

Cite this: *Analyst*, 2012, **137**, 269

www.rsc.org/analyst

PAPER

## Use of micellar mobile phases for the chromatographic determination of melamine in dietetic supplements

Beatriz Beltrán-Martinavarró, Juan Peris-Vicente, Sergio Marco-Peiró, Josep Esteve-Romero, Maria Rambla-Alegre and Samuel Carda-Broch\*

Received 19th July 2011, Accepted 4th October 2011

DOI: 10.1039/c1an15622e

Melamine is a nitrogen-rich industrial chemical which is occasionally used to increase the apparent protein content of different products destined for human and animal consumption. In this work, a liquid chromatographic procedure that uses micellar mobile phases of sodium dodecyl sulfate (SDS) buffered at pH 3, a C18 column and UV detection is reported for the determination of melamine in dietetic supplements. Samples were reconstituted with a SDS solution and were directly injected, thus avoiding long extraction and experimental procedures. Melamine was eluted in less than 10 min with no interference by other compounds of the matrices. The optimum mobile phase composition was taken by a chemometrical approach that considers the retention factor, efficiency and peak shape. Validation was performed following the indications of the European Commission (Decision 2002/657/EC). The following parameters were considered: linearity ( $0.02\text{--}100\ \mu\text{g mL}^{-1}$ ;  $R^2 = 0.9996$ ), intra- and inter-day precisions ( $<12.4\%$ ), accuracy ( $90.0\text{--}101.3\%$ ), and robustness (less than  $9.8\%$  and  $5.1\%$ , for retention time and peak area, respectively). The limits of detection and quantification were  $9$  and  $20\ \text{ng mL}^{-1}$ , respectively. Recoveries for several spiked samples were in the  $85.8\text{--}114.3\%$  range. These results indicate that the proposed methodology is useful for routine analysis of control quality of infant formula and adult dietetic supplements.

### 1. Introduction

Melamine, or 1,3,5-triazine-2,4,6-triamine (Fig. 1), is an organic compound commonly used in the synthesis of melamine formaldehyde resins for the manufacture of laminates, plastics and glues or adhesives. Melamine can be found at  $\mu\text{g mL}^{-1}$  levels in food and beverages due to migration from melamine-containing resins<sup>1</sup> or as a metabolite product of cyromazine, an insecticide used on animals and crops.<sup>2</sup> When deliberately added to a number of different types of materials in both animal and

human food, it can increase the nitrogen concentration, which suggests a false increase in the protein concentration measured by the Kjeldahl method.<sup>3</sup>

Dietetic supplements are a complex matrix due to the presence of many compounds destined to correct deficiencies in both adult and infant food. In March 2007, a pet food manufacturer alerted the US Food and Drug Administration (US FDA) to animal deaths in the United States which appeared to be linked to certain batches of pet food. Further investigation showed that raw materials (wheat gluten and rice protein), which had been imported from China and used to manufacture pet food,<sup>4</sup> were apparently contaminated with melamine intentionally to increase their total nitrogen concentration and, consequently, the calculated protein content. In September 2008, melamine-tainted milk resulted in nephrolithiasis and renal failure in infants in China. More than 50 000 infants were hospitalised and six infant deaths were confirmed.<sup>5</sup> Melamine could form crystals in combination with cyanuric acid. Very recent Chinese data on the composition of renal calculi (stones) from 15 infants in China who had consumed contaminated infant formula (Chinese Center for Disease Control and Prevention, unpublished data, 2008) confirmed that these stones were composed of uric acid and melamine (in a 1.2 : 1 to 2.1 : 1 molar ratio), and no cyanuric acid was detected.<sup>6</sup>

Due to the proved toxicity of melamine, the safety limit of melamine ingestion has been officially set by the US FDA at

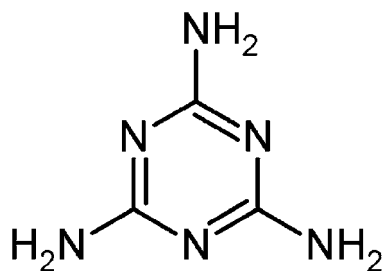


Fig. 1 Melamine structure.

