
 139 assigned, were obtained upon the same conditions of Fig. S10.



140

141 **Fig. S16.** Detection and identification of compounds with possible chemical structure:
 142 $C_{28}H_{22}O_2S_2$ in extracts obtained upon electrolysis of 27 mg L^{-1} 4,6-DMDBT at 1.5 V for 90
 143 min. The LE and HE TOF mass spectra for (a) and (b), as well as the possible structure
 144 assigned, were obtained upon the same conditions of Fig. S10.



145

146 **Fig. S17.** Total ion chromatograms obtained from the UHPLC-ESI-Q-TOF-MS analysis of
 147 samples collected after electrolysis of 27 mg L⁻¹ 4,6-DMDBT in ACN (93.5% v/v)–water
 148 (6.5% v/v, 0.010 M LiClO₄) for 240 min in a BDD/BDD cell, at (a) 1.50 and (b) 2.00 V. HE
 149 and LE mass spectra for (a') 4,6-DMDBTO at retention time of 8.30 min (compound 1) and
 150 (b') 4,6-DMDBTO₂ at retention time of 8.43 min (compound 2).

151 Table S1. Products, t_R and ESI-TOF-MS mass spectral properties of fragments formed from bulk
 152 electrolysis carried out for 90 min at $E_{an} = 1.5$ V.

Peak	t_R (min)	Experimental mass (m/z)	mDa	Elemental composition	Fragments (m/z)	mDa	Lost fragments
P1	6.04	439.1191	0.1	$C_{28}H_{23}OS_2^+$	422.1147	-1.6	OH
					392.1247	1.2	CH ₃
					226.0452	0.0	C ₁₄ H ₁₃ S
					199.0588	0.7	C ₁₅ H ₁₂ OS
P2	7.01	247.0982	1.2	$C_{14}H_{15}O_4^+$	229.0865	-1.0	H ₂ O
					211.0793	-1.3	2H ₂ O
					184.0546	2.2	C ₂ H ₇ O ₂
					173.0606	0.3	C ₄ H ₆ O ₂
					151.0404	0.9	C ₆ H ₈ O
					123.0458	1.2	C ₇ H ₈ O ₂
P3	7.83	489.1197	0.3	$C_{28}H_{25}O_4S_2^+$	373.1088	0.3	C ₄ H ₄ O ₄
					267.0462	-1.8	C ₁₂ H ₁₄ O ₂ S
					245.0640	0.4	C ₁₄ H ₁₂ O ₂ S
					228.0612	0.3	C ₁₄ H ₁₃ O ₃ S
P4	8.04	437.1031	-0.3	$C_{28}H_{21}OS_2^+$	422.0816	-0.8	CH ₃
					409.1082	1.7	CO
					394.0845	-0.5	C ₂ H ₃ O
P5	8.35	229.0693	0.6	$C_{14}H_{13}OS^+$	212.0665	0.5	OH
					197.0431	0.6	CH ₄ O
					178.0783	0.0	H ₃ OS
					152.0628	0.2	C ₂ H ₅ OS
P6	9.96	423.1248	0.7	$C_{28}H_{23}S_2^+$	408.1010	0.4	CH ₃
					393.0776	0.4	2CH ₃
					287.0911	1.7	C ₈ H ₈ S
					211.0592	1.1	C ₁₀ H ₁₂ S
					178.0796	1.3	C ₁₄ H ₁₃ S ₂
*P7	6.74	455.1138	-0.1	$C_{28}H_{23}O_2S_2^+$	440.0905	-0.2	CH ₃
					425.0680	1.0	2CH ₃
					228.0244	-	C ₁₄ H ₁₁ OS
					211.0588	0.7	C ₁₄ H ₁₂ O ₂ S

153 * P7 Product obtained for long electrolysis time of 240 min