

1 **The role of analytical chemistry in exposure science: focus on the aquatic**
2 **environment**

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32 † In Memoriam

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34 **Summary**

35 Exposure science, in its broadest sense, studies the interactions between stressors
36 (chemical, biological, and physical agents) and receptors (e.g. humans and other living
37 organisms, and non-living items like buildings), together with the associated pathways and
38 processes potentially leading to negative effects on human health and the environment. The
39 aquatic environment may contain thousands of compounds, many of them still unknown, that
40 can pose a risk to ecosystems and human health. Due to the unquestionable importance of the
41 aquatic environment, one of the main challenges in the field of exposure science is the
42 comprehensive characterization and evaluation of complex environmental mixtures beyond
43 the classical/priority contaminants to new emerging contaminants.

44 The role of advanced analytical chemistry to identify and quantify potential chemical
45 risks that might cause adverse effects to the aquatic environment, is essential. In this paper,
46 we present the strategies and tools that analytical chemistry has nowadays, focused on
47 chromatography hyphenated to (high-resolution) mass spectrometry because of its relevance
48 in this field. Key issues, such as the application of effect direct analysis to reduce the
49 complexity of the sample, the investigation of the huge number of transformation/degradation
50 products that may be present in the aquatic environment, the analysis of urban wastewater as
51 a source of valuable information on our lifestyle and substances we consumed and/or are
52 exposed to, or the monitoring of drinking water, are discussed in this article. The trends and
53 perspectives for the next few years are also highlighted, when it is expected that new
54 developments and tools will allow a better knowledge of chemical composition in the aquatic
55 environment. This will help regulatory authorities to protect water bodies and to advance
56 towards improved regulations that enable practical and efficient abatements for
57 environmental and public health protection.

58

59 **Keywords:** environmental analytical chemistry, water, exposure science, high resolution mass
60 spectrometry, chromatography, emerging contaminants

61 **1. Introduction**

62 The production and use of chemicals is an indispensable aspect of the current worldwide
63 economy and modern life. A key driver for the increasing use of chemicals is the growth of the
64 global population. This, combined with rapid, albeit uneven, economic and technological
65 development, has led to a considerable increase in global production, consumption of goods
66 and mobility, together with increased demand for food and energy (EEA, 2013). In addition,
67 demographic changes such as the ageing population and the tendency to move from rural
68 areas to urban areas will lead to changes and possible increase in the use and emission
69 patterns of, for instance, pharmaceuticals (Boecking et al., 2012; Tränckner and Koegst, 2010).
70 Worldwide, over 125 million organic compounds are registered in the Chemical Abstract
71 Service (CAS) database. Today, there are more than 100,000 chemicals in the inventory of the
72 European Chemicals Agency (ECHA, 2017). The release of these compounds and resulting risks
73 for human health, the environment and the water cycle, have raised the concern of scientists
74 and policy makers.

75 The increasing awareness of potential risks of organic contaminants for aquatic
76 ecosystems and human health has encouraged scientific and public discussion and political
77 action to introduce measures to improve water quality. As minimization of sources is difficult
78 to achieve, often end-of-pipe solutions such as advanced technologies to remove harmful
79 compounds are implemented (e.g. additional processes in wastewater and drinking water
80 treatment). The conventional processes applied in wastewater treatment plants (WWTPs) are
81 designed to remove the bulk of organic load, measured as biochemical oxygen demand (BOD)
82 or total organic carbon (TOC). However, many organic micropollutants (OMs) are not
83 efficiently removed, and wastewater effluents usually contain amounts of pharmaceuticals,
84 personal care products (PCPs), or illicit drugs, among others, that end up in the aquatic
85 environment (aus der Beek et al., 2016; Bijlsma et al., 2014; Gracia-Lor et al., 2012; Gros et al.,
86 2010; Jelic et al., 2011; J. Liu et al., 2015; Loos et al., 2013; Schymanski et al., 2014b).
87 Additional treatments, such as advanced oxidation or activated carbon, to reduce the input of
88 these contaminants into the aquatic environment are being implemented progressively on
89 certain plants (Eggen et al., 2014). Drinking water treatments, such as nanofiltration or
90 reversed osmosis, can also contribute to the removal of OMs.

91 Chemical pollution has been identified as one of the nine planetary or regional
92 boundaries for which anthropogenic impact needs to be reduced (Rockström et al., 2009b,
93 2009a; Steffen et al., 2015). Despite improvements in regulations and the efforts to reduce the
94 production and use of the most hazardous chemicals, chemical pollution still poses a
95 significant risk to nearly half of the water bodies monitored in Europe (Malaj et al., 2014). Only

96 a few contaminants have been included in the list of priority substances (PS) to be monitored
97 according to the Water Framework Directive (WFD) of the European Commission (EC, 2000).
98 The amended directive for PS (EC, 2013) introduced a Watch List of emerging pollutants or
99 other chemicals for which the available monitoring data are insufficient to assess the
100 environmental risk. Environmental Quality Standards (EQS) have been established for PS (EC,
101 2008) to protect the aquatic environment from adverse effects of chemical substances. The 1st
102 Watch List, launched in 2015, has been recently updated (EC, 2018), and with these measures
103 it is expected that pollution from PS will be progressively reduced. Nevertheless, the number
104 of PS and river basin specific pollutants will still remain very small compared with the number
105 of chemicals released into the environment.