*Química Bioanalítica, QFA, ESTCE, Universitat Jaume I, 12071, Castelló, Spain. E-mail: scarda@qfa.uji.es*

2.5  $\mu\text{g mL}^{-1}$  for adult's food,<sup>7,8</sup> and at 1  $\mu\text{g mL}^{-1}$  for infant formula.<sup>7</sup> Therefore, a reliable method is needed to determine melamine residues in food, particularly in products for children, in order to eliminate the potential threat to human health.

Different methods based on capillary zone electrophoresis with diode array detection,<sup>9</sup> gas chromatography-tandem mass spectrometry,<sup>10,11</sup> high performance liquid chromatography with UV<sup>11,12</sup> or mass spectrometry detection<sup>2,11,13,14</sup> have been developed for the quantification of melamine in a large number of different food and agricultural matrices such as feeds,<sup>11</sup> milk and milk products,<sup>9–11</sup> dairy products and fish feed,<sup>9</sup> cereal flours,<sup>13</sup> chard,<sup>2</sup> animal feed<sup>14</sup> and royal jelly.<sup>12</sup> However, due to the complexity of the matrices, these analytical methodologies involve time-consuming extraction, preconcentrations and purification steps such as molecular imprinting and solid-phase,<sup>11</sup> solid–liquid<sup>2,12–14</sup> and liquid–liquid<sup>9</sup> extraction. Moreover, because of the need for high selectivity, mobile phases are programmed as gradients, which make the analysis of a large amount of samples a difficult task, due to the time of stabilization of the column at the end of each analysis.

The use of surfactant-mediated mobile phases<sup>15</sup> usually simplifies the experimental procedures that involve biological matrices, since their hydrophobic compounds (mainly proteins) are solubilised and become harmless for the chromatographic system. Moreover they are eluted with, or shortly after, the solvent front, and do not interfere with the analytes.<sup>16,17</sup> The introduction of micellar media in the mobile phase and the consequent modification of the stationary phase increase the number of interactions inside the column (between stationary phase, mobile phase, micelles and analyte). A chemometrics approach provides a simulation of the elution conditions, from the data obtained by the study of the analytes in few several mobile phases following an experimental design.<sup>18,19</sup>

In a previous work, the authors analysed melamine in milk samples using a hybrid micellar mobile phase of sodium dodecyl sulfate and propanol.<sup>20</sup> Nowadays, it is desirable to work with non-pollutant mobile phases, which means free of organic solvents or containing low amounts of them. The aim of this work is to perform an easy, fast, accurate and reliable analytical methodology to quantify the level of melamine in dietetic supplements using pure micellar mobile phases containing only the anionic surfactant sodium dodecyl sulfate in the absence of any organic solvent. The analyte has to be resolved from the other compounds of the matrix with sufficient sensitivity to reach the safety levels marked by the FDA. The proposed method was validated following EC indications in terms of selectivity, linearity, sensitivity, repeatability, intermediate precision, robustness and recovery.<sup>21</sup> Since safety limits are not indicated by EU regulations, those established by the US FDA have been considered.<sup>7,8</sup>

## 2. Materials and methods

### 2.1. Chemicals and reagents

Melamine (>99% purity) was purchased from Aldrich (St Louis, MO, USA). Sodium dodecyl sulfate (SDS, >99% purity) was from Merck (Darmstadt, Germany). Ultrapure water (Millipore S.A.S., Molsheim, France) was used throughout to prepare the

mobile phases. Sodium dihydrogenophosphate monohydrate and HCl were obtained from Panreac (Barcelona, Spain). Methanol was bought from J. T. Baker (Deventer, The Netherlands) and 1-propanol and 1-butanol obtained from Scharlab (Barcelona). The characteristics of the studied dietetic supplements are shown in Table 1 (they were all purchased in a local pharmacy).

### 2.2. Instrumentation

The chromatographic system, an Agilent Technologies series 1100 CA, USA, was equipped with an isocratic pump, a degasifier, an autosampler and a DAD (range 190–700 nm). The column used was a Kromasil C18 (150 mm  $\times$  4.6 mm i.d., pore size 100 Å and 5  $\mu\text{m}$  particle size) from Scharlab. The pH of solutions was measured with a potentiometer model GLP 22 Crison (Barcelona), equipped with a combined Ag/AgCl/glass electrode. The analytical balance used was an AX105 Delta-Range (Mettler-Toledo, Switzerland). The mobile phases and the injected solutions were filtered through 0.45  $\mu\text{m}$  nylon membranes (Micron Separations, MA, USA). An ultrasonic bath was used to dissolve the standards (model Ultrasons-H, Selecta, Barcelona).

