

1 **Responses of plants to climate change: Metabolic changes during**
2 **abiotic stress combination in plants**

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25 Reprogramming plant metabolism under combined stresses.

26 **Highlight**

27 The specific and complex metabolic responses of plants to a combination of different
28 abiotic stresses, that is predicted to worsen with climate change, is reviewed.

29

30 **Abstract**

31 Climate change is predicted to increase the frequency and intensity of abiotic stress
32 combinations that negatively impact plants and pose a serious threat to crop yield and
33 food supply. Plants respond to episodes of stress combination by activating specific
34 physiological and molecular responses, as well as by adjusting different metabolic
35 pathways, to mitigate the negative effects of stress combination on plant growth,
36 development, and reproduction. Plants synthesize a wide range of metabolites that
37 regulate many aspects of plant growth and development, as well as plant responses to
38 stress. Although metabolic responses to individual abiotic stresses have been studied
39 extensively in different plant species, recent efforts have been directed at understanding
40 metabolic responses that occur when different abiotic factors are combined. In this review
41 we examine recent studies of metabolomic changes under stress combination in different
42 plants and suggest new avenues for the development of stress combination-resilient crops
43 based on metabolites as breeding targets.

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46

47 **Key words**

48 Abiotic stress, Climate change, Global warming, Hormones, Metabolites, Primary
49 metabolism, Secondary metabolism, Stress combination.

50 **Abbreviations**

51 ABA, abscisic acid; ABR1, ABA Repressor 1; APX1, Ascorbate Peroxidase 1; ET,
52 ethylene; GABA, γ -aminobutyric acid; JA, jasmonic acid; MBF1c, Multiprotein Bridging
53 Factor 1c; RD26, Responsive to Desiccation 26; ROS, reactive oxygen species; SA,
54 salicylic acid; TCA, tricarboxylic acid.

55 **Introduction**

56 Plant growth, yield and survival are highly affected by global warming and climate
57 change that cause a constant increase in global atmospheric CO₂ levels, as well as the
58 intensity and frequency of different abiotic stresses including drought, extreme
59 temperatures, and flooding; resulting in many cases in a multifactorial stress combination
60 phenomenon (Steg, 2018; Masson-Delmotte *et al.*, 2021; Zandalinas *et al.*, 2021*b,a*; von
61 der Gathen *et al.*, 2021; Houtan *et al.*, 2021; Rivero *et al.*, 2022). The predicted increase
62 in soil-associated stresses such as salinity, inorganic nutrient imbalances or heavy metals,
63 alongside biotic stresses, further adds to the complexity of the future plant environment
64 and make it difficult to forecast how plants will respond to the challenges posed by the
65 interactions of multiple stresses. In addition to the activation of specific physiological and
66 molecular responses to stress, plants adjust several metabolic pathways to mitigate the
67 effects of stress on growth and development and to adapt to new energetic demands
68 imposed by different climatic and environmental scenarios (Dusenge *et al.*, 2019; Fernie
69 *et al.*, 2020). The efficiency of reconfiguring metabolic networks, resuming active growth
70 and seed production, and restoring a new state of homeostasis after stress release will
71 further determine how successful a plant is in acclimating to stress (Shulaev *et al.*, 2008;
72 Dusenge *et al.*, 2019).

73 Plants produce a wide range of different metabolites in a spatiotemporal- and/or
74 environment-dependent manner (Fang *et al.*, 2019), including carbohydrates, amino
75 acids, phenolics, polyols, polyamines, lipids, and others, with many different biological
76 functions. This large diversity is traditionally divided into primary metabolites, which are
77 directly required for plant growth [*e.g.*, carbohydrates, tricarboxylic acid (TCA) cycle
78 intermediates or amino acids]; secondary metabolites, which are involved in regulating
79 plant–environment interactions (*e.g.*, phenolics, terpenes or nitrogen-containing
80 compounds); and hormones, that control many aspects of plant growth and development,
81 as well as plant responses to stress [*e.g.*, abscisic acid (ABA), jasmonic acid (JA), salicylic
82 acid (SA) or ethylene (ET)] (Fang *et al.*, 2019; Erb and Kliebenstein, 2020). It was
83 recently proposed that some secondary metabolites are multifunctional and could function
84 as potent regulators of plant growth and development processes, as well as defense against
85 pathogens, thus acting as plant regulatory compounds or hormones. For example,
86 secondary metabolites such as flavonoids were implicated in regulating plant growth and
87 development, as well as environmental responses (Erb and Kliebenstein, 2020). In

88 addition to being considered as downstream targets of many stress-response signaling
89 pathways, it was suggested that different metabolites also function as stress signals. In
90 this sense, metabolic responses generated by specific abiotic stresses that persist in the
91 plant could prime metabolic responses for subsequent environmental stresses
92 (Schwachtje *et al.*, 2019). Examples of stress-induced metabolic signals include the
93 amino acid proline, that is triggered by drought in *Periploca sepium* (An *et al.*, 2013) or
94 tobacco plants (Vanková *et al.*, 2012). Other metabolic signals consisting of primary
95 metabolites (*e.g.*, sugars, sugar phosphates and different amino acids) and/or secondary
96 metabolism, were previously described in response to drought (*e.g.*, Moyankova *et al.*,
97 2014; Wedeking *et al.*, 2018), cold (*e.g.*, Kaplan *et al.*, 2004; Mazzucotelli *et al.*, 2006)
98 or heat stress (*e.g.*, Hemme *et al.*, 2014). Plants can therefore produce a wide array of
99 variable and flexible regulators to adjust their growth and development, and to survive
100 challenging environmental habitats.

101 Metabolic responses to single abiotic stresses are well described for many plant systems
102 and in both perennial and annual plants (*e.g.*, Wang *et al.*, 2016b, 2020; Kang *et al.*, 2019;
103 Muchate *et al.*, 2019; Itam *et al.*, 2020; Cai *et al.*, 2020). However, plants growing in the
104 field are typically subjected to more than one abiotic stress condition at a time (*i.e.*, stress
105 combination), imposing new and unique metabolic demands on plants (Mittler, 2006;
106 Zandalinas *et al.*, 2018; Rivero *et al.*, 2022). Although considerable research efforts have
107 been invested in the last decade in dissecting plant responses to a combination of two or
108 three different stresses (*e.g.*, Vile *et al.*, 2012; Iyer *et al.*, 2013; Zandalinas *et al.*, 2020a,b,
109 2021b; Cohen *et al.*, 2021), a deeper understanding of the metabolic response of plants to
110 multiple stress factors under field conditions is crucial for improving stress-tolerance of
111 different crops. Because the plant metabolome plays such a crucial role in defining the
112 phenotype of plants during stress (Fang *et al.*, 2019; Sharma *et al.*, 2021; Carrera *et al.*,
113 2021), studies of the metabolome of plants grown under multiple stress combinations
114 could be used to identify stress resistance-associated metabolites, which can then be used
115 in future metabolic engineering strategies, or breeding programs, as selective markers
116 (Abdelrahman *et al.*, 2018). In addition, since annual and perennial plant species
117 generally adopt different strategies to cope with abiotic stresses, future research should
118 consider these differences in dissecting and identifying stress-specific markers under
119 multiple stress combinations.

120 In this review, we describe recent research efforts focused on studying the metabolomic
121 responses of plants to different abiotic stress conditions and their combination and
122 propose stress combination-resistance metabolites as potential targets for plant breeding.
123 Unraveling the complexities of the plant metabolomic responses to a combination of
124 multiple abiotic stresses could facilitate the development of climate-resilient crops,
125 improving global food supply.

126

127 **Primary metabolic pathways impacted by abiotic stress combination**

128 The general primary response of plants to abiotic stress includes a reduction in energy
129 consumption and overall protein biosynthesis in order to shift from plant growth to
130 protective mechanism, adjusting various primary metabolic pathways, including
131 carbohydrate, amino acid and polyamine metabolism (Zhang *et al.*, 2022). Below we
132 describe specific and common changes in primary metabolic pathways in response to
133 different abiotic stresses and their combination (Fig. 1).

134 *Sugars*

135 Different abiotic conditions and their combinations induce stomatal closure to minimize
136 water loss. Examples for these include drought (*e.g.*, Flexas *et al.*, 2002, 2004; Carmo-
137 Silva *et al.*, 2012; Medina and Gilbert, 2016; Zhang *et al.*, 2021), high light (*e.g.*,
138 Devireddy *et al.*, 2018, 2020a; Zandalinas *et al.*, 2020a), and salinity (*e.g.*, Flexas *et al.*,
139 2004; Orsini *et al.*, 2012; Zhu *et al.*, 2014; Hedrich and Shabala, 2018), as well as
140 combined drought and heat (Rizhsky *et al.*, 2002, 2004; Carmo-Silva *et al.*, 2012;
141 Zandalinas *et al.*, 2016a; Zhou *et al.*, 2017; Shaar-Moshe *et al.*, 2017), salinity and heat
142 (Shaar-Moshe *et al.*, 2017; Balfagón *et al.*, 2019b), or salinity and drought (Shaar-Moshe
143 *et al.*, 2017; Dugasa *et al.*, 2019). This response is accompanied by a decrease in the
144 intercellular CO₂ concentration of plants, limiting photosynthesis (Chaves *et al.*, 2003),
145 that in turn alters carbohydrate levels. Carbohydrates are direct products of photosynthetic
146 CO₂ fixation and are substrates for biomass accumulation and biosynthesis of other
147 organic molecules such as sugar alcohols. In addition, they play a central role in stress
148 perception and signaling, function as a regulatory hub for stress-mediated gene
149 expression, and act as stabilizers of membranes and proteins as well as osmoprotectants
150 and reactive oxygen species (ROS) quenchers (Sami *et al.*, 2016; Pommerrenig *et al.*,
151 2018). Therefore, a fine-tuned reconfiguration of sugar accumulation under stress

152 combination is crucial for plant acclimation and survival. Different abiotic stresses result
153 in distinct, and sometimes opposing, patterns of sugar accumulation (Fig. 1), and this
154 pattern depends on the type of stress as well as the plant species. For example, in response
155 to cold stress, the pathway leading to raffinose was shown to be crucial for freezing
156 tolerance in *Arabidopsis* (Korn *et al.*, 2010). Heat stress caused a decrease in glucose in
157 soybean plants (Das *et al.*, 2017), but an increase in soluble sugars in *Arabidopsis* plants
158 (Wang *et al.*, 2020a). In addition, in both plant species (*Arabidopsis* and soybean), heat
159 stress inhibited the accumulation of metabolites involved in the TCA cycle (Das *et al.*,
160 2017; Wang *et al.*, 2020a). In response to drought stress, many of the major carbohydrates
161 such as glucose, fructose and sucrose accumulated in soybean (Mutava *et al.*, 2015; Das
162 *et al.*, 2017) as well as in *Arabidopsis* (Rizhsky *et al.*, 2004). In addition to drought,
163 changes in the amounts of carbohydrates (including trehalose, glucose and fructose) were
164 determined in leaves and roots of soybean plants subjected to flooding (Coutinho *et al.*,
165 2018). Salinity stress caused an increased abundance of sugars and sugar derivatives such
166 as arabinose, inositol, mannose, sucrose, trehalose, xylose, and galactose in sugar beet
167 (Hossain *et al.*, 2017). Moreover, high amounts of hexoses, sugar alcohols and organic
168 acids were found in the salt-tolerant *Thellungiella halophila* plants (Gong *et al.*, 2005),
169 demonstrating the protective functions of sugars under salt stress.

170 In contrast to individual stresses, different combinations of adverse conditions displayed
171 unique patterns of sugar accumulation (Rizhsky *et al.*, 2004; Wulff-Zottele *et al.*, 2010;
172 Rivero *et al.*, 2014; Obata *et al.*, 2015; Zandalinas *et al.*, 2016c; Zinta *et al.*, 2018;
173 Weiszmann *et al.*, 2018; Balfagón *et al.*, 2022a) (Fig. 1). A metabolite profiling analysis
174 of *Arabidopsis* subjected to a combination of drought and heat stress revealed that plants
175 accumulated sucrose and other sugars such as maltose and gulose exclusively under this
176 stress combination. In addition, it was suggested that a combination of drought and heat
177 stress required sucrose rather than proline (accumulated only under drought applied
178 individually) as an osmoprotectant, and that sucrose biosynthesis could occur from starch
179 in *Arabidopsis* plants (Rizhsky *et al.*, 2004). In maize subjected to a combination of
180 drought and heat, metabolite profiles were analyzed and contrasted with grain yield in the
181 field (Obata *et al.*, 2015). Metabolic responses under this stress combination could be
182 mostly predicted from the sum of those in single stresses, but eight metabolites, including
183 succinate, raffinose and xylose, were specifically regulated under the combined stress
184 conditions (Obata *et al.*, 2015). Interestingly, elevated CO₂ levels reduced the impact of

185 a combination of drought and heat on sugar metabolism, showing less accumulation of
186 soluble sugars in Arabidopsis plants, pointing towards the role of high CO₂ in stress
187 combination mitigation (Zinta *et al.*, 2018). In addition to the possible beneficial role of
188 CO₂ in alleviating damages from a combination of drought and heat stress, it was reported
189 that elevated CO₂ decreased iron and zinc concentration in soybean seeds when plants
190 where growing in the field, heat stress had the opposite effect, and the combination of
191 elevated CO₂ and heat generally restored seed iron and zinc concentrations to levels
192 obtained under ambient CO₂ and temperature conditions (Köhler *et al.*, 2019). These
193 results suggest that increased levels of CO₂ due to climate change could be beneficial for
194 the nutritional quality of crops. However, it should be noted that C3 grains and legumes
195 had lower concentrations of zinc and iron when grown under field conditions at elevated
196 atmospheric CO₂ concentration (Myers *et al.*, 2014), highlighting the complexity of
197 predicting climate change effects on food and nutritional security. In addition to the
198 combination of drought and heat, it was recently reported in Arabidopsis that a
199 combination of high light and heat stress resulted in a unique metabolic response that
200 included increased accumulation of sugars (including maltose, glucose and fructose),
201 coupled with decreased levels of metabolites participating in the TCA cycle (including
202 citrate and fumarate), compared to the individual stresses (Balfagón *et al.*, 2022a).
203 Similarly, the combination of high light and sulfur depletion had a profound effect on
204 central metabolic pathways including the TCA cycle and glycolysis. Specifically, sugars
205 such as glucose, fructose and raffinose were elevated compared to control (Wulff-Zottele
206 *et al.*, 2010). Another stress combination that affected the accumulation of sugars in
207 Arabidopsis was cold and high light stress. The freezing-tolerant Arabidopsis Rsch
208 accession significantly increased sucrose, fructose and glucose concentrations in response
209 to this stress combination, and a role for vacuolar invertase activity in preventing a
210 limitation in cytosolic hexose metabolism under stress was suggested (Weiszmann *et al.*,
211 2018). An increased content of fructose, glucose and UDP-glucose was also found in the
212 combined stress-tolerant tomato line RIL-76 under a combination of heat and salinity,
213 indicating that the level of this specific sugar could be used for the selection of varieties
214 more resistant to conditions of climate change (Lopez-Delacalle *et al.*, 2021).
215 Furthermore, tomato plants subjected to a combination of heat and salinity specifically
216 accumulated trehalose, that could have an important role in protecting photosynthetic
217 proteins from this stress combination (Rivero *et al.*, 2014).

