Femtosecond digital lensless holographic microscopy to image biological samples

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The use of femtosecond laser radiation in digital lensless holographic microscopy (DLHM) to image biological samples is presented. A mode-locked Ti:Sa laser that emits ultrashort pulses of 12 fs intensity FWHM, with 800 nm mean wavelength, at 75 MHz repetition rate is used as a light source. For comparison purposes, the light from a light-emitting diode is also utilized. For the two-arm DHM, the limited coherence time of this source imposes experimental configurations with very short optical path differences and compensating devices to extend the field of interference [4,6]. Because the single-arm DHM relaxes the conditions over the coherence time, it simplifies the utilization of femtosecond lasers, as was presented by Brunel et al. [5]. In that work, the authors utilized 20 fs plane wave laser radiation to image with a lensless microscope a circular thin film of indium tin oxide 260 μm in diameter. Beyond this type of application, the use of femtosecond laser radiation in DHM may be useful, for instance, to study the spectral response of complex internal structures in biological samples. For these applications, lensless microscopes are the preferred imaging tool because they preserve the temporal pulse duration and do not introduce chromatic aberrations. However, the imaging approach must offer larger magnification and resolution power than that provided by plane wave lensless holographic microscopes [5]. A microscopy architecture that uses no lenses and provides micrometer resolution at multiple wavelengths is digital lensless holographic microscopy (DLHM) [7,8]. DLHM can offer the features needed to image biological specimens with femtosecond laser radiation. In this Letter, the use of femtosecond laser radiation in DLHM and its application to image biological samples is presented. The variation of the pinhole size shows new and unaware effects on the performance of DLHM that are compared with the previously known.

In DLHM, the sample is illuminated by a spherical wavefront with a wavelength \( \lambda \). A digital screen records the amplitude superposition of the portion of the spherical wavefront that is scattered by the specimen, \( U_{\text{scat}} \), with the portion that travels from the point source to the screen with no distortion, \( U_{\text{ref}} \). The former is called the object wave and the latter is the reference wave. The intensity recorded by the digital screen is transferred to a PC for further processing, which includes the retrieving of \( U_{\text{scat}} \). A DLHM setup is illustrated in Fig. 1. For the light sources in this study, we utilize a LED and a mode-locked Ti:Sa laser with 12 fs intensity FWHM and 800 nm mean wavelength at 75 MHz repetition rate and energy per pulse of 4 nJ. For all types of illumination sources, perhaps the most important challenge in order for DLHM to work is the production of the point source. Usually this point source is produced by focusing down light onto a pinhole with diameter \( d_p = \lambda \); namely, the light is shrunk into an area of the order of \( \pi d_p^2/4 \). For instance, for a typical CW He–Ne laser of 10 mW, the power per unit area over the surface of the pinhole is of the order of...
0.7 MW/cm². This figure is enough below the damage threshold of the commercial pinholes, which is of the order of 70 TW/cm². In our application, the peak power over the surface of the pinhole after focusing the ultrashort pulse is of the order of 2.5 TW/cm². This value makes advisable some reduction of the power of the light for safety reasons while enough energy is kept for recording the hologram. Hence, the peak power is reduced to about 1.1 TW/cm² by decreasing the pulse energy with neutral filters.

For comparison of the noise reduction achieved with the use of LEDs [1,2] and that with the fs laser in lensless holographic setups [5], we add to our setup a superbright 655 ± 20 nm LED manufactured by Luxeon. For the fs DLHM as well as for the LED-DLHM, pinholes with 5 and 1 μm diameters are utilized. While the 5 μm pinhole is utilized for enhancing the light throughput, the 1 μm pinhole optimizes the spatial resolution. For both sources, the light from the optical source is focused down onto the pinhole by using an M-40X microscope objective (manufactured by Newport). The microscope objective has a numerical aperture of 0.65, working distance of 0.6 mm, and effective focal length \( F = 4.5 \text{ mm} \). The smallest (largest) spot diameter over the plane of the pinhole in our experiment is 1.2 (2) μm. The holograms are recorded on a CCD camera (manufactured by Balser) with 1024 × 1024 square pixels of 6 μm side. The camera (sample) is located at a distance 17(5) mm from the pinhole.