### 2.3. Mobile phase, sample and standard preparation

The micellar mobile phases were prepared by dissolving the appropriate amount of SDS and dihydrogenophosphate monohydrate in ultrapure water, and then the pH was adjusted to the desired value. Finally, the solution was topped up with ultrapure water to the mark on the volumetric flask, sonicated and filtered.

Samples were spiked by adding the appropriate amount of melamine to 1 mL of liquid dietetic supplement, and by filling up to 10 mL with a solution of 0.05 M SDS at pH 3. Powdered samples were reconstituted by dissolving 1 g in 10 mL of the same micellar solution.

Stock solutions with 1, 100, and 200  $\mu\text{g mL}^{-1}$  of melamine were prepared by dissolving the appropriate amount in methanol.

### 2.4. Chromatographic conditions

Several mobile phases were tested by varying the SDS concentration, while the pH was buffered at 3. Separation was performed by running the mobile phase at 1 mL  $\text{min}^{-1}$  through a Kromasil C18 column thermostatted at 25 °C. The injection volume was 20  $\mu\text{L}$  and the UV wavelength was set at 210 nm. The optimal mobile phase was finally an aqueous solution (0.15 M SDS, pH 3) without any organic solvent. Chromatographic signals were acquired with an Agilent ChemStation (Rev. A.10.01). *Michrom* software was used for processing the chromatographic data.<sup>19</sup>

### 2.5. Method validation

Method validation was performed following the criteria specified by the European Commission Decision 2002/657/EC (2002),<sup>21</sup> and using spiked dietetic supplements, all of which provided similar results.

Linearity and sensitivity were obtained by injecting the analyte at 15 different concentration levels to cover the whole working

**Table 1** Characteristics and recoveries obtained for the analysed dietetic supplements

Commercial name	Supplier	State	Consumer	Spiked conc. / $\mu\text{g mL}^{-1}$	Recovery (%) ( $n = 6$ )
PediaSure Vanilla	Abbot Laboratories Spain (Madrid, Spain)	Powder	Infants	0.050	97.8
				0.100	87.8
				0.150	94.4
				5	114.0
PediaSure Drink Vanilla		Liquid	Infants	0.050	90.9
				0.100	91.9
				0.150	102.1
				5	114.0
PediaSure Chocolate		Powder	Infants	0.050	93.4
				0.100	112.3
				0.150	88.5
				5	109.4
PediaSure Drink Chocolate		Liquid	Infants	0.050	85.3
				0.100	86.2
				0.150	90.5
				5	111.9
Nutrinovex Complex Chocolate	Grupo Farmacéutico de Levante, (Castelló, Spain)	Powder	Adults	0.125	114.1
				0.250	101.6
				0.375	87.9
				5	114.3
Nutrinovex Whey Chocolate		Powder	Adults	0.125	90.4
				0.250	107.6
				0.375	92.5
				5	112.3
Nutrinovex Endurance		Powder	Adults	0.125	87.9
				0.250	113.8
				0.375	100.0
				5	111.8
Nutrinovex Complex Strawberry		Powder	Adults	0.125	106.7
				0.250	85.8
				0.375	87.6
				5	102.5
Optifast Coffee	Nestle Spain, (Espulgues de Llobregat, Spain)	Powder	Adults	0.125	89.7
				0.250	113.9
				0.375	96.4
				5	95.9
Optifast Strawberry		Powder	Adults	0.125	107.0
				0.250	113.5
				0.375	102.1
				5	110.6
Meritene Chocolate		Powder	Adults	0.125	105.0
				0.250	95.2
				0.375	87.5
				5	101.2
Meritene Junior Chocolate		Powder	Infants	0.050	112.0
				0.100	105.2
				0.150	87.2
				5	94.5

range (0.02–100  $\mu\text{g mL}^{-1}$ ). Calibration curves of the spiked dietetic supplement samples were calculated by plotting the peak area of melamine *versus* melamine concentration using the least squares linear regression analysis method. The limit of detection (LOD) was based on the  $3s$  criterion ( $3s/b$ , three times the standard deviation of the lowest concentration solution included in the calibration divided by the slope of the calibration curve using a series of 10 solutions containing a low melamine concentration), and the limit of quantification (LOQ) was selected as the low concentration used in the calibration curve.