218 Taken together, a general increase in soluble sugar accumulation seems to be a common
219 response to different stress combinations, that could be a result of reduced plant growth
220 (decreasing the demand for primary metabolites) as well as impaired photosynthetic rate.
221 In turn, the decreased photosynthetic rate under stress combination would indicate that
222 the increase in sugar levels could come from starch breakdown instead of carbon fixation.
223 Therefore, remobilizing starch reserves to produce energy, and sugar and derived
224 metabolites when photosynthesis is suppressed could provide metabolites that act as
225 osmoprotectants and compatible solutes to alleviate the negative effect of different stress
226 combinations (Thalmann and Santelia, 2017). It should be noted that in the future the
227 combination of different abiotic stresses will be accompanied by an elevation in CO₂
228 levels under climate change conditions, and high CO₂ concentrations are expected to
229 increase leaf photosynthetic rates. However, the degree to which this will occur is
230 unknown, given that the stimulation of photosynthesis by CO₂ depends on leaf
231 temperature, and water and nutrient availability (Dusenge *et al.*, 2019). Plant respiration
232 could be also altered by elevated CO₂ levels due to its effects on stomatal aperture.
233 Dusenge *et al.* (2019) reviewed recent literature focused on increased CO₂ levels and plant
234 respiration, concluding that responses of respiration to changes in CO₂ are highly variable
235 and that a consistent theoretical basis for making strong predictions for how respiration
236 will change when CO₂ levels are high are still unclear (Dusenge *et al.*, 2019).

237 *Amino acids*

238 Like sugars, the accumulation of amino acids during different abiotic stresses can also
239 contribute to osmotic adjustments and ROS scavenging (Hildebrandt, 2018). Moreover,
240 amino acids can function as alternative substrates for mitochondrial respiration when
241 carbohydrate supply is insufficient due to a decrease in photosynthesis rate, as well as
242 serve as precursors of secondary metabolites (Hildebrandt, 2018; Batista-Silva *et al.*,
243 2019; Heinemann and Hildebrandt, 2021). The accumulation of amino acids resulting
244 from protein breakdown or *de novo* biosynthesis can therefore minimize the adverse
245 effects of different abiotic stresses (Lugan *et al.*, 2010; Krasensky and Jonak, 2012; Obata
246 and Fernie, 2012; Hildebrandt, 2018). For example, during cold stress, amino acids play
247 an important role in N fixation into glutamine, and the synthesis of different proteins as
248 well as polyamines, phenylpropanoids, glucosinolates, auxins and indole alkaloids
249 (Alcázar *et al.*, 2006; Bernard and Habash, 2009; Amir, 2010; Tzin and Galili, 2010;
250 Hildebrandt, 2018). In *Arabidopsis*, the catabolic intermediates of lysine and the

251 branched-chain amino acids valine, leucine and isoleucine represent key factors in
252 drought tolerance (Pires *et al.*, 2016), and the activation of biosynthetic pathways of
253 serine, arginine, glutamine and alanine during osmotic and salt stress are associated with
254 storage of precursors for protein synthesis to prepare for rapid recovery of plant
255 metabolism following stress (Batista-Silva *et al.*, 2019). Proline is thought to be crucial
256 for acclimation to drought, cold stress and salinity but not to heat stress (Rizhsky *et al.*,
257 2004; Kaplan *et al.*, 2004; Gagneul *et al.*, 2007; Usadel *et al.*, 2008; Kempa *et al.*, 2008;
258 Urano *et al.*, 2009; Szabados and Savouré, 2010; Lugan *et al.*, 2010; Lv *et al.*, 2011; Per
259 *et al.*, 2017). In addition, the non-proteinogenic amino acid γ -aminobutyric acid (GABA)
260 accumulates in response to drought, hypoxia, anoxia, heat stress, salinity, zinc, osmotic
261 stress, wounding and oxidative stress (Bouché and Fromm, 2004; Xing *et al.*, 2007;
262 Miyashita and Good, 2008; Fait *et al.*, 2008; Allan *et al.*, 2008; Bor *et al.*, 2009; Renault
263 *et al.*, 2010; Akçay *et al.*, 2012; Nayyar *et al.*, 2014; Vijayakumari and Puthur, 2016; Daş
264 *et al.*, 2016; Mei *et al.*, 2016; Mekonnen *et al.*, 2016; Salvatierra *et al.*, 2016; Bown and
265 Shelp, 2016; Wang *et al.*, 2017; Scholz *et al.*, 2017; Priya *et al.*, 2019; Xu *et al.*, 2021;
266 Wu *et al.*, 2021) (Table 1).

267 As shown for sugar metabolism, a combination of different abiotic stresses results in
268 different patterns of amino acid accumulation compared to those caused by individual
269 stresses. These unique patterns can be correlated with the specific demands the combined
270 stress imposes on plants. For example, valine, asparagine and glutamine significantly
271 accumulated in the tomato tolerant line RIL-76 under a combination of salinity and heat,
272 and this accumulation was correlated with better nitrogen use efficiency compared to that
273 of the sensitive tomato line RIL-66 (Lopez-Delacalle *et al.*, 2020). In response to a
274 combination of drought and heat stress, the specific accumulation of certain amino acids
275 including glutamate, tyrosine, valine, and tryptophan in Purslane plants was thought to
276 play a role in the cellular osmotic adjustment aimed at maintaining leaf turgor during
277 stress combination (Jin *et al.*, 2016). In addition, a stress combination induced by two
278 metals, Co and Cu, led to a higher amino acid accumulation in the tolerant barley
279 genotype Yan66 (Lwalaba *et al.*, 2020), suggesting that amino acids could confer
280 resistance to toxic levels of metals due to their potential metal-binding capacity (Anjum
281 *et al.*, 2015; Yuan *et al.*, 2020; Lwalaba *et al.*, 2020). Special attention has been recently
282 given to GABA in response to different abiotic stress combinations (Table 1). The tomato
283 tolerant line RIL-76 subjected to a combination of salinity and heat (Lopez-Delacalle *et*

284 *al.*, 2020), as well as *Arabidopsis* plants subjected to a combination of high light and heat
285 stress (Balfagón *et al.*, 2022a), specifically accumulated this amino acid in response to
286 stress combination. GABA also accumulates under combinations of salinity and drought,
287 and heat and drought, as well as salinity, drought and heat, in *Brachypodium distachyon*
288 plants (Shaar-Moshe *et al.*, 2019). In addition to its involvement in plant growth
289 regulation and stomatal responses, and its proposed role in enhancing plant tolerance to
290 different environmental stresses (Bouché and Fromm, 2004; Shi *et al.*, 2010; Shang *et al.*,
291 2011; Yu *et al.*, 2014; Mekonnen *et al.*, 2016; Salvatierra *et al.*, 2016; Li *et al.*, 2016b;
292 Seifikalhor *et al.*, 2019, 2020; Priya *et al.*, 2019; Fromm, 2020; Xu *et al.*, 2021), GABA
293 was also proposed to regulate autophagy during stress and stress combination (Signorelli
294 *et al.*, 2019; Li *et al.*, 2020; Wang *et al.*, 2021; Balfagón *et al.*, 2022a). Therefore, GABA
295 could play a key role regulating plant responses to individual as well as combined abiotic
296 stresses (Table 1), emerging as a potential metabolic marker to assist breeding programs
297 in the development of climate-resilient crops. Further research is needed to decipher the
298 role of GABA in plant tolerance of crops growing in the field subjected to a multifactorial
299 stress combination.

300 ***Polyamines***

301 Amino acids such as arginine and ornithine are precursors of polyamines, including
302 spermine, spermidine and putrescine. These metabolites are aliphatic compounds induced
303 by several abiotic stresses such as salt, heat, drought and oxidative stress (Alcázar *et al.*,
304 2010; Fu *et al.*, 2014; Glaubitz *et al.*, 2015; Sang *et al.*, 2017; Ebeed *et al.*, 2017;
305 Podlešáková *et al.*, 2019; Jing *et al.*, 2020; Upadhyay *et al.*, 2020). Elevated levels of the
306 main polyamines putrescine, spermidine and spermine have been shown to confer
307 tolerance to different abiotic stresses. For example, a T-DNA mutational analysis of
308 *Oryza sativa* Lysine Decarboxylase-like Protein 1 (OsLDC-like 1) revealed that an
309 increased polyamine biosynthesis enhanced oxidative stress tolerance by preventing the
310 accumulation of ROS (Jang *et al.*, 2012). In addition, spermidine could be involved in the
311 expressions of proteins related to cell rescue and defense and the activation of antioxidant
312 enzymes in tomato seedlings exposed to high temperatures (Sang *et al.*, 2017) and has
313 also been suggested to protect *Arabidopsis* from heat stress by increasing the expression
314 of genes encoding heat shock proteins (Sagor *et al.*, 2012). In addition to high
315 temperatures, drought responses can be modulated by polyamines in wheat plants
316 growing in the field through osmolytes accumulation (Ebeed *et al.*, 2017). Spermidine

317 has been proposed to alleviate salinity damages in tomato plants (Zhang *et al.*, 2015) and
318 in sorghum seedlings (Yin *et al.*, 2016), as well as to reduce chilling injury during low
319 temperature storage of grape berries from vineyards (Harindra Champa *et al.*, 2015). In
320 addition, an Arabidopsis mutant plant (*acl5/spms*), which cannot produce spermine, was
321 hypersensitive to high salt and drought (Jang *et al.*, 2012). Therefore, the role of
322 polyamines in protecting plant cells from a wide range of different stress-associated
323 damages in several plant species has been extensively demonstrated. However, the
324 potential role of polyamines in regulating plant responses to abiotic stress combination
325 remains unclear. It was previously suggested that proline could be involved in the
326 protection of tobacco plants against a combination of drought and heat stress by
327 modulating polyamine biosynthesis (Cvikrová *et al.*, 2013). In addition, spermine could
328 confer tolerance to combined high temperature and drought stress in trifoliolate orange
329 seedlings by modulation of antioxidant capacity (Fu *et al.*, 2014). However, the
330 combination of high light and heat stress repressed the accumulation of putrescine in
331 Arabidopsis plants, suggesting that the role of polyamines under this stress combination
332 might be marginal (Balfagón *et al.*, 2022a).

333 Although different transgenic approaches indicated that polyamines could be essential for
334 abiotic stress tolerance, underlying the possibility of exploiting them as markers to
335 improve plant tolerance (Alcázar *et al.*, 2006), further research is needed to identify the
336 role of polyamines in plants subjected to stress combination.