A section of the head of a drosophila melanogaster fly is utilized as sample to study. For the fs-DLHM, the 800 nm mean wavelength \( \lambda_0 \) of the ultrashort pulse has been chosen as the reconstruction wavelength. The reconstructed holograms from this complex biological specimen for both the fs-DLHM and LED-DLHM are shown in Fig. 2. For all the panels, no coherence noise is visible, as was expected from previous works [1,2,5]. As the panels of the top row are compared with those in the bottom row, one can conclude that the reduction of the coherence noise that is achieved with both sources is comparable; however, there is an important difference in the spatial resolution of the reconstructed holograms. It has been previously reported that in DLHM, the achievable spatial resolution is improved as the pinhole size is reduced, essentially because the spatial coherence of the illuminating source over the plane of the sample is enhanced [1]. From Figs. 2(c) and 2(d) this premise is validated, but for Figs. 2(a) and 2(b), this is no longer true; hence, a further analysis is needed.

To understand why the reconstructed images for the fs-DLHM are blurred, one can regard the broadband illuminating wave as being composed by a bundle of wavelengths. Thus, the recorded in-line hologram is given by the weighted incoherent superposition of multiple monochromatic in-line holograms, each one of the form

\[
I(\mathbf{r}) = \int_{\Delta \nu} 2^{-\Delta \lambda/2} \rho(\lambda) I(\mathbf{r}, \lambda) d\lambda.
\]

In Eq. (1), \( |U_{\text{scat}}(\mathbf{r}, \lambda)|^2 \) is negligible because the samples utilized in DLHM lightly scatter the impinging wavefront. The terms in the square brackets are the so-called twin images [7]. As spherical illumination is utilized, the presence of the twin images does not introduce any nuisance on the reconstructed image, as has been extensively studied in previous works [2,7].

The spectral power \( \rho(\lambda) \) is the corresponding weighting factor such that the recorded in-line hologram in the fs-DLHM is given by

\[
\tilde{I}(\mathbf{r}) = \int_{\lambda_0 - \Delta \nu} 2^{\Delta \lambda/2} \rho(\lambda) I(\mathbf{r}, \lambda) d\lambda.
\]

Because the integration time of the digital screen is much longer than the temporal pulse width, the contrast hologram can be correctly expressed by the intensity superposition given in Eq. (3) [9]. The complex amplitude wave field scattered by the sample \( U_{\text{scat}} \) can be retrieved by computing the diffraction that a reference wave \( U_{\text{ref}} \) undergoes as it illuminates the contrast hologram \( I(\mathbf{r}) \) [7].

As the contrast hologram can be represented by Eq. (3), the information about the individual wavelengths is merged. For that reason, the utilized reconstructing wavelength only changes the scale of the reconstructed image. Details about the numerical implementation of the diffraction process for the case of DLHM can be found elsewhere [10]. The above reasoning leads us to
understand $U_{\text{scat}}$ as the incoherent superposition of multiple diffracted fields from the same specimen, each one with a slightly different scale proportional to $\lambda/\lambda_0$; this superposition results in a blurred reconstructed image, as shown in Figs. 2(a) and 2(b).

The reason why the reconstructed images for the fs-DLHM are nearly invariant with respect to the pinhole size can be supported by considering the DLHM as a wavefront-splitting interferometric microscope. Under this consideration, similar to how it happens in a Young’s-like interferometer, only the spatial coherence controls the performance of the microscope. As the femtosecond laser is essentially a fully spatially coherent source, the different pinhole sizes do not modify the spatial coherence properties over the sample plane; hence, no change in the performance of the microscope is observable. For a spatially partially coherent source such as the LED, the diameter of the area that is almost coherently illuminated $D_{\text{coh}}$ in a plane located at a distance $Z$ from the pinhole is given by the Van Cittert–Zernike theorem, $D_{\text{coh}} = 0.32\lambda_0 Z/d_p$. The above expression indicates that the smaller the pinhole diameter, the larger the spatial coherence, and therefore the better the achievable spatial resolution of the DLHM [1, 2]. This feature is verified in Figs. 2(c) and 2(d). Because the fs laser is a fully spatially coherent source, in the framework of this analysis the performance of the fs-DLHM is not modified essentially by the size of the pinhole, as shown in Figs. 2(a) and 2(b). The blurring of the reconstructed images is therefore due to the superposition of the multiple reconstructed fields with different scaling, as was mentioned above.

The above analysis is supported by measuring the spectrum of the light that illuminates the sample. The spectra that are measured for no pinhole and the pinholes of 5 and 1 $\mu$m are the same, as shown in Fig. 3. The almost identical spectra are consistent with the fact that the quality of the reconstructed images does not change with the size of the pinhole.

In summary, we have shown the use of femtosecond laser radiation in DLHM. With a suitable spectral filtering method, the information gathered with the fs-DLHM for the multiple wavelengths can be unscrambled, providing a new tool for life science imaging, as was recently shown by our group [11].

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