Decision limits ( $CC\alpha$ ) and detection capabilities ( $CC\beta$ ) were also calculated.  $CC\alpha$  was calculated by analysing 20 samples spiked with melamine at the LOQ and safety limits (1  $\mu\text{g mL}^{-1}$  for infants and 2.5  $\mu\text{g mL}^{-1}$  for adults) of melamine.  $CC\alpha$  was calculated as the concentration spiked plus 1.64 of the

corresponding standard deviation. To obtain the  $CC\beta$  values, 20 samples were spiked at the calculated  $CC\alpha$  levels, and were analysed. The  $CC\beta$  was calculated as the theoretical value of  $CC\alpha$  previously obtained plus 1.64 times the standard deviation.<sup>21</sup>

Accuracy and precision were also determined by analysing three different concentration levels corresponding to 0.5, 1 and 1.5 times the proposed safety limits. Method robustness was also evaluated.

### 3. Results and discussion

#### 3.1. Mobile phase selection and chromatographic optimization

Melamine is a polar compound ( $\log P_{o/w} = -1.14$ ,  $pK_a = 5.10$ ), which means that adequate analysis times will be obtained using

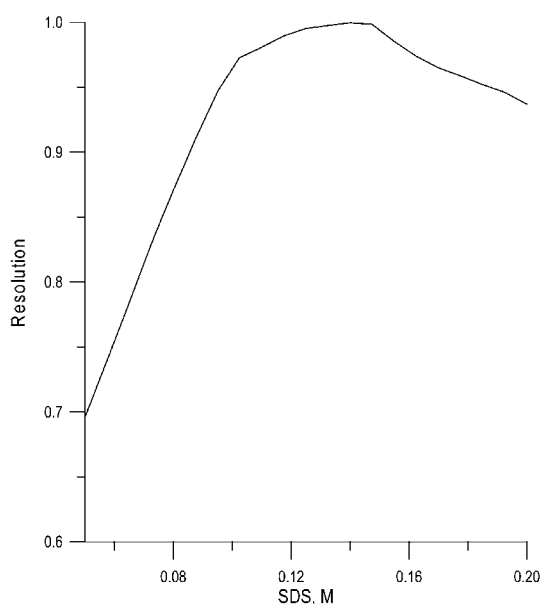
a C18 column and a pure micellar mobile phase. Mobile phases containing 0.05 and 0.1 M SDS were tested, but the retention time was too long. At 0.15 M SDS, the melamine peak eluted at 8.3 min, and was sufficiently separated from the matrix interferences.

The addition of a short-chain alcohol (1-propanol and 1-butanol) can be used in micellar mobile phases to improve efficiency and to reduce the retention time of the compounds.<sup>22</sup> However, in this case, the addition of 1-butanol produces a too fast elution (near the dead volume), and 1-propanol does not provide any significant advantage. Therefore, no alcohol was added to the mobile phase.

An interpretive optimisation strategy was followed to select the best surfactant concentration for the determination of melamine in dietetic supplements. Thus, several mobile phases containing the following SDS (M) concentrations were assayed: 0.05, 0.1, 0.15 and 0.2 M, all buffered at pH 3. This concentration range was selected to avoid excessive retention times or an elution near the void volume. Retention factors ( $k$ ), efficiencies ( $N$ ) and asymmetries ( $B/A$ ) were measured for melamine, as well as for the front of the chromatogram and two unknown matrix compounds, which could overlap with the analyte. These data were processed with the Michrom software.<sup>19</sup> The retention of the compounds was modelled according to:<sup>15</sup>

$$k = \frac{K_{AS}}{1 + K_{AM}[M]} \quad (1)$$

where  $[M]$  is the surfactant concentration;  $K_{AS}$  and  $K_{AM}$  correspond to the equilibrium constants between the solute in pure water and the stationary phase or micelle, respectively. A global resolution criterion based on the equations developed by Lapasio *et al.* was used to predict the chromatograms.<sup>23</sup> This criterion measures the non-overlapped fractions for each individual peak and facilitates the understanding of the information obtained in the optimisation process. Different regions of the variable space