337

338 **Changes in secondary metabolites in response to stress combination**

339 A wide variety of secondary metabolites in higher plants are synthesized from primary
340 metabolic pathways such as glycolysis, TCA cycle, aliphatic and aromatic amino acids,
341 pentose phosphate pathway and shikimate pathway (Aharoni and Galili, 2011; Erb and
342 Kliebenstein, 2020). The major secondary metabolites groups generated by plants are
343 typically divided into three groups: phenolic compounds (including
344 flavonoids/isoflavonoids, lignins and tannins), terpenes (including terpenoids or
345 isoprenoids), and nitrogen- or sulfur-containing compounds including alkaloids and
346 glucosinolates, respectively (Aharoni and Galili, 2011). In addition to their role in
347 regulating plant defense against herbivores and pathogens, secondary compounds are
348 essential for the acclimation of plants to different abiotic stresses (including temperature,

349 drought, nutrient deficiencies, elevated CO₂, salinity and UV light) by protecting key
350 biomolecules such as proteins and nucleic acids from stress-induced oxidative damage
351 (Ramakrishna and Ravishankar, 2011; Austen *et al.*, 2019; Ahanger *et al.*, 2020; Jan *et*
352 *al.*, 2021). For example, flavonoids are accumulated in response to almost all abiotic
353 stresses (Nakabayashi and Saito, 2015) and their antioxidant function has been
354 experimentally identified under different stress conditions (Hernández *et al.*, 2004;
355 Kusano *et al.*, 2011; Nakabayashi *et al.*, 2014; Nakabayashi and Saito, 2015; Wang *et al.*,
356 2016a). Terpenoids are also involved in plant responses to different abiotic stresses
357 providing antioxidant protection to drought, temperature, light and salt stresses (Schuh *et*
358 *al.*, 1997; Loreto and Delfine, 2000; Munné-Bosch *et al.*, 2001; Vallat *et al.*, 2005; Zhang
359 *et al.*, 2019; Ahanger *et al.*, 2020). Similarly, nitrogen-containing compounds such as
360 alkaloids increase in their levels in response to different stresses and are considered potent
361 mitigators of oxidative damage resulting from stress (Kleinwächter and Selmar, 2015;
362 Zhu *et al.*, 2015; Benjamin *et al.*, 2019; Patel *et al.*, 2020; Yadav *et al.*, 2021). In addition
363 to alkaloids, glucosinolates accumulate in response to different stresses (Ramakrishna and
364 Ravishankar, 2011) to mediate tolerance to, for example, low temperatures (Ljubej *et al.*,
365 2021), high temperatures (Rao *et al.*, 2021), or drought stress (Eom *et al.*, 2018; Salehin
366 *et al.*, 2019). Metabolic changes of *Lonicerae Japonicae Flos* exposed to salt stress
367 included altered patterns of the biosynthesis of phenolic acid, flavonoids, and iridoids
368 (Cai *et al.*, 2020).

369 Besides their protective role during different individual abiotic stresses, several studies
370 provided evidence for the function of secondary metabolites in the response of plants to
371 stress combination. Some reports showed that, depending on the plant species and the
372 type of the abiotic stresses involved, the secondary metabolism response to one of the
373 single stresses prevails over the other under combined stress. For example, Austen *et al.*,
374 (2019) reported that heat responses (inducing the accumulation of isoprene) overcame the
375 effects of elevated CO₂ (suppressing the accumulation of isoprene) under controlled
376 growth conditions of combined heat and elevated CO₂ in a woody plant (*Salix* spp.),
377 indicating that the antioxidant function and capacity to protect cell membrane of isoprene
378 compensates for its energetic cost under this stress combination (Austen *et al.*, 2019). In
379 contrast, phenylpropanoid accumulation in tomato plants under a combination of heat and
380 salinity was more similar to that raised by salinity than that caused by heat. While several
381 compounds belonging to the caffeoylquinic acids family were repressed under heat, they

382 were induced under salinity or the combination of salinity and heat. By contrast, flavonol-
383 related compounds were accumulated under heat, whereas under salinity and salinity
384 combined with heat these metabolites were repressed, correlating with higher oxidative
385 damage (Martinez et al. 2016). In addition, whereas phenylpropanoids were important for
386 plant tolerance to a combination of salinity and heat (Martinez et al. 2016), a key role for
387 tocopherols was found in the tolerance of tomato plants to the combination of light and
388 heat stress (Spicher *et al.*, 2017). Different stress combinations that included heavy metals
389 resulted in different patterns of accumulation of secondary metabolites depending on the
390 plant species (Zhao *et al.*, 2016; Chrysargyris *et al.*, 2019). For example, in *Rhodiola*
391 *rosea* seedlings growing under semi-controlled conditions, heat stress enhanced the
392 negative effects of heavy metals, showing a higher accumulation of secondary
393 metabolites such as phenolic compounds under this stress combination (Zhao *et al.*,
394 2016). The combination of salinity stress and Cu exposure altered the primary metabolic
395 pathways in favor of production of different secondary metabolites including limonene in
396 *Mentha spicata* plants (Chrysargyris *et al.*, 2019). Furthermore, the combination of
397 drought and heat resulted in activation of secondary metabolites that led to tolerance or
398 sensibility to this stress combination depending on the plant tested. For example,
399 tolerance to a combination of drought and heat stress was correlated with elevated levels
400 of flavonoids and phenols in the cotton genotype Zhongmian 23 (Ibrahim *et al.*, 2019),
401 whereas the activation of secondary metabolism was associated to sensitivity to this stress
402 combination in citrus plants (Zandalinas *et al.*, 2016c).

403 Overall, the ability of plants to synthesize a wide range of largely specialized secondary
404 compounds is thought to facilitate their adaptation to a changing environment (Wink,
405 2010), at the expense of their growth (Panda *et al.*, 2021). The fact that plants invest large
406 amounts of energy synthesizing secondary metabolites indicate the importance of these
407 metabolites to plant survival, especially under combined stresses, in which damages tend
408 to be higher compared to those caused by individual stresses. However, the complexity
409 of metabolic networks, that include tens of thousands of plant secondary metabolites
410 (Wink, 2010), and the findings that the production of secondary compounds is specific to
411 the type of stress combination, as well as the plant species involved, render the use of
412 secondary metabolites in breeding efforts to develop combined stress-resilient crops, a
413 challenge that could be difficult to overcome.

414

415 **Changes in hormone levels in response to stress combination**

416 Together with primary and secondary metabolites, hormones can display a stress-specific
417 accumulation pattern in response to different stresses and their combination. Because the
418 role of different hormones under individual abiotic and biotic stresses is too vast to cover
419 here, readers are directed to excellent reviews on the subject (*e.g.*, Shi *et al.*, 2015; Dar *et al.*,
420 *et al.*, 2015; Eremina *et al.*, 2016; Li *et al.*, 2016a, 2021; Pandey *et al.*, 2016; Verma *et al.*,
421 2016; Raja *et al.*, 2017; Shigenaga *et al.*, 2017; Shu *et al.*, 2018; Dermastia, 2019; Wang
422 *et al.*, 2020b; Devireddy *et al.*, 2020b; Yu *et al.*, 2020). The ability of plants to regulate
423 specific and unique hormonal responses when subjected to stress combinations has an
424 important role in plant acclimation (Table 2). One of the main hormones involved in the
425 response of plants to different abiotic stresses and their combination is ABA. It is thought
426 to be involved in the acclimation of *Arabidopsis* to high temperatures combined with
427 salinity (Suzuki *et al.*, 2016), or with drought (Zandalinas *et al.*, 2016a), by modulating
428 gene expression and the accumulation of Ascorbate Peroxidase 1 (APX1) and
429 Multiprotein Bridging Factor 1c (MBF1c), respectively (Suzuki *et al.*, 2016; Zandalinas
430 *et al.*, 2016a). Furthermore, ABA accumulated in response to a combination of salinity
431 and high temperatures in citrus plants (Balfagón *et al.*, 2019b), and in tomato plants
432 subjected to salinity and drought (Xue *et al.*, 2021). The transcription of ABA-response
433 genes such as Responsive to Desiccation 26 (RD26) and ABA Repressor 1 (ABR1) in
434 poplar plants was also elevated under salinity and heat stress combination (Jia *et al.*,
435 2017), further suggesting that ABA could be a key component in signaling pathways
436 involved in plant acclimation to different stress combinations. However, due to the
437 complex (and sometimes opposing) interactions different stresses impose on hormonal
438 signaling, other hormones can participate in plant responses to stress combination (Table
439 2). For example, in contrast to ABA, JA was shown to be involved in plant acclimation
440 to a combination of high light and heat stress by modulating the expression of different
441 JA-response genes (Balfagón *et al.*, 2019a). In citrus plants, SA accumulated under the
442 combination of drought and heat stress (Zandalinas *et al.*, 2016b), and at low temperatures
443 combined with wounding citrus plants accumulated JA (Balfagón *et al.*, 2019b). In
444 contrast, wounded citrus plants subjected to high temperatures did not accumulate JA
445 whereas SA levels increased, suggesting antagonistic interactions between SA and JA
446 under a combination of heat and wounding (Balfagón *et al.*, 2019b). Interestingly, a recent
447 study of signal integration during the combination of high light and heat stress revealed

448 that the interplay between JA and SA could be involved in how an Arabidopsis leaf
449 simultaneously subjected to high light and heat stress initiates systemic signaling and
450 plant acclimation (Zandalinas *et al.*, 2020a).

451 In summary, during stress combination, different hormone interactions specific for
452 particular stresses may collide, making the study of plant responses to stress combination
453 a complex challenge. In order to identify new targets for crop improvements, deciphering
454 the unique crosstalk modes between the different hormones under different stress
455 combinations should be addressed in future studies of plant acclimation to the
456 combination of multiple stresses.

457

458 **Interconnection among primary metabolism, secondary metabolism and hormones** 459 **under stress**

460 The biosynthesis of secondary metabolites and hormones is closely associated with
461 pathways of primary metabolism (Aharoni and Galili, 2011) (Fig. 2). For example,
462 increments in the sugar/amino acid ratio leads to elevation in phenylpropanoid levels and
463 a decline in the concentration of the alkaloid nicotine. In situations where photosynthesis
464 is inhibited, the decrease in carbohydrates is accompanied by a collapse in the absolute
465 levels of carbon- and nitrogen-rich secondary metabolites (Matt *et al.*, 2002). In addition,
466 amino acids represent major precursors for the synthesis of secondary metabolites and
467 some hormones. Examples include the secondary metabolites nicotine, anthocyanins,
468 glucosinolates, and terpenoid indole alkaloids, that are synthesized from proteinogenic
469 amino acids (Wasternack and Strnad, 2019), or the hormones indole-3-acetic acid (IAA)
470 and ET, that are synthesized from tryptophan and methionine, respectively (Yang *et al.*,
471 2020). Other primary metabolites involved in the biosynthesis of secondary metabolites
472 are polyamines. The interaction of polyamines such as putrescine with secondary
473 metabolism is well established in some plants. The enzyme Putrescine N-
474 methyltransferase (PMT) catalyzes the methylation of putrescine, and the product of this
475 reaction (N-methyl putrescine) is required for the synthesis of nicotine, tropane and
476 nortropane alkaloids in Solanaceae and Convolvulaceae plants (Biastoff *et al.*, 2009).
477 Hormones, in turn, can alter both primary and secondary metabolism (Fig. 2). The
478 hormone ABA can trigger changes in the metabolism of sugars and organic acids in guard
479 cells (Jin *et al.*, 2014) and crosstalk between hormones and secondary metabolism was

480 described in response to different stresses. For example, under drought stress, methyl
481 jasmonate (MeJA) and JA elicit the production of secondary metabolites such as
482 alkaloids, taxanes, terpenoids, coumarins, and phenolic phytoalexins in several plant
483 species (Jogawat *et al.*, 2021). Supporting the link between JA and secondary metabolism
484 reprogramming, it was also reported that in tomato plants, constitutive activation of the
485 jasmonate signaling pathway enhanced the production of secondary metabolites (Chen *et*
486 *al.*, 2006) and that the jasmonate-responsive transcriptional regulator ORCA3 controlled
487 the production of terpenoid indole alkaloids (Van Der Fits and Memelink, 2000). In
488 addition to JA, SA together with putrescine can trigger the accumulation of primary and
489 secondary metabolites including sugars, succinate, leucine, and phenolic compounds to
490 protect plants against drought stress (Khan *et al.*, 2019). Other examples of the interaction
491 of hormones and secondary metabolites under abiotic stress conditions were described
492 previously in Arabidopsis plants (Hectors *et al.*, 2012; Salehin *et al.*, 2019). Auxins play
493 a key role in UV acclimation by regulating flavonoid concentration (Hectors *et al.*, 2012)
494 and the auxin-sensitive Aux/IAA repressors IAA5, IAA6, and IAA19 regulate the
495 biosynthesis of aliphatic glucosinolates under drought stress (Salehin *et al.*, 2019).

496

497 **Conclusions and future perspectives**

498 Among the many processes affected by harsh environmental conditions, plant metabolism
499 is perhaps the most influenced, resulting in significant adjustments to the growth,
500 development, and reproduction of plants. The ability of plants to modulate their primary
501 and secondary metabolism in response to different stresses and/or stress combination is
502 key for the reallocation of resources from growth and reproduction to stress acclimation
503 that is crucial for plant survival (Fig. 2). Progress in our understanding of the changes that
504 occur in different metabolic processes in response to stress combination would identify
505 new targets for crop improvement. However, major challenges exist for metabolic studies
506 and engineering projects attempting to alter these processes. These include: (i) the
507 identification of the key metabolic processes occurring under multiple combined stresses;
508 (ii) designing the proper engineering strategy to increase flux of the metabolic processes
509 required to enhance plant acclimation to stress combination; and (iii) the identification of
510 necessary genetic manipulations to implement this strategy. In addition, due to the
511 conflicting nature of some metabolic changes triggered during different stress

512 combinations, together with the different stress intensities impacting plants in the field, it
513 is difficult to predict the specific metabolites that could be good candidates for breeding
514 programs. Perhaps special attention should be given to common metabolites that function
515 during a wide range of single and combined stresses. In this sense, different studies of
516 metabolic changes in plants subjected to different stress combinations have already
517 provided clues regarding key metabolites that could play a key role in the tolerance of
518 plants growing under multiple stress conditions. An example could be GABA, which
519 accumulate in response to different abiotic stresses and their combination in several plant
520 species (Table 1). Interestingly, GABA is not considered a simple metabolite (Bouché
521 and Fromm, 2004; Bown and Shelp, 2016), and it has been suggested to provide a direct
522 link with the metabolic status of plant cells under stress conditions as a bypass of different
523 reactions of the TCA cycle (Fait *et al.*, 2008; Xu *et al.*, 2021). In addition, GABA has
524 recently been proposed to be a stomatal aperture regulatory signal of economic
525 significance, since genetic manipulation of cell-type specific GABA metabolism could
526 reduce water loss by fine-tuning stomatal aperture (Xu *et al.*, 2021). GABA is therefore
527 a good example for a primary metabolite that could be used for the improvement of plant
528 stress resilience to multiple co-occurring environmental stress conditions. Other potential
529 metabolites that may serve as candidates for crop improvement are secondary
530 metabolites. These are known to have a role in protecting plants from oxidative stress
531 caused by different abiotic stresses. Plant varieties containing high amounts of specific
532 secondary metabolites could therefore provide new avenues for the development of crops
533 with high tolerance to multiple abiotic stress combinations. However, due to the multiple
534 functions secondary metabolites have in plant cells (*e.g.*, flavonoids are developmental
535 regulators, but also function in the response of higher plants to a wide range of abiotic
536 stresses as antioxidants; Agati and Tattini, 2010), future research should focus on how
537 secondary metabolites affect plant survival under multiple stresses, and how these effects
538 are linked to plant development and growth. In addition to primary and secondary
539 metabolites, hormones have been reported to play a key role in plant acclimation to
540 several stress combinations (Table 2), making them potential candidates to improve the
541 climate-resilience of crops. Although ABA has been reported as key metabolite for plant
542 acclimation to a wide range of abiotic stress combinations (Suzuki *et al.*, 2016; Zandalinas
543 *et al.*, 2016a; Jia *et al.*, 2017; Balfagón *et al.*, 2019b), its integration with other hormones
544 and/or other signaling pathways including ROS regulatory systems might be different
545 depending on the different stress combinations (Suzuki, 2016; Devireddy *et al.*, 2020b).

546 Identifying essential master regulators that activate different hormone signaling pathways
547 in response to different stress combinations could therefore provide an avenue to tailor
548 responses of plants under multiple abiotic stress conditions. Further studies should be
549 conducted to address this possibility.

550 Finally, it is important to keep in mind that all metabolic studies of stress combination
551 reported to date focused on metabolite changes in plants subjected to two or at most three
552 co-occurring stress conditions. By contrast, no study has revealed how the different
553 metabolic pathways will be affected by a higher number of different abiotic factors (*i.e.*,
554 multifactorial stress combination; Zandalinas *et al.*, 2021*b,a*). Future climate scenarios
555 may include the simultaneous or sequential exposure of crops to high temperature in
556 combination with many other stresses including salinity, drought, flooding, nutrient
557 deficiency, high CO₂ and/or other biotic and abiotic stressors. It was recently reported in
558 *Arabidopsis* that with the increasing number and complexity of multiple stress factors (up
559 to 6 different stress factors) acting simultaneously, plant growth and survival declines,
560 even though the individual level of each stress applied had a negligible effect on plants
561 (Zandalinas *et al.*, 2021*b*). In this study it was also found that the molecular response of
562 *Arabidopsis* to the different multifactorial stress combinations was unique and may
563 involve pathways or metabolites not identified yet (Zandalinas *et al.*, 2021*b*). In addition,
564 biotic stressors can modify plant responses to abiotic stresses and vice versa (Rivero *et*
565 *al.*, 2022 and references therein), making the study of multifactorial stress combination a
566 difficult challenge to overcome. Further studies addressing the identification of potential
567 metabolites acting under multifactorial abiotic/biotic stress combination could therefore
568 be key to mitigating the impact of global warming and climate change on crop
569 productivity.

570

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577

578 **Author contributions**

579 SIZ, DB, AG-C and RM wrote and approved the final version of the manuscript.

580

581 **Conflict of interest**

582 The authors declare no conflict of interest.

583

584 **Data Availability**

585 All data supporting the findings of this study are available within the paper.

References

Abdelrahman M, Burritt D, Tran L. 2018. The use of metabolomic quantitative trait locus mapping and osmotic adjustment traits for the improvement of crop yields under environmental stresses. *Seminars in Cell & Developmental Biology* **83**, 86–94.

Agati G, Tattini M. 2010. Multiple functional roles of flavonoids in photoprotection. *New Phytologist* **186**, 786–793.

Ahanger MA, Bhat JA, Siddiqui MH, Rinklebe J, Ahmad P. 2020. Integration of silicon and secondary metabolites in plants: a significant association in stress tolerance. *Journal of Experimental Botany* **71**, 6758–6774.

Aharoni A, Galili G. 2011. Metabolic engineering of the plant primary–secondary metabolism interface. *Current Opinion in Biotechnology* **22**, 239–244.

Akçay N, Bor M, Karabudak T, Özdemir F, Türkan I. 2012. Contribution of Gamma amino butyric acid (GABA) to salt stress responses of *Nicotiana sylvestris* CMSII mutant and wild type plants. *Journal of Plant Physiology* **169**, 452–458.

Alcázar R, Altabella T, Marco F, Bortolotti C, Reymond M, Koncz C, Carrasco P, Tiburcio AF. 2010. Polyamines: Molecules with regulatory functions in plant abiotic stress tolerance. *Planta* **231**, 1237–1249.

Alcázar R, Marco F, Cuevas JC, Patron M, Ferrando A, Carrasco P, Tiburcio AF, Altabella T. 2006. Involvement of polyamines in plant response to abiotic stress. *Biotechnology Letters* **28**, 1867–1876.

Allan WL, Simpson JP, Clark SM, Shelp BJ. 2008. γ -Hydroxybutyrate accumulation in *Arabidopsis* and tobacco plants is a general response to abiotic stress: putative regulation by redox balance and glyoxylate reductase isoforms. *Journal of Experimental Botany* **59**, 2555–2564.

Amir R. 2010. Current understanding of the factors regulating methionine content in vegetative tissues of higher plants. *Amino Acids* **39**, 917–931.

An Y, Zhang M, Liu G, Han R, Liang Z. 2013. Proline Accumulation in Leaves of *Periploca sepium* via Both Biosynthesis Up-Regulation and Transport during Recovery from Severe Drought. *PLoS One* **8**, e69942.

- Anjum NA, Hasanuzzaman M, Hossain MA, et al.** 2015. Jacks of metal/metalloid chelation trade in plants—an overview. *Frontiers in Plant Science* **6**, 192.
- Austen N, Walker HJ, Lake JA, Phoenix GK, Cameron DD.** 2019. The Regulation of Plant Secondary Metabolism in Response to Abiotic Stress: Interactions Between Heat Shock and Elevated CO₂. *Frontiers in Plant Science* **10**, 1–12.
- Balfagón D, Gómez-Cadenas A, Rambla JL, Granell A, De Ollas C, Mittler R, Zandalinas SI.** 2022a. GABA plays a key role in plant acclimation to a combination of high light and heat stress. *Plant Physiology*, doi: 10.1093/plphys/kiac010.
- Balfagón D, Rambla JL, Granell A, Arbona V, Gómez-Cadenas A.** 2022b. Grafting can improve citrus tolerance to drought and heat stress combination by modifying scion metabolic response. *Environmental and Experimental Botany*, **195**, 104793.
- Balfagón D, Sengupta S, Gómez-Cadenas A, Fritschi FB, Azad R, Mittler R, Zandalinas SI.** 2019a. Jasmonic acid is required for plant acclimation to a combination of high light and heat stress. *Plant Physiology* **181**, 1668–1682.
- Balfagón D, Zandalinas SI, Gómez-Cadenas A.** 2019b. High temperatures change the perspective: Integrating hormonal responses in citrus plants under co-occurring abiotic stress conditions. *Physiologia Plantarum* **165**, 183–197.
- Batista-Silva W, Heinemann B, Rugen N, Nunes-Nesi A, Araújo WL, Braun H, Hildebrandt TM.** 2019. The role of amino acid metabolism during abiotic stress release. *Plant, Cell & Environment* **42**, 1630–1644.
- Benjamin JJ, Lucini L, Jothiramshekar S, Parida A.** 2019. Metabolomic insights into the mechanisms underlying tolerance to salinity in different halophytes. *Plant Physiology and Biochemistry* **135**, 528–545.
- Bernard SM, Habash DZ.** 2009. The importance of cytosolic glutamine synthetase in nitrogen assimilation and recycling. *New Phytologist* **182**, 608–620.
- Biastoff S, Brandt W, Dräger B.** 2009. Putrescine N-methyltransferase--the start for alkaloids. *Phytochemistry* **70**, 1708–1718.
- Bor M, Seekin B, Ozgur R, Yilmaz O, Ozdemir F, Turkan I.** 2009. Comparative effects of drought, salt, heavy metal and heat stresses on gamma-aminobutyric acid levels of sesame (*Sesamum indicum* L.). *Acta Physiologiae Plantarum* **31**, 655–659.

- Bouché N, Fromm H.** 2004. GABA in plants: Just a metabolite? *Trends in Plant Science* **9**, 110–115.
- Bown AW, Shelp BJ.** 2016. Plant GABA: Not Just a Metabolite. *Trends in Plant Science* **21**, 811–813.
- Cai Z, Chen H, Chen J, et al.** 2020. Metabolomics characterizes the metabolic changes of *Lonicerae Japonicae* Flos under different salt stresses. *PLoS One* **15**, e0243111.
- Carmo-Silva AE, Gore MA, Andrade-Sanchez P, French AN, Hunsaker DJ, Salvucci ME.** 2012. Decreased CO₂ availability and inactivation of Rubisco limit photosynthesis in cotton plants under heat and drought stress in the field. *Environmental and Experimental Botany* **83**, 1–11.
- Carrera FP, Noceda C, Maridueña-Zavala MG, Cevallos-Cevallos JM.** 2021. Metabolomics, a Powerful Tool for Understanding Plant Abiotic Stress. *Agronomy* **11**, 824.
- Chaves M, Maroco J, Pereira J.** 2003. Understanding plant responses to drought - from genes to the whole plant. *Functional plant biology* **30**, 239–264.
- Chen H, Jones AD, Howe GA.** 2006. Constitutive activation of the jasmonate signaling pathway enhances the production of secondary metabolites in tomato. *FEBS Letters* **580**, 2540–2546.
- Chrysargyris A, Papakyriakou E, Petropoulos SA, Tzortzakis N.** 2019. The combined and single effect of salinity and copper stress on growth and quality of *Mentha spicata* plants. *Journal of Hazardous Materials* **368**, 584–593.
- Cohen I, Zandalinas SI, Huck C, Fritschi FB, Mittler R.** 2021. Meta-analysis of drought and heat stress combination impact on crop yield and yield components. *Physiologia Plantarum* **171**, 66–76.
- Coutinho ID, Henning LMM, Döpp SA, Nepomuceno A, Moraes LAC, Marcolino-Gomes J, Richter C, Schwalbe H, Colnago LA.** 2018. Flooded soybean metabolomic analysis reveals important primary and secondary metabolites involved in the hypoxia stress response and tolerance. *Environmental and Experimental Botany* **153**, 176–187.

- Cvikrová M, Gemperlová L, Martincová O, Vanková R.** 2013. Effect of drought and combined drought and heat stress on polyamine metabolism in proline-over-producing tobacco plants. *Plant Physiology and Biochemistry* **73**, 7–15.
- Dar TA, Uddin M, Khan MMA, Hakeem KR, Jaleel H.** 2015. Jasmonates counter plant stress: A Review. *Environmental and Experimental Botany* **115**, 49–57.
- Daş ZA, Dimlioğlu G, Bor M, Özdemir F.** 2016. Zinc induced activation of GABA-shunt in tobacco (*Nicotiana tabaccum* L.). *Environmental and Experimental Botany* **122**, 78–84.
- Das A, Rushton PJ, Rohila JS.** 2017. Metabolomic profiling of soybeans (*Glycine max* L.) reveals the importance of sugar and nitrogen metabolism under drought and heat stress. *Plants* **6**, 199–208.
- Dermastia M.** 2019. Plant hormones in phytoplasma infected plants. *Frontiers in Plant Science* **10**, 477.
- Devireddy AR, Arbogast J, Mittler R.** 2020a. Coordinated and rapid whole-plant systemic stomatal responses. *The New Phytologist* **225**, 21–25.
- Devireddy AR, Zandalinas SI, Fichman Y, Mittler R.** 2020b. Integration of reactive oxygen species and hormone signaling during abiotic stress. *The Plant Journal* **105**, 459–476.
- Devireddy AR, Zandalinas SI, Gómez-Cadenas A, Blumwald E, Mittler R.** 2018. Coordinating the overall stomatal response of plants: Rapid leaf-to-leaf communication during light stress. *Science Signaling* **11**, eaam9514.
- Dugasa MT, Cao F, Ibrahim W, Wu F.** 2019. Differences in physiological and biochemical characteristics in response to single and combined drought and salinity stresses between wheat genotypes differing in salt tolerance. *Physiologia Plantarum* **165**, 134–143.
- Dusenge ME, Duarte AG, Way DA.** 2019. Plant carbon metabolism and climate change: elevated CO₂ and temperature impacts on photosynthesis, photorespiration and respiration. *New Phytologist* **221**, 32–49.
- Ebeed HT, Hassan NM, Aljarani AM.** 2017. Exogenous applications of Polyamines modulate drought responses in wheat through osmolytes accumulation, increasing free

polyamine levels and regulation of polyamine biosynthetic genes. *Plant Physiology and Biochemistry* **118**, 438–448.

Eom SH, Baek S-A, Kim JK, Hyun TK. 2018. Transcriptome Analysis in Chinese Cabbage (*Brassica rapa* ssp. *pekinensis*) Provides the Role of Glucosinolate Metabolism in Response to Drought Stress. *Molecules* **23**, 1186.