**Fig. 2** Contour map of global resolution for the separation of melamine in dietetic supplements.

are often associated with different critical peak-pairs. Thus, the resolution of a multicomponent mixture requires an analysis that involves all the components in the whole variable space. Inspection of the contour maps of global resolution will allow the robustness of the optimum to be evaluated.

Fig. 2 depicts the resolution diagram. As can be observed, the resolution is poor below a concentration of 0.10 M SDS and above 0.16 M SDS due to several overlapping peaks. Although the best resolution was found at 0.14 M ( $R = 0.9997$ ), the mobile phase selected was 0.15 M SDS, because the resolution was similar, and the retention time of melamine was significantly reduced. Melamine was adequately resolved from the other matrix peaks in less than 10 min using this mobile phase. The chromatographic parameters for melamine were: retention time,  $t_R = 8.3$  min, retention factor,  $k = 8.1$ , efficiency,  $N = 2450$ , and asymmetry factor,  $B/A = 1.1$ .

### 3.2. Method validation

**3.2.1 Selectivity.** Ten blanks of each studied dietetic supplement samples were analysed. Fig. 3 shows the chromatograms obtained from the samples before and after the addition of  $2 \mu\text{g mL}^{-1}$  of melamine. In the blanks, the protein band and a large number of unknown peaks appear both before and after the peak of melamine, but were sufficiently separated to avoid any overlapping.

**3.2.2 Linearity and sensitivity.** To study the variability of the calibration parameters, calibration curves were obtained for a different set of concentration levels (six replicates each, between  $0.02$  and  $100 \mu\text{g mL}^{-1}$ ) for five days over a 2 month period (preparing samples on each occasion).

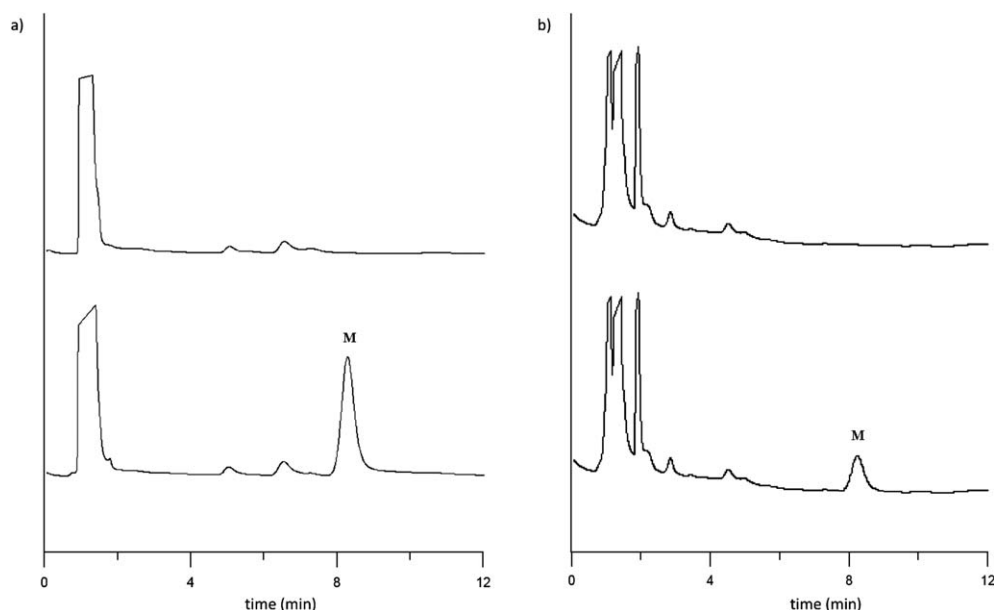
The slope and intercept were determined using the least squares linear regression analysis method. The adjusted equation and regression coefficient ( $R^2$ ), taken as the average of the five calibration curves, were:

$$\text{Peak area} = (4.03 \pm 0.06)[\text{melamine}] - (0.000 \pm 0.003), \\ R^2 = 0.9996$$

The LOQ was the lower level reached in the calibration ( $20 \text{ ng mL}^{-1}$ ), and the LOD was set at  $9 \text{ ng mL}^{-1}$  by the  $3s$  criterion.

**3.2.3 Accuracy and precision.** The intra- and inter-day accuracy and precision of the proposed method were determined by means of recovery experiments. The concentrations chosen were 0.5, 1 and 1.5 times the safety limits proposed by the FDA for infant ( $1 \mu\text{g mL}^{-1}$ ) and adult ( $2.5 \mu\text{g mL}^{-1}$ ) food by considering the 10-fold dilution in the experimental procedure (Section 2.3), and at the three high concentrations ( $2, 5$  and  $10 \mu\text{g mL}^{-1}$ ). Results are shown in Table 2.

The intra-day analysis was determined by injecting the aliquots of these samples six times on the same day. Suitable precision ( $\text{RSD} < 12.4\%$ ) and accuracy ( $90.0\text{--}101.3\%$ ) were found. The inter-day analyses correspond to the average of five measurements of the intra-day values taken over a 3 month period. Precision, expressed as method repeatability, and accuracy were estimated from the recovery experiments ( $n = 5$ ) at



**Fig. 3** Chromatograms of: (a) blank (top) and spiked PediaSure Vanilla powder (infants) (bottom) at  $5 \mu\text{g mL}^{-1}$ , and (b) blank (top) and spiked Nutriben endurance (adults) (bottom) at  $25 \mu\text{g mL}^{-1}$ . Peak identification: M = melamine. Extracts were analysed following the optimised condition methodology.

**Table 2** Intra- and inter-day precisions and accuracy of melamine

Added concentration/ $\mu\text{g mL}^{-1}$	Found <sup>a</sup> (mean $\pm$ SD)/ $\mu\text{g mL}^{-1}$	Accuracy (%)	Intra-day RSD (%)	Found <sup>b</sup> (mean $\pm$ SD)/ $\mu\text{g mL}^{-1}$	Accuracy (%)	Inter-day RSD (%)
0.05	$0.045 \pm 0.005$	90.0	11.1	$0.050 \pm 0.004$	100	8.0
0.1	$0.097 \pm 0.012$	97.0	12.4	$0.101 \pm 0.007$	101	6.9
0.15	$0.138 \pm 0.006$	92.0	4.4	$0.148 \pm 0.007$	98.7	4.7
0.125	$0.116 \pm 0.007$	92.8	6.0	$0.120 \pm 0.012$	96.0	10.0
0.25	$0.25 \pm 0.02$	100.0	8.0	$0.25 \pm 0.02$	100	8.0
0.375	$0.38 \pm 0.03$	101.3	7.9	$0.38 \pm 0.04$	101.3	10.5
2	$1.92 \pm 0.02$	96.0	1.0	$1.90 \pm 0.09$	95.0	4.7
5	$5.04 \pm 0.07$	101	1.4	$4.8 \pm 0.4$	96.0	8.3
10	$9.73 \pm 0.04$	97.3	0.4	$9.5 \pm 0.2$	95.0	2.1

<sup>a</sup>  $n = 6$ . <sup>b</sup>  $n = 5$ .

each concentration level. Excellent precision (RSD < 10.5%) and accuracy (95.0–101.3%) were obtained for all the matrices. The results indicate that the proposed methodology is suitable for the routine analysis of dietetic supplements.

**3.2.4 Robustness.** In order to study the robustness of the method, six replicate injections of a spiked sample at a  $1 \mu\text{g mL}^{-1}$  concentration, with slight changes made to the SDS concentration, pH and flow rate, were examined. The results are shown in Table 3 and indicate that a slight variation in these parameters does not significantly alter either the retention factor of melamine (RSD < 9.3%) or its sensitivity (RSD < 5%).