Erb M, Kliebenstein DJ. 2020. Plant Secondary Metabolites as Defenses, Regulators, and Primary Metabolites: The Blurred Functional Trichotomy. *Plant Physiology* **184**, 39–52.

Eremina M, Rozhon W, Poppenberger B. 2016. Hormonal control of cold stress responses in plants. *Cellular and Molecular Life Sciences* **73**, 797–810.

Fàbregas N, Fernie AR. 2019. The metabolic response to drought. *Journal of Experimental Botany* **70**, 1077–1085.

Fait A, Fromm H, Walter D, Galili G, Fernie AR. 2008. Highway or byway: the metabolic role of the GABA shunt in plants. *Trends in Plant Science* **13**, 14–19.

Fang C, Fernie AR, Luo J. 2019. Exploring the Diversity of Plant Metabolism. *Trends in Plant Science* **24**, 83–98.

Fernie AR, Bachem CWB, Helariutta Y, et al. 2020. Synchronization of developmental, molecular and metabolic aspects of source–sink interactions. *Nature Plants* **6**, 55–66.

Van Der Fits L, Memelink J. 2000. ORCA3, a jasmonate-responsive transcriptional regulator of plant primary and secondary metabolism. *Science* **289**, 295–297.

Flexas J, Bota J, Escalona JM, Sampol B, Medrano H. 2002. Effects of drought on photosynthesis in grapevines under field conditions: an evaluation of stomatal and mesophyll limitations. *Functional Plant Biology* **29**, 461–471.

Flexas J, Bota J, Loreto F, Cornic G, Sharkey TD. 2004. Diffusive and Metabolic Limitations to Photosynthesis under Drought and Salinity in C3 Plants. *Plant Biology* **6**, 269–279.

Fromm H. 2020. GABA signaling in plants: targeting the missing pieces of the puzzle. *Journal of Experimental Botany* **71**, 6238–6245.

- Fu X-Z, Xing F, Wang N-Q, Peng L-Z, Chun C-P, Cao L, Ling L-L, Jiang C-L.** 2014. Exogenous spermine pretreatment confers tolerance to combined high-temperature and drought stress in vitro in trifoliolate orange seedlings via modulation of antioxidative capacity and expression of stress-related genes. *Biotechnology, Biotechnological Equipment* **28**, 192–198.
- Gagneul D, Aïnouche A, Duhazé C, Lugan R, Larher FR, Bouchereau A.** 2007. A Reassessment of the Function of the So-Called Compatible Solutes in the Halophytic Plumbaginaceae *Limonium latifolium*. *Plant Physiology* **144**, 1598–1611.
- von der Gathen P, Kivi R, Wohltmann I, Salawitch RJ, Rex M.** 2021. Climate change favours large seasonal loss of Arctic ozone. *Nature Communications* **12**, 3886.
- Glaubitz U, Erban A, Kopka J, Hinch DK, Zuther E.** 2015. High night temperature strongly impacts TCA cycle, amino acid and polyamine biosynthetic pathways in rice in a sensitivity-dependent manner. *Journal of Experimental Botany* **66**, 6385–6397.
- Gong Q, Li P, Ma S, Rupassara SI, Bohnert HJ.** 2005. Salinity stress adaptation competence in the extremophile *Thellungiella halophila* in comparison with its relative *Arabidopsis thaliana*. *The Plant Journal* **44**, 826–839.
- Harindra Champa WA, Gill MIS, Mahajan BVC, Bedi S.** 2015. Exogenous treatment of spermine to maintain quality and extend postharvest life of table grapes (*Vitis vinifera* L.) cv. Flame Seedless under low temperature storage. *LWT - Food Science and Technology* **60**, 412–419.
- Hectors K, Van Oevelen S, Guisez Y, Prinsen E, Jansen MAK.** 2012. The phytohormone auxin is a component of the regulatory system that controls UV-mediated accumulation of flavonoids and UV-induced morphogenesis. *Physiologia Plantarum* **145**, 594–603.
- Hedrich R, Shabala S.** 2018. Stomata in a saline world. *Current Opinion in Plant Biology* **46**, 87–95.
- Heinemann B, Hildebrandt TM.** 2021. The role of amino acid metabolism in signaling and metabolic adaptation to stress-induced energy deficiency in plants. *Journal of Experimental Botany* **72**, 4634–4645.

- Hemme D, Veyel D, Mühlhaus T, et al.** 2014. Systems-Wide Analysis of Acclimation Responses to Long-Term Heat Stress and Recovery in the Photosynthetic Model Organism *Chlamydomonas reinhardtii*. *The Plant Cell* **26**, 4270–4297.
- Hernández I, Alegre L, Munné-Bosch S.** 2004. Drought-induced changes in flavonoids and other low molecular weight antioxidants in *Cistus clusii* grown under Mediterranean field conditions. *Tree Physiology* **24**, 1303–1311.
- Hildebrandt TM.** 2018. Synthesis versus degradation: directions of amino acid metabolism during *Arabidopsis* abiotic stress response. *Plant Molecular Biology* **98**, 121–135.
- Hossain MS, Persicke M, ElSayed AI, Kalinowski J, Dietz K-J.** 2017. Metabolite profiling at the cellular and subcellular level reveals metabolites associated with salinity tolerance in sugar beet. *Journal of Experimental Botany* **68**, 5961–5976.
- Houtan KS Van, Tanaka KR, Gagné TO, Becker SL.** 2021. The geographic disparity of historical greenhouse emissions and projected climate change. *Science Advances* **7**, eabe4342.
- Ibrahim W, Zhu Y-M, Chen Y, Qiu C-W, Zhu S, Wu F.** 2019. Genotypic differences in leaf secondary metabolism, plant hormones and yield under alone and combined stress of drought and salinity in cotton genotypes. *Physiologia Plantarum* **165**, 343–355.
- Itam M, Mega R, Tadano S, Abdelrahman M, Matsunaga S, Yamasaki Y, Akashi K, Tsujimoto H.** 2020. Metabolic and physiological responses to progressive drought stress in bread wheat. *Scientific Reports* **10**, 1–14.
- Iyer NJ, Tang Y, Mahalingam R.** 2013. Physiological, biochemical and molecular responses to a combination of drought and ozone in *Medicago truncatula*. *Plant, Cell and Environment* **36**, 706–720.
- Jan R, Asaf S, Numan M, Lubna, Kim KM.** 2021. Plant secondary metabolite biosynthesis and transcriptional regulation in response to biotic and abiotic stress conditions. *Agronomy* **11**, 1–31.
- Jang SJ, Wi SJ, Choi YJ, An G, Park KY.** 2012. Increased polyamine biosynthesis enhances stress tolerance by preventing the accumulation of reactive oxygen species: T-

DNA mutational analysis of *Oryza sativa* lysine decarboxylase-like protein 1. *Molecules and Cells* **34**, 251–262.

Jia J, Zhou J, Shi W, Cao X, Luo J, Polle A, Luo Z Bin. 2017. Comparative transcriptomic analysis reveals the roles of overlapping heat-/drought-responsive genes in poplars exposed to high temperature and drought. *Scientific Reports* **7**, 43215.

Jin R, Wang Y, Liu R, Gou J, Chan Z. 2016. Physiological and Metabolic Changes of Purslane (*Portulaca oleracea* L.) in Response to Drought, Heat, and Combined Stresses. *Frontiers in Plant Science* **6**, 1123.

Jin X, Wang RS, Zhu M, Jeon BW, Albert R, Chen S, Assmann SM. 2014. Abscisic Acid-Responsive Guard Cell Metabolomes of *Arabidopsis* Wild-Type and *gpa1* G-Protein Mutants. *The Plant Cell* **25**, 4789–4811.

Jing J, Guo S, Li Y, Li W. 2020. The alleviating effect of exogenous polyamines on heat stress susceptibility of different heat resistant wheat (*Triticum aestivum* L.) varieties. *Scientific Reports* **10**, 7467.

Jogawat A, Yadav B, Chhaya, Lakra N, Singh AK, Narayan OP. 2021. Crosstalk between phytohormones and secondary metabolites in the drought stress tolerance of crop plants: A review. *Physiologia Plantarum* **172**, 1106–1132.

Kang Z, Babar MA, Khan N, Guo J, Khan J, Islam S, Shrestha S, Shahi D. 2019. Comparative metabolomic profiling in the roots and leaves in contrasting genotypes reveals complex mechanisms involved in post-anthesis drought tolerance in wheat. *PLoS One* **14**, e0213502.

Kaplan F, Kopka J, Haskell DW, Zhao W, Schiller KC, Gatzke N, Sung DY, Guy CL. 2004. Exploring the temperature-stress metabolome of *Arabidopsis*. *Plant Physiology* **136**, 4159–4168.

Kempa S, Krasensky J, Dal Santo S, Kopka J, Jonak C. 2008. A Central Role of Abscisic Acid in Stress-Regulated Carbohydrate Metabolism. *PLoS One* **3**, e3935.

Khan N, Bano A, Babar MA. 2019. Metabolic and physiological changes induced by plant growth regulators and plant growth promoting rhizobacteria and their impact on drought tolerance in *Cicer arietinum* L. *PLoS One* **14**, e0213040.

- Kleinwächter M, Selmar D.** 2015. New insights explain that drought stress enhances the quality of spice and medicinal plants: potential applications. *Agronomy for Sustainable Development* **35**, 121–131.
- Köhler IH, Huber SC, Bernacchi CJ, Baxter IR.** 2019. Increased temperatures may safeguard the nutritional quality of crops under future elevated CO₂ concentrations. *The Plant Journal* **97**, 872–886.
- Korn M, Gärtner T, Erban A, Kopka J, Selbig J, Hinch DK.** 2010. Predicting Arabidopsis Freezing Tolerance and Heterosis in Freezing Tolerance from Metabolite Composition. *Molecular Plant* **3**, 224–235.
- Krasensky J, Jonak C.** 2012. Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. *Journal of Experimental Botany* **63**, 1593–1608.
- Kusano M, Tohge T, Fukushima A, et al.** 2011. Metabolomics reveals comprehensive reprogramming involving two independent metabolic responses of Arabidopsis to UV-B light. *The Plant Journal* **67**, 354–369.
- Li C, Li J, Chong K, et al.** 2016a. Toward a Molecular Understanding of Plant Hormone Actions. *Molecular Plant* **9**, 1–3.
- Li Y, Liu C, Sun X, Liu B, Zhang X, Liang W, Huo L, Wang P, Ma F, Li C.** 2020. Overexpression of MdATG18a enhances alkaline tolerance and GABA shunt in apple through increased autophagy under alkaline conditions. *Tree Physiology* **40**, 1509–1519.
- Li Z, Yu J, Peng Y, Huang B.** 2016b. Metabolic pathways regulated by γ -aminobutyric acid (GABA) contributing to heat tolerance in creeping bentgrass (*Agrostis stolonifera*). *Scientific Reports* **6**, 30338.
- Li SM, Zheng HX, Zhang XS, Sui N.** 2021. Cytokinins as central regulators during plant growth and stress response. *Plant Cell Reports* **40**, 271–282.
- Ljubej V, Redovniković IR, Salopek-Sondi B, Smolko A, Roje S, Šamec D.** 2021. Chilling and Freezing Temperature Stress Differently Influence Glucosinolates Content in *Brassica oleracea* var. *acephala*. *Plants* **10**, 1305.
- Lopez-Delacalle M, Camejo DMD, García-Martí M, et al.** 2020. Using tomato recombinant lines to improve plant tolerance to stress combination through a more efficient nitrogen metabolism. *Frontiers in Plant Science* **10**, 1702.

- Lopez-Delacalle M, Camejo D, Garcia-Marti M, Lopez-Ramal MJ, Nortes PA, Martinez V, Rivero RM.** 2021. Deciphering fruit sugar transport and metabolism from tolerant and sensitive tomato plants subjected to simulated field conditions. *Physiologia Plantarum* **173**, 1715–1728.
- Loreto F, Delfine S.** 2000. Emission of Isoprene from Salt-Stressed *Eucalyptus globulus* Leaves. *Plant Physiology* **123**, 1605–1610.
- Lugan R, Niogret M-FF, Leport L, Guégan J-PP, Larher FR, Savouré A, Kopka J, Bouchereau A.** 2010. Metabolome and water homeostasis analysis of *Thellungiella salsuginea* suggests that dehydration tolerance is a key response to osmotic stress in this halophyte. *The Plant Journal* **64**, 215–229.
- Lv W-T, Lin B, Zhang M, Hua X-J.** 2011. Proline Accumulation Is Inhibitory to *Arabidopsis* Seedlings during Heat Stress. *Plant Physiology* **156**, 1921–1933.
- Lwalaba JLW, Zvobgo G, Mwamba TM, et al.** 2020. High accumulation of phenolics and amino acids confers tolerance to the combined stress of cobalt and copper in barley (*Hordeum vulgare*). *Plant Physiology and Biochemistry* **155**, 927–937.
- Maruyama K, Takeda M, Kidokoro S, et al.** 2009. Metabolic pathways involved in cold acclimation identified by integrated analysis of metabolites and transcripts regulated by DREB1A and DREB2A. *Plant Physiology* **150**, 1972–1980.
- Masson-Delmotte V, Zhai P, Pirani A, et al. (Eds.).** 2021. *IPCC 2021: Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. UK: Cambridge University Press.
- Matt P, Krapp A, Haake V, Mock HP, Stitt M.** 2002. Decreased Rubisco activity leads to dramatic changes of nitrate metabolism, amino acid metabolism and the levels of phenylpropanoids and nicotine in tobacco antisense RBCS transformants. *The Plant Journal* **30**, 663–677.
- Mazzucotelli E, Tartari A, Cattivelli L, Forlani G.** 2006. Metabolism of gamma-aminobutyric acid during cold acclimation and freezing and its relationship to frost tolerance in barley and wheat. *Journal of Experimental Botany* **57**, 3755–3766.