**3.2.5 Decision limit and detection capability.** The decision limit ( $CC\alpha$ ) indicates the limit at and above which it can be concluded with an error probability of  $\alpha$  that a sample's concentration is over the established limits. The detection capability ( $CC\beta$ ) is the lowest concentration at which the method is able to detect permitted limit concentrations with a statistical

**Table 3** Evaluation of the robustness of the MLC method

Chromatographic parameters	Level	Retention time/min	Area (arbitrary unit)
<i>Concentration SDS (M)</i>			
0.145	-0.005	8.33	3.85
0.150	0	8.73	3.80
0.155	+0.005	7.93	3.98
Mean $\pm$ SD		$8.3 \pm 0.4$	$3.88 \pm 0.09$
RSD (%)		4.8	2.3
<i>pH</i>			
2.9	-0.1	7.9	3.95
3.0	0	8.73	3.80
3.1	+0.1	8.23	4.20
Mean $\pm$ SD		$8.3 \pm 0.6$	$4.0 \pm 0.2$
RSD (%)		7.2	5.0
<i>Flow/mL min<sup>-1</sup></i>			
0.9	-0.1	9.37	3.62
1.0	0	8.73	3.80
1.1	+0.1	7.70	3.87
Mean $\pm$ SD		$8.6 \pm 0.8$	$3.76 \pm 0.12$
RSD (%)		9.3	3.2

certainty of  $1-\beta$ .<sup>21</sup> These parameters allow the assessment of the critical concentrations above which method reliability distinguishes and quantifies a substance by taking into account the variability of the method and the statistical risk of making a wrong decision.<sup>24</sup> Error probabilities  $\alpha$  and  $\beta$  were set at 5%.<sup>21</sup>

In this case, three sets of  $CC\alpha$  and  $CC\beta$  were calculated, and the LOQ and the safety limits for infant formula and adult dietetic supplements were taken as the established limit.<sup>25</sup> The obtained results were: spiking 20 ng mL<sup>-1</sup> (LOQ level):  $CC\alpha = 21$  ng mL<sup>-1</sup> and  $CC\beta = 25$  ng mL<sup>-1</sup>; spiking 0.1  $\mu$ g mL<sup>-1</sup> (safety limit for infants):  $CC\alpha = 0.120$   $\mu$ g mL<sup>-1</sup> and  $CC\beta = 0.140$   $\mu$ g mL<sup>-1</sup>; and spiking 0.25  $\mu$ g mL<sup>-1</sup> (safety limit for adults):  $CC\alpha = 0.280$   $\mu$ g mL<sup>-1</sup> and  $CC\beta = 0.210$   $\mu$ g mL<sup>-1</sup>. The obtained values indicate that melamine can be detected in contaminated samples with the established limits.

**3.2.6 Recovery.** Recovery was determined by analysing the concentration of the spiked samples (of all the studied formulations of dietetic supplements), at several levels and by a comparison between the concentration provided by the proposed method and the known spiked amount. The concentration levels selected for the study were 0.05, 0.1, 0.15 and 5  $\mu$ g mL<sup>-1</sup> for infant products and 0.125, 0.25, 0.375 and 5  $\mu$ g mL<sup>-1</sup> for adult products. Powder samples were prepared and analysed following the previously indicated procedure. The results shown in Table 1 indicate that adequate recovery values (85.8–114.3%) were obtained for all the samples and concentration levels.

#### 4. Conclusions

Micellar liquid chromatography has proved to be a fast, sensitive, and selective technique for the determination of melamine in a wide variety of dietetic supplements for adults and infant formula. The developed method allows the rapid determination of melamine by the direct injection of the reconstituted samples into the chromatographic system, after filtration, thus avoiding long, tedious extractions. The analysis time was under 10 min. Validation was performed according to EC guidelines with satisfactory results in the linearity, selectivity, accuracy, robustness and recovery studies. This method meets the requirements of the “green chemistry” concept since no organic solvent has been used. Besides, it is relatively inexpensive compared to other methods, thus making it a more appealing method. The proposed technique could be recommended as a routine method for the analysis of melamine in dietetic supplements for adults and infant formula.

#### Acknowledgements

This work has been supported by a project from the Spanish Ministry of Education and Science (MEC) CTQ 2007 764473/

BQU and the Foundation Caixa-Castelló, Bancaixa P1-1B2006-12. María Rambla-Alegre also wishes to acknowledge the MEC for the FPU grant.