- Medina V, Gilbert ME.** 2016. Physiological trade-offs of stomatal closure under high evaporative gradients in field grown soybean. *Functional Plant Biology* **43**, 40–51.
- Mei X, Chen Y, Zhang L, Fu X, Wei Q, Grierson D, Zhou Y, Huang Y, Dong F, Yang Z.** 2016. Dual mechanisms regulating glutamate decarboxylases and accumulation of gamma-aminobutyric acid in tea (*Camellia sinensis*) leaves exposed to multiple stresses. *Scientific Reports* **6**, 23685.
- Mekonnen D, Flügge U, Ludewig F.** 2016. Gamma-aminobutyric acid depletion affects stomata closure and drought tolerance of *Arabidopsis thaliana*. *Plant Science* **245**, 25–34.
- Mittler R.** 2006. Abiotic stress, the field environment and stress combination. *Trends in Plant Science* **11**, 15–19.
- Miyashita Y, Good AG.** 2008. Contribution of the GABA shunt to hypoxia-induced alanine accumulation in roots of *Arabidopsis thaliana*. *Plant & Cell Physiology* **49**, 92–102.
- Moyankova D, Mladenov P, Berkov S, Peshev D, Georgieva D, Djilianov D.** 2014. Metabolic profiling of the resurrection plant *Haberlea rhodopensis* during desiccation and recovery. *Physiologia Plantarum* **152**, 675–687.
- Muchate NS, Rajurkar NS, Suprasanna P, Nikam TD.** 2019. NaCl induced salt adaptive changes and enhanced accumulation of 20-hydroxyecdysone in the in vitro shoot cultures of *Spinacia oleracea* (L.). *Scientific Reports* **9**, 1–10.
- Munné-Bosch S, Mueller M, Schwarz K, Alegre L.** 2001. Diterpenes and antioxidative protection in drought-stressed *Salvia officinalis* plants. *Journal of Plant Physiology* **158**, 1431–1437.
- Mutava RN, Prince SJK, Syed NH, Song L, Valliyodan B, Chen W, Nguyen HT.** 2015. Understanding abiotic stress tolerance mechanisms in soybean: A comparative evaluation of soybean response to drought and flooding stress. *Plant Physiology and Biochemistry* **86**, 109–120.
- Myers SS, Zanobetti A, Kloog I, et al.** 2014. Increasing CO₂ threatens human nutrition. *Nature* **510**, 139–142.
- Nakabayashi R, Saito K.** 2015. Integrated metabolomics for abiotic stress responses in plants. *Current Opinion in Plant Biology* **24**, 10–16.

- Nakabayashi R, Yonekura-Sakakibara K, Urano K, et al.** 2014. Enhancement of oxidative and drought tolerance in Arabidopsis by overaccumulation of antioxidant flavonoids. *The Plant Journal* **77**, 367–379.
- Nayyar H, Kaur R, Kaur S, Singh R.** 2014. γ -Aminobutyric Acid (GABA) Imparts Partial Protection from Heat Stress Injury to Rice Seedlings by Improving Leaf Turgor and Upregulating Osmoprotectants and Antioxidants. *Journal of Plant Growth Regulation* **33**, 408–419.
- Nishizawa A, Yabuta Y, Shigeoka S.** 2008. Galactinol and raffinose constitute a novel function to protect plants from oxidative damage. *Plant Physiology* **147**, 1251–1263.
- Obata T, Fernie AR.** 2012. The use of metabolomics to dissect plant responses to abiotic stresses. *Cellular and Molecular Life Sciences* **69**, 3225–3243.
- Obata T, Witt S, Lisek J, Palacios-Rojas N, Florez-Sarasa I, Yousfi S, Araus JL, Cairns JE, Fernie AR.** 2015. Metabolite profiles of maize leaves in drought, heat, and combined stress field trials reveal the relationship between metabolism and grain yield. *Plant Physiology* **169**, 2665–2683.
- Orsini F, Alnayef M, Bona S, Maggio A, Gianquinto G.** 2012. Low stomatal density and reduced transpiration facilitate strawberry adaptation to salinity. *Environmental and Experimental Botany* **81**, 1–10.
- Panda S, Kazachkova Y, Aharoni A.** 2021. Catch-22 in specialized metabolism: balancing defense and growth. *Journal of Experimental Botany* **72**, 6027–6041.
- Pandey A, Sharma M, Pandey GK.** 2016. Emerging roles of strigolactones in plant responses to stress and development. *Frontiers in Plant Science* **7**, 434.
- Patel MK, Kumar M, Li W, Luo Y, Burritt DJ, Alkan N, Tran L-SP.** 2020. Enhancing Salt Tolerance of Plants: From Metabolic Reprogramming to Exogenous Chemical Treatments and Molecular Approaches. *Cells* **9**, 2492.
- Per TS, Khan NA, Reddy PS, Masood A, Hasanuzzaman M, Khan MIR, Anjum NA.** 2017. Approaches in modulating proline metabolism in plants for salt and drought stress tolerance: Phytohormones, mineral nutrients and transgenics. *Plant Physiology and Biochemistry* **115**, 126–140.

- Pires M V., Júnior AAP, Medeiros DB, et al.** 2016. The influence of alternative pathways of respiration that utilize branched-chain amino acids following water shortage in Arabidopsis. *Plant, Cell & Environment* **39**, 1304–1319.
- Podlešáková K, Ugena L, Spíchal L, Doležal K, De Diego N.** 2019. Phytohormones and polyamines regulate plant stress responses by altering GABA pathway. *New Biotechnology* **48**, 53–65.
- Pommerrenig B, Ludewig F, Cvetkovic J, Trentmann O, Klemens PAW, Neuhaus HE.** 2018. In Concert: Orchestrated Changes in Carbohydrate Homeostasis Are Critical for Plant Abiotic Stress Tolerance. *Plant and Cell Physiology* **59**, 1290-1299.
- Priya M, Sharma L, Kaur R, Bindumadhava H, Nair RM, Siddique KHM, Nayyar H.** 2019. GABA (γ -aminobutyric acid), as a thermo-protectant, to improve the reproductive function of heat-stressed mungbean plants. *Scientific Reports* **9**, 1–14.
- Raja V, Majeed U, Kang H, Andrabi KI, John R.** 2017. Abiotic stress: Interplay between ROS, hormones and MAPKs. *Environmental and Experimental Botany* **137**, 142–157.
- Ramakrishna A, Ravishankar GA.** 2011. Influence of abiotic stress signals on secondary metabolites in plants. *Plant Signaling & Behavior* **6**, 1720.
- Rao SQ, Chen XQ, Wang KH, Zhu ZJ, Yang J, Zhu B.** 2021. Effect of short-term high temperature on the accumulation of glucosinolates in Brassica rapa. *Plant Physiology and Biochemistry* **161**, 222–233.
- Renault H, Roussel V, El Amrani A, Arzel M, Renault D, Bouchereau A, Deleu C.** 2010. The Arabidopsis pop2-1 mutant reveals the involvement of GABA transaminase in salt stress tolerance. *BMC Plant Biology* **10**, 20.
- Rivero RM, Mestre TC, Mittler R, Rubio F, Garcia-Sanchez F, Martinez V.** 2014. The combined effect of salinity and heat reveals a specific physiological, biochemical and molecular response in tomato plants. *Plant, Cell and Environment* **37**, 1059–1073.
- Rivero RM, Mittler R, Blumwald E, Zandalinas SI.** 2022. Developing climate-resilient crops: Improving plant tolerance to stress combination. *The Plant Journal* **109**, 373-389.
- Rizhsky L, Liang H, Mittler R.** 2002. The combined effect of drought stress and heat shock on gene expression in tobacco. *Plant Physiology* **130**, 1143–1151.

- Rizhsky L, Liang H, Shuman J, Shulaev V, Davletova S, Mittler R.** 2004. When defense pathways collide. The response of Arabidopsis to a combination of drought and heat stress. *Plant Physiology* **134**, 1683–1696.
- Sagor GHM, Berberich T, Takahashi Y, Niitsu M, Kusano T.** 2012. The polyamine spermine protects Arabidopsis from heat stress-induced damage by increasing expression of heat shock-related genes. *Transgenic Research* **22**, 595–605.
- Salehin M, Li B, Tang M, Katz E, Song L, Ecker JR, Kliebenstein DJ, Estelle M.** 2019. Auxin-sensitive Aux/IAA proteins mediate drought tolerance in Arabidopsis by regulating glucosinolate levels. *Nature Communications* **10**, 4021.
- Salvatierra A, Pimentel P, Almada R, Hinrichsen P.** 2016. Exogenous GABA application transiently improves the tolerance to root hypoxia on a sensitive genotype of *Prunus* rootstock. *Environmental and Experimental Botany* **125**, 52–66.
- Sami F, Yusuf M, Faizan M, Faraz A, Hayat S.** 2016. Role of sugars under abiotic stress. *Plant Physiology and Biochemistry* **109**, 54–61.
- Sang Q, Shan X, An Y, Shu S, Sun J, Guo S.** 2017. Proteomic Analysis Reveals the Positive Effect of Exogenous Spermidine in Tomato Seedlings' Response to High-temperature Stress. *Frontiers in Plant Science* **8**, 120.
- Schmitz J, Heinrichs L, Scossa F, Fernie AR, Oelze ML, Dietz KJ, Rothbart M, Grimm B, Flügge UI, Häusler RE.** 2014. The essential role of sugar metabolism in the acclimation response of *Arabidopsis thaliana* to high light intensities. *Journal of Experimental Botany* **65**, 1619–1636.
- Scholz SS, Malabarba J, Reichelt M, Heyer M, Ludewig F, Mithöfer A.** 2017. Evidence for GABA-induced systemic GABA accumulation in *Arabidopsis* upon wounding. *Frontiers in Plant Science* **8**, 388.
- Schuh G, Heiden AC, Hoffmann T, Kahl J, Rockel P, Rudolph J, Wildt J.** 1997. Emissions of Volatile Organic Compounds from Sunflower and Beech: Dependence on Temperature and Light Intensity. *Journal of Atmospheric Chemistry* **27**, 291–318.
- Schwachtje J, Whitcomb SJ, Firmino AAP, Zuther E, Hinch DK, Kopka J.** 2019. Induced, imprinted, and primed responses to changing environments: Does metabolism store and process information? *Frontiers in Plant Science* **10**, 106.

- Seifikalhor M, Aliniaefard S, Bernard F, Seif M, Latifi M, Hassani B, Didaran F, Bosacchi M, Rezadoost H, Li T.** 2020. γ -Aminobutyric acid confers cadmium tolerance in maize plants by concerted regulation of polyamine metabolism and antioxidant defense systems. *Scientific Reports* **10**, 3356.
- Seifikalhor M, Aliniaefard S, Hassani B, Niknam V, Lastochkina O.** 2019. Diverse role of γ -aminobutyric acid in dynamic plant cell responses. *Plant Cell Reports* **38**, 847–867.
- Shaar-Moshe L, Blumwald E, Peleg Z.** 2017. Unique physiological and transcriptional shifts under combinations of salinity, drought, and heat. *Plant Physiology* **174**, 421–434.
- Shaar-Moshe L, Hayouka R, Roessner U, Peleg Z.** 2019. Phenotypic and metabolic plasticity shapes life-history strategies under combinations of abiotic stresses. *Plant Direct* **3**, e00113.
- Shang H, Cao S, Yang Z, Cai Y, Zheng Y.** 2011. Effect of exogenous γ -aminobutyric acid treatment on proline accumulation and chilling injury in peach fruit after long-term cold storage. *Journal of Agricultural and Food Chemistry* **59**, 1264–1268.
- Sharma V, Gupta P, Priscilla K, et al.** 2021. Metabolomics Intervention Towards Better Understanding of Plant Traits. *Cells* **10**, 346.
- Shi Y, Ding Y, Yang S.** 2015. Cold Signal Transduction and its Interplay with Phytohormones During Cold Acclimation. *Plant and Cell Physiology* **56**, 7–15.
- Shi SQ, Shi Z, Jiang ZP, Qi LW, Sun XM, Li CX, Liu JF, Xiao WF, Zhang SG.** 2010. Effects of exogenous GABA on gene expression of *Caragana intermedia* roots under NaCl stress: Regulatory roles for H₂O₂ and ethylene production. *Plant, Cell and Environment* **33**, 149–162.
- Shigenaga AM, Berens ML, Tsuda K, Argueso CT.** 2017. Towards engineering of hormonal crosstalk in plant immunity. *Current Opinion in Plant Biology* **38**, 164–172.
- Shu K, Zhou W, Chen F, Luo X, Yang W.** 2018. Abscisic acid and gibberellins antagonistically mediate plant development and abiotic stress responses. *Frontiers in Plant Science* **9**, 416.
- Shulaev V, Cortes D, Miller G, Mittler R.** 2008. Metabolomics for plant stress response. *Physiologia Plantarum* **132**, 199–208.

- Signorelli S, Tarkowski LP, Van den Ende W, Bassham DC.** 2019. Linking Autophagy to Abiotic and Biotic Stress Responses. *Trends in Plant Science* **24**, 413–430.
- Spicher L, Almeida J, Gutbrod K, Pipitone R, Dörmann P, Glauser G, Rossi M, Kessler F.** 2017. Essential role for phytol kinase and tocopherol in tolerance to combined light and temperature stress in tomato. *Journal of Experimental Botany* **68**, 5845–5856.
- Steg L.** 2018. Limiting climate change requires research on climate action. *Nature Climate Change* **8**, 759–761.
- Suzuki N.** 2016. Hormone signaling pathways under stress combinations. *Plant Signaling and Behavior* **11**, 1–5.
- Suzuki N, Bassil E, Hamilton JS, et al.** 2016. ABA is required for plant acclimation to a combination of salt and heat stress. *PLoS One* **11**, e0147625.
- Szabados L, Saviouré A.** 2010. Proline: a multifunctional amino acid. *Trends in Plant Science* **15**, 89–97.
- Thalman M, Santelia D.** 2017. Starch as a determinant of plant fitness under abiotic stress. *New Phytologist* **214**, 943–951.
- Tzin V, Galili G.** 2010. New Insights into the Shikimate and Aromatic Amino Acids Biosynthesis Pathways in Plants. *Molecular Plant* **3**, 956–972.
- Upadhyay RK, Fatima T, Handa AK, Mattoo AK.** 2020. Polyamines and Their Biosynthesis/Catabolism Genes Are Differentially Modulated in Response to Heat Versus Cold Stress in Tomato Leaves (*Solanum lycopersicum* L.). *Cells* **9**, 1749.
- Urano K, Maruyama K, Ogata Y, et al.** 2009. Characterization of the ABA-regulated global responses to dehydration in Arabidopsis by metabolomics. *The Plant Journal* **57**, 1065–1078.
- Usadel B, Bläsing OE, Gibon Y, Poree F, Höhne M, Günter M, Trethewey R, Kamlage B, Poorter H, Stitt M.** 2008. Multilevel genomic analysis of the response of transcripts, enzyme activities and metabolites in Arabidopsis rosettes to a progressive decrease of temperature in the non-freezing range. *Plant, Cell and Environment* **31**, 518–547.

- Vallat A, Gu H, Dorn S.** 2005. How rainfall, relative humidity and temperature influence volatile emissions from apple trees in situ. *Phytochemistry* **66**, 1540–1550.
- Vanková R, Dobrá J, Štorchová H.** 2012. Recovery from drought stress in tobacco: An active process associated with the reversal of senescence in some plant parts and the sacrifice of others. *Plant Signaling & Behavior* **7**, 19–21.
- Verma V, Ravindran P, Kumar PP.** 2016. Plant hormone-mediated regulation of stress responses. *BMC Plant Biology* **16**, 1–10.
- Vijayakumari K, Puthur JT.** 2016. γ -Aminobutyric acid (GABA) priming enhances the osmotic stress tolerance in *Piper nigrum* Linn. plants subjected to PEG-induced stress. *Plant Growth Regulation* **78**, 57–67.
- Vile D, Pervent M, Belluau M, Vasseur F, Bresson J, Muller B, Granier C, Simonneau T.** 2012. Arabidopsis growth under prolonged high temperature and water deficit: Independent or interactive effects? *Plant, Cell and Environment* **35**, 702–718.
- Wang P, Dong Y, Zhu L, Hao Z, Hu L, Hu X, Wang G, Cheng T, Shi J, Chen J.** 2021. The role of γ -aminobutyric acid in aluminum stress tolerance in a woody plant, *Liriodendron chinense* \times *tulipifera*. *Horticulture Research* **8**, 80.
- Wang Y, Gu W, Meng Y, Xie T, Li L, Li J, Wei S.** 2017. γ -Aminobutyric Acid Imparts Partial Protection from Salt Stress Injury to Maize Seedlings by Improving Photosynthesis and Upregulating Osmoprotectants and Antioxidants. *Scientific Reports* **7**, 43609.
- Wang F, Kong W, Wong G, Fu L, Peng R, Li Z, Yao Q.** 2016a. AtMYB12 regulates flavonoids accumulation and abiotic stress tolerance in transgenic *Arabidopsis thaliana*. *Molecular Genetics and Genomics* **291**, 1545–1559.
- Wang L, Ma K-B, Lu Z-G, Ren S-X, Jiang H-R, Cui J-W, Chen G, Teng N-J, Lam H-M, Jin B.** 2020a. Differential physiological, transcriptomic and metabolomic responses of *Arabidopsis* leaves under prolonged warming and heat shock. *BMC Plant Biology* **20**, 86.
- Wang J, Song L, Gong X, Xu J, Li M.** 2020b. Functions of Jasmonic Acid in Plant Regulation and Response to Abiotic Stress. *International Journal of Molecular Sciences* **21**, 1446.

Wang W-S, Zhao X-Q, Li M, Huang L-Y, Xu J-L, Zhang F, Cui Y-R, Fu B-Y, Li Z-K. 2016*b*. Complex molecular mechanisms underlying seedling salt tolerance in rice revealed by comparative transcriptome and metabolomic profiling. *Journal of Experimental Botany* **67**, 405–419.

Wasternack C, Strnad M. 2019. Jasmonates are signals in the biosynthesis of secondary metabolites — Pathways, transcription factors and applied aspects — A brief review. *New Biotechnology* **48**, 1–11.

Wedeking R, Maucourt M, Deborde C, Moing A, Gibon Y, Goldbach HE, Wimmer MA. 2018. ¹H-NMR metabolomic profiling reveals a distinct metabolic recovery response in shoots and roots of temporarily drought-stressed sugar beets. *PLoS One* **13**, e0196102.

Weiszmann J, Fürtauer L, Weckwerth W, Nägele T. 2018. Vacuolar sucrose cleavage prevents limitation of cytosolic carbohydrate metabolism and stabilizes photosynthesis under abiotic stress. *The FEBS Journal* **285**, 4082–4098.

Wink M. 2010. Introduction: Biochemistry, Physiology and Ecological Functions of Secondary Metabolites. In: Wink M, ed. *Biochemistry of Plant Secondary Metabolism: Second Edition*. John Wiley & Sons, Ltd, 1–19.

Wu Q, Su N, Huang X, Cui J, Shabala L, Zhou M, Yu M, Shabala S. 2021. Hypoxia-induced increase in GABA content is essential for restoration of membrane potential and preventing ROS-induced disturbance to ion homeostasis. *Plant Communications* **2**, 100188.

Wulff-Zottele C, Gatzke N, Kopka J, Orellana A, Hoefgen R, Fisahn J, Hesse H. 2010. Photosynthesis and metabolism interact during acclimation of *Arabidopsis thaliana* to high irradiance and sulphur depletion. *Plant, Cell & Environment* **33**, 1974–1988.

Xing SG, Jun YB, Hau ZW, Liang LY. 2007. Higher accumulation of γ -aminobutyric acid induced by salt stress through stimulating the activity of diamine oxidases in *Glycine max* (L.) Merr. roots. *Plant Physiology and Biochemistry* **45**, 560–566.

Xu B, Long Y, Feng X, et al. 2021. GABA signalling modulates stomatal opening to enhance plant water use efficiency and drought resilience. *Nature Communications* **12**, 1952.

- Xue F, Liu W, Cao H, Song L, Ji S, Tong L, Ding R.** 2021. Stomatal conductance of tomato leaves is regulated by both abscisic acid and leaf water potential under combined water and salt stress. *Physiologia Plantarum* **172**, 2070–2078.
- Yadav B, Jogawat A, Rahman MS, Narayan OP.** 2021. Secondary metabolites in the drought stress tolerance of crop plants: A review. *Gene Reports* **23**, 101040.
- Yang Q, Zhao D, Liu Q.** 2020. Connections Between Amino Acid Metabolisms in Plants: Lysine as an Example. *Frontiers in Plant Science* **11**, 928.
- Yin L, Wang S, Tanaka K, Fujihara S, Itai A, Den X, Zhang S.** 2016. Silicon-mediated changes in polyamines participate in silicon-induced salt tolerance in *Sorghum bicolor* L. *Plant, Cell & Environment* **39**, 245–258.
- Yu Z, Duan X, Luo L, Dai S, Ding Z, Xia G.** 2020. How Plant Hormones Mediate Salt Stress Responses. *Trends in Plant Science* **25**, 1117–1130.
- Yu GH, Zou J, Feng J, Peng XB, Wu JY, Wu YL, Palanivelu R, Sun MX.** 2014. Exogenous γ -aminobutyric acid (GABA) affects pollen tube growth via modulating putative Ca^{2+} -permeable membrane channels and is coupled to negative regulation on glutamate decarboxylase. *Journal of Experimental Botany* **65**, 3235–3248.
- Yuan K, Wang C, Zhang C, Huang Y, Wang P, Liu Z.** 2020. Rice grains alleviate cadmium toxicity by expending glutamate and increasing manganese in the cadmium contaminated farmland. *Environmental Pollution* **262**, 114236.
- Zandalinas SI, Balfagón D, Arbona V, Gómez-Cadenas A, Inupakutika MA, Mittler R.** 2016a. ABA is required for the accumulation of APX1 and MBF1c during a combination of water deficit and heat stress. *Journal of Experimental Botany* **67**, 5381–5390.
- Zandalinas SI, Fichman Y, Devireddy AR, Sengupta S, Azad RK, Mittler R.** 2020a. Systemic signaling during abiotic stress combination in plants. *Proceedings of the National Academy of Sciences of the United States of America* **117**, 13810–13820.
- Zandalinas SI, Fritschi FB, Mittler R.** 2020b. Signal transduction networks during stress combination. *Journal of Experimental Botany* **71**, 1734–1741.

Zandalinas SI, Fritschi FB, Mittler R. 2021*a*. Global warming, climate change, and environmental pollution: Recipe for a multifactorial stress combination disaster. *Trends in Plant Science* **26**, 588–599.

Zandalinas SI, Mittler R, Balfagón D, Arbona V, Gómez-Cadenas A, Balfagon D, Arbona V, Gomez-Cadenas A. 2018. Plant adaptations to the combination of drought and high temperatures. *Physiologia Plantarum* **162**, 2–12.

Zandalinas SI, Rivero RM, Martínez V, Gómez-Cadenas A, Arbona V. 2016*b*. Tolerance of citrus plants to the combination of high temperatures and drought is associated to the increase in transpiration modulated by a reduction in abscisic acid levels. *BMC Plant Biology* **16**, 105.

Zandalinas S, Sales C, Beltrán J, Gómez-Cadenas A, Arbona V. 2016*c*. Activation of Secondary Metabolism in Citrus Plants Is Associated to Sensitivity to Combined Drought and High Temperatures. *Frontiers in Plant Science* **7**, 1954.

Zandalinas SI, Sengupta S, Fritschi FB, Azad RK, Nechushtai R, Mittler R. 2021*b*. The impact of multifactorial stress combination on plant growth and survival. *New Phytologist* **230**, 1034–1048.

Zhang J, Deng L, Jiang H, Peng C, Huang C, Zhang M, Zhang X. 2021. The effects of elevated CO₂, elevated O₃, elevated temperature, and drought on plant leaf gas exchanges: a global meta-analysis of experimental studies. *Environmental Science and Pollution Research* **28**, 15274–15289.

Zhang X, Niu M, Silva JAT da, et al. 2019. Identification and functional characterization of three new terpene synthase genes involved in chemical defense and abiotic stresses in *Santalum album*. *BMC Plant Biology* **19**, 115.

Zhang Y, Zhang H, Zou ZR, Liu Y, Hu XH. 2015. Deciphering the protective role of spermidine against saline–alkaline stress at physiological and proteomic levels in tomato. *Phytochemistry* **110**, 13–21.

Zhang H, Zhu J, Gong Z, Zhu J-K. 2022. Abiotic stress responses in plants. *Nature Reviews Genetics* **23**, 104–119.

Zhao YH, Jia X, Wang WK, Liu T, Huang SP, Yang MY. 2016. Growth under elevated air temperature alters secondary metabolites in *Robinia pseudoacacia* L.

seedlings in Cd- and Pb-contaminated soils. *Science of The Total Environment* **565**, 586–594.

Zhou R, Yu X, Ottosen CO, Rosenqvist E, Zhao L, Wang Y, Yu W, Zhao T, Wu Z. 2017. Drought stress had a predominant effect over heat stress on three tomato cultivars subjected to combined stress. *BMC Plant Biology* **17**, 24.

Zhu W, Yang B, Komatsu S, Lu X, Li X, Tian J. 2015. Binary stress induces an increase in indole alkaloid biosynthesis in *Catharanthus roseus*. *Frontiers in Plant Science* **6**, 582.

Zhu M, Zhou M, Shabala L, Shabala S, Zhu M, Zhou M, Shabala L, Shabala S. 2014. Linking osmotic adjustment and stomatal characteristics with salinity stress tolerance in contrasting barley accessions. *Functional Plant Biology* **42**, 252–263.

Zinta G, Abdelgawad H, Peshev D, Weedon JT, Van den Ende W, Nijs I, Janssens IA, Beemster GTS, Asard H. 2018. Dynamics of metabolic responses to periods of combined heat and drought in *Arabidopsis thaliana* under ambient and elevated atmospheric CO₂. *Journal of Experimental Botany* **69**, 2159–2170.

Tables

Table 1. GABA levels are enhanced in different plant species in response to different abiotic stresses and their combination. C letter in references means experiments conducted under controlled conditions; SM letters in references mean experiments conducted under semi-controlled conditions; F letter in references means experiments conducted in the field.