#### References

- H. Ishiwata, T. Inoue, T. Yamazaki and K. Yoshihira, *J. Assoc. Off. Anal. Chem.*, 1987, **70**, 457–460.
- J. V. Sancho, M. Ibañez, S. Grimalt, O. J. Pozo and F. Hernandez, *Anal. Chim. Acta*, 2005, **530**, 237–243.
- P. G. Wiles, I. K. Gray and R. C. Kissling, *J. AOAC Int.*, 1998, **81**, 620–632.
- FDA, *Interim Melamine and Analogues Swafty/Risk Assessment*, May 25 2007.
- Times on line (From the Times), *Chinese Milk Powder Contaminated with Melamine Sickens 1253 Babies*, <http://www.timesonline.co.uk/tol/news/world/asia/article4758549.ece>, September 16 2008.
- Report of a WHO Expert Meeting in Collaboration with FAO Supported by Health Canada*, Ottawa, 1–4 December 2008.
- L. Zhu, G. Gamez, H. Chen, K. Chingjin and R. Zenobi, *Chem. Commun.*, 2009, 559–561.
- <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/2008/ucm116960.htm>.
- N. Yan, L. Zhou, Z. Zhu and X. Chen, *J. Agric. Food Chem.*, 2009, **57**, 807–811.
- H. Miao, S. Fan, Y. N. Wu, L. Zhang, P. P. Zhou, J. G. Li, H. J. Chen and Y. F. Zhao, *Biomed. Environ. Sci.*, 2009, **22**, 87–94.
- L. He, Y. Su, Y. Zheng, X. Huang, L. Wu, Y. Liu, Z. Zeng and Z. Chen, *J. Chromatogr., A*, 2009, **1216**, 6196–6203.
- J. Zhou, J. Zhao, X. Xue, F. Chen, J. Zhang, Y. Li, L. Wu and L. Chen, *J. Sep. Sci.*, 2010, **33**, 167–173.
- S. Ehling, S. Tefera and I. P. Ho, *Food Addit. Contam.*, 2007, **24**, 1319–1325.
- D. N. Heller and C. B. Nochetto, *Rapid Commun. Mass Spectrom.*, 2008, **22**, 3624–3632.
- A. Berthod and M. C. García-Álvarez-Coque, *Micellar Liquid Chromatography*, Marcel-Dekker, New York, USA, 2000.
- J. Esteve-Romero, E. Ochoa-Aranda, D. Bose, M. Rambla-Alegre, J. Peris-Vicente and A. Martinavarró-Domínguez, *Anal. Bioanal. Chem.*, 2010, **397**, 1557–1561.
- M. Rambla-Alegre, J. Esteve-Romero and S. Carda-Broch, *Anal. Chim. Acta*, 2009, **633**, 250–256.
- M. Rambla-Alegre, J. Peris-Vicente, J. Esteve-Romero and S. Carda-Broch, *Food Chem.*, 2010, **123**, 1294–1302.
- J. R. Torres-Lapasió, *Michrom Software*, Marcel-Dekker, New York, USA, 2000.
- M. Rambla-Alegre, J. Peris-Vicente, S. Marco-Peiró, B. Beltrán-Martinavarró and J. Esteve-Romero, *Talanta*, 2010, **81**, 894–900.
- 2002/657/EC, Commission decision of 12 August 2002 implementing council directive 96/23/EC concerning the performance of analytical methods and the interpretation of results, *Off. J. Eur. Communities: Legis.*, 2002, **221**, 8–62.
- M. Rambla-Alegre, M. T. Gil-Agustí, M. E. Capella-Peiró, S. Carda-Broch and J. Esteve-Romero, *J. Chromatogr., B: Anal. Technol. Biomed. Life Sci.*, 2006, **839**, 89–94.
- J. R. Torres-Lapasió, J. J. Baeza-Baeza and M. C. García-Álvarez-Coque, *Anal. Chem.*, 1997, **69**, 3822–3831.
- E. Verdon, D. Hurtaud-Pessel and P. Sanders, *Accredit. Qual. Assur.*, 2006, **11**, 58–62.
- K. Verschuere, *Handbook of Environmental Data on Organic Chemicals*, Van Nostrand Reinhold Co., New York, 3rd edn, 1996.