Species	Stress	References
Individual stresses		
<i>Agrostis stolonifera</i>	Heat	^C Li <i>et al.</i> , 2016b
<i>Arabidopsis thaliana</i>	Salinity	^C Kempa <i>et al.</i> , 2008; ^C Renault <i>et al.</i> , 2010; ^C Allan <i>et al.</i> , 2008
	Flooding	^C Allan <i>et al.</i> , 2008
	Cold	^C Allan <i>et al.</i> , 2008
	Heat	^C Allan <i>et al.</i> , 2008
	Drought	^C Mekonnen <i>et al.</i> , 2016; ^C Allan <i>et al.</i> , 2008; ^C Xu <i>et al.</i> , 2021
	Hypoxia	^C Miyashita and Good, 2008; ^C Wu <i>et al.</i> , 2021
	Wounding	^C Scholz <i>et al.</i> , 2017
<i>Camellia sinensis</i>	Anoxia	^C Mei <i>et al.</i> , 2016
<i>Glycine max</i>	Salinity	^C Xing <i>et al.</i> , 2007
<i>Nicotiana sylvestris</i>	Salinity	^C Akçay <i>et al.</i> , 2012
<i>Nicotiana tabacum</i>	Flooding	^C Allan <i>et al.</i> , 2008
	Zinc	^C Daş <i>et al.</i> , 2016
<i>Oryza Sativa</i>	Heat	^C Nayyar <i>et al.</i> , 2014
<i>Piper nigrum</i>	Osmotic stress	^C Vijayakumar and Puthur, 2016
<i>Prunus</i>	Hypoxia	^{SC} Salvatierra <i>et al.</i> , 2016
<i>Sesamum indicum</i>	Osmotic stress	^C Bor <i>et al.</i> , 2008
	Salinity	^C Bor <i>et al.</i> , 2008
	Selenium	^C Bor <i>et al.</i> , 2008
	Heat	^C Bor <i>et al.</i> , 2008
<i>Vigna radiata</i>	Heat	^{SC} Priya <i>et al.</i> , 2019
<i>Zea mays</i>	Salinity	^C Wang <i>et al.</i> , 2017
	Drought	^F Obata <i>et al.</i> , 2015
	Heat	^F Obata <i>et al.</i> , 2015
Combined stresses		
<i>Arabidopsis thaliana</i>	High light + Heat	^C Balfagón <i>et al.</i> , 2022a
<i>Brachypodium distachyon</i>	Salinity + Heat	^C Shaar-Moshe <i>et al.</i> , 2019
	Salinity + Drought	^C Shaar-Moshe <i>et al.</i> , 2019
	Salinity + Drought + Heat	^C Shaar-Moshe <i>et al.</i> , 2019
<i>Camellia sinensis</i>	Anoxia + Wounding	^C Mei <i>et al.</i> , 2016
<i>Citrus sinensis</i>	Drought + Heat	^C Balfagón <i>et al.</i> , 2022b
<i>Zea mays</i>	Drought + Heat	^F Obata <i>et al.</i> , 2015

Table 2. Involvement of different hormones in plant responses to different abiotic stress combinations under controlled conditions.

Hormone	Species	Combined stress	Hormone involvement	References
ABA	<i>Arabidopsis</i>	Salinity + Heat	Regulation of gene expression	Suzuki <i>et al.</i> , 2016
	<i>Citrus</i>	Salinity + Heat	Increased ABA accumulation	Balfagón <i>et al.</i> , 2019b
	<i>Poplar</i>	Salinity + Heat	Increased ABA accumulation	Jia <i>et al.</i> , 2017
	<i>Tomato</i>	Salinity + Drought	Stomatal closure	Xue <i>et al.</i> , 2021
	<i>Arabidopsis</i>	Drought + Heat	Induction of acclimation proteins	Zandalinas <i>et al.</i> , 2016a
JA	<i>Arabidopsis</i>	High light + Heat	Regulation of gene expression	Balfagón <i>et al.</i> , 2019a; Zandalinas <i>et al.</i> , 2020a
	<i>Citrus</i>	Cold + Wounding	Increased JA accumulation	Balfagón <i>et al.</i> , 2019b
SA	<i>Citrus</i>	Drought + Heat	Increased SA accumulation	Zandalinas <i>et al.</i> , 2016b
	<i>Citrus</i>	Heat + Wounding	Increased SA accumulation	Balfagón <i>et al.</i> , 2019b

Figure legends

Fig. 1. Changes in primary metabolism of *Arabidopsis* plants subjected to different individual or combined stresses. Over- or under-accumulation of different sugars, amino acids, TCA cycle-derived metabolites and other metabolites, in response to single abiotic stresses and selected stress combinations, are shown. Yellow squares represent contradictory results depending on different experimental designs. References used: Rizhsky *et al.*, 2004; Kaplan *et al.*, 2004; Nishizawa *et al.*, 2008; Maruyama *et al.*, 2009; Wulff-Zottele *et al.*, 2010; Kusano *et al.*, 2011; Schmitz *et al.*, 2014; Mekonnen *et al.*, 2016; Zinta *et al.*, 2018; Weiszmann *et al.*, 2018; Fàbregas and Fernie, 2019; Balfagón *et al.*, 2022a. Abbreviations: e[CO₂], elevated CO₂; S, sulfur; TCA, tricarboxylic acid.

Fig. 2. Multiple stress factors, including environmental, biotic and/or anthropogenic stresses may simultaneously impact plants in the field. Plants respond by reprogramming their metabolic networks, accumulating specific primary and secondary metabolites, as well as different hormones. The integration of different metabolic and hormonal pathways (dotted arrows) allows plants to adjust their growth and adopt a survival, escape, acclimation and/or defense strategies.

		Individual stresses						Combined stresses				
		Cold	Drought	Oxidative stress	UV-B	High light	Heat	High light + Heat	Drought + Heat	S depletion + High light	Cold + High light	Drought + Heat + e[CO ₂]
Sugars	Galactose		Over-accumulation						Over-accumulation			
	Maltose		Over- or under-accumulation		Over-accumulation	Over-accumulation	Under-accumulation		Over-accumulation	Over-accumulation		
	Galactinol	Over-accumulation	Over-accumulation	Over-accumulation			Over-accumulation					
	Raffinose		Over-accumulation				Over-accumulation					Over-accumulation
	Sucrose		Over- or under-accumulation		Over-accumulation	Over-accumulation			Over-accumulation	Over-accumulation	Over-accumulation	Over-accumulation
	Glucose	Over-accumulation	Over-accumulation			Over-accumulation			Over-accumulation	Over-accumulation	Over-accumulation	Over-accumulation
	Mannose								Over-accumulation			
	Fructose	Over-accumulation	Over-accumulation			Over-accumulation			Over-accumulation	Over-accumulation	Over-accumulation	Over-accumulation
	Trehalose	Over-accumulation	Over-accumulation	Under-accumulation		Over-accumulation			Over-accumulation	Over-accumulation		
	Myo-inositol		Over- or under-accumulation				Over-accumulation	Under-accumulation				
Amino acids	Serine	Over-accumulation										
	Glycine	Over-accumulation		Over-accumulation	Over-accumulation				Over-accumulation			
	Leucine	Over-accumulation							Over-accumulation	Over-accumulation		Over-accumulation
	Valine	Over-accumulation							Over-accumulation	Over-accumulation		Over-accumulation
	Isoleucine	Over-accumulation						Under-accumulation	Over-accumulation	Over-accumulation		Over-accumulation
	Alanine	Over-accumulation							Over-accumulation	Over-accumulation		Under-accumulation
	Asparagine			Over-accumulation	Over-accumulation				Over-accumulation	Under-accumulation		Over-accumulation
	Methionine	Over-accumulation										Over-accumulation
	Lysine	Over-accumulation		Over-accumulation					Over-accumulation	Over-accumulation		Over-accumulation
	Threonine	Over-accumulation					Over-accumulation					Over-accumulation
	Tryptophan	Over-accumulation		Over-accumulation					Over-accumulation	Over-accumulation		Over-accumulation
	Phenylalanine	Over-accumulation		Over-accumulation					Over-accumulation	Over-accumulation		Over-accumulation
	Tyrosine	Over-accumulation							Over-accumulation	Over-accumulation		Over-accumulation
	Glutamate					Over-accumulation		Under-accumulation	Over-accumulation	Over-accumulation		Over-accumulation
	Glutamine								Over-accumulation	Over-accumulation		Under-accumulation
	Arginine	Over-accumulation							Under-accumulation	Over-accumulation		Over-accumulation
	GABA	Over-accumulation	Over-accumulation							Over-accumulation	Over-accumulation	
Proline	Over-accumulation				Over-accumulation			Under-accumulation	Over-accumulation	Over-accumulation	Over-accumulation	
Aspartate	Over-accumulation					Under-accumulation		Under-accumulation	Over-accumulation	Over-accumulation	Over-accumulation	
Cysteine	Over-accumulation							Over-accumulation	Over-accumulation		Over-accumulation	
TCA cycle metabolites	Citrate		Over- or under-accumulation					Under-accumulation	Over-accumulation	Over-accumulation		
	2-Oxoglutarate		Over- or under-accumulation						Over-accumulation	Over-accumulation		
	Succinate		Over- or under-accumulation						Over-accumulation	Over-accumulation		
	Fumarate				Under-accumulation		Over-accumulation		Under-accumulation			
	Malate	Over-accumulation	Over- or under-accumulation		Over-accumulation		Over-accumulation		Under-accumulation	Over-accumulation		
Others	Putrescine				Over-accumulation	Over-accumulation		Under-accumulation		Over-accumulation		
	L-ascorbate		Over- or under-accumulation						Over-accumulation	Over-accumulation		
	Nicotinic acid							Under-accumulation				
	Lactate			Over-accumulation					Under-accumulation			

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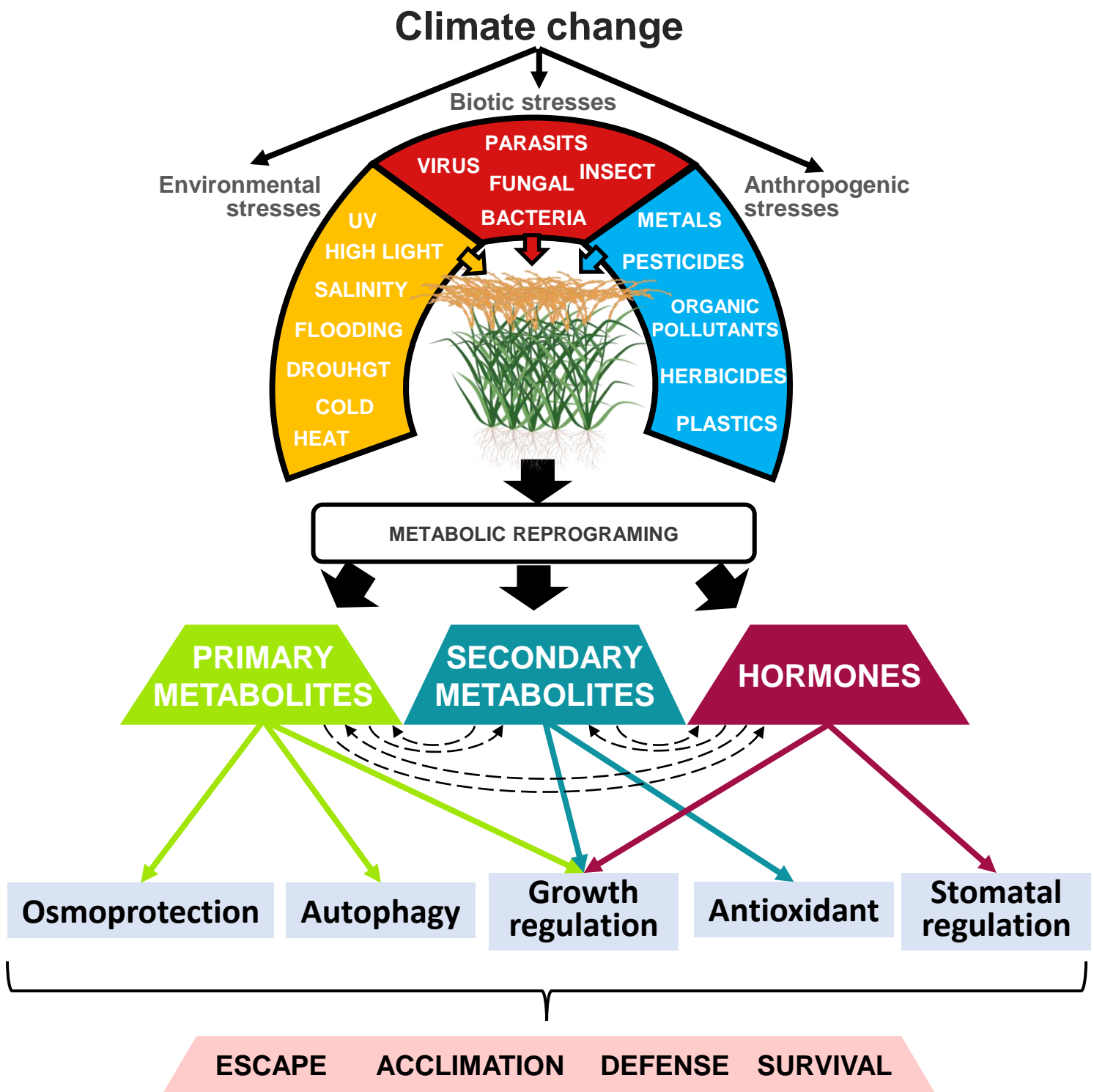


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