106 A working group headed by the NORMAN network identified that risks and impacts of
107 complex mixtures of chemicals on water quality (Brack et al., 2017) will challenge monitoring,
108 prioritization, assessment and management within the WFD, and an integrated strategy
109 accounting for knowledge gaps is recommended. One of the major challenges is to improve
110 the monitoring and strengthen comprehensive prioritization and risk assessment of complex
111 mixtures. Integrative effect-based tools (EBTs), such as bioassays and biomarkers, are
112 recommended in monitoring water quality. EBTs are particularly useful to bridge the gap
113 between chemical contamination and ecological status; they can cover a broad range of
114 exposure and toxicity mechanisms in diverse organisms, and include effects of non-studied
115 compounds and mixtures, as well as the combined effect of mixtures of compounds (whether
116 or not identified) (Brack et al., 2012).

117 Prioritization, monitoring and assessment under the WFD tend to emphasize regulated,
118 well-known substances, while emerging substances are not adequately addressed (Heiss and
119 Küster, 2015). The present evaluation in European water bodies is mostly based on
120 information on occurrence of target substances versus EQS values. Although some substance
121 classes, such as pesticides, have been widely covered (Moschet et al., 2014), only a small
122 percentage is currently considered in environmental risk assessment (de Zwart et al., 2018).
123 Identifying potential chemical risks in the aquatic environment requires extensive information
124 on the occurrence of a large number of contaminants.

125 Besides the difficulties to identify what to measure and where in the aquatic scenario,
126 the additional challenge is to understand which concentration of an individual substance or
127 combination of substances (mixtures) raises concern in terms of potential risk. To evaluate the
128 potential harmful effects, both hazard and exposure information is needed. The current EU
129 Regulation on chemicals management (Registration, Evaluation, Authorization and restriction
130 of Chemicals (REACH)), created a mandate to deliver adequate exposure information to enable

131 appropriate safety assessments, and foster the safe use and management of chemicals.
132 Exposure information is also required in other European regulatory frameworks, including
133 regulations on plant protection products, biocidal products, construction products, general
134 product safety, classification, labelling and packaging, control of air quality and major-accident
135 hazards. Furthermore, EU strategies promoting moving to a non-toxic environment by 2050,
136 striving towards a circular and bio-based economy, and encouraging green and sustainable
137 chemistry, add additional challenges requiring adequate exposure information. Proper
138 measurement and analyses strategies need to be developed to fulfill the regulatory
139 requirements and to facilitate informed decision-making supporting the EU 2050 goals. To
140 generate and analyze adequate exposure information, the application of excellent exposure
141 science is crucial.

142 Exposure science, in its broadest sense, studies the interaction between stressors
143 (primarily chemical, biological, and physical agents) and receptors (e.g. molecules, cells,
144 organs, humans, other living organisms, and non-living items like buildings), and the associated
145 pathways and processes potentially leading to negative effects on human health and the
146 natural and built environment (Bruinen de Bruin et al., 2018). Therefore, the need for
147 Programs to promote increased generation of exposure information and fit-for-purpose
148 exposure assessment tools, followed by independent assessments and use by risk managers
149 and policy makers, is increasingly important. Analytical chemistry is one of the indispensable
150 disciplines in this field (**Figure 1**). The application of appropriate analytical strategies and tools
151 is essential for a comprehensive characterization of the environment and to understand and
152 evaluate the exposure of the population to emerging chemical risks.

153

154

Insert Figure 1

155

156 Environmental analytical chemistry has several challenges, such as:

- 157 • A huge number of compounds can be present in the environment, and many of them are
158 still unknown. The great diversity in chemical composition of organic contaminants, from
159 non-polar to highly polar compounds, and from low to high volatility, makes the detection
160 and identification of potentially hazardous compounds an analytical challenge. No
161 universal analytical method can be applied to detect and identify all contaminants that
162 may be present in environmental samples. Despite the analytical efforts, a notable
163 number of compounds still requires specific methods and experimental conditions for
164 their determination (e.g. highly volatile compounds, highly polar/ionic compounds, or

165 compounds with distinct polar and apolar functionalities (surfactants)), hampering their
166 inclusion in multi-residue multiclass methods.

- 167 • Most organic contaminants are present at very low concentrations in complex/unknown,
168 ever-changing matrices, and the analytical methodology for their detection, identification
169 and quantification is far from being routinely applicable. It is highly specialized and
170 expensive, and requires the use of sophisticated instrumentation and experienced
171 analysts, particularly when applying wide-scope screening methodologies for suspect and
172 non-target analysis (see later in the manuscript). In addition, valuable techniques
173 commonly used for elucidation of the identity of organic compounds, such as infrared
174 spectroscopy (IR), nuclear magnetic resonance (NMR) and X-Ray, cannot be applied (or
175 have very limited applicability) in the field of environmental analytical chemistry because
176 of their limited sensitivity, making (HR)MS the almost sole analytical technique able to
177 provide information for identification of these compounds.
- 178 • Transformation via biotic or abiotic processes in the environment becomes an additional
179 difficulty, as this multiplies the number of compounds of potential interest, many of which
180 are still unknown. The almost infinite number of transformation products (TPs) in the
181 aqueous environment makes it extraordinary difficult to perform a comprehensive
182 evaluation of their presence and effects on the environment. The lack of occurrence data
183 for TPs imposes a notable limitation in the evaluation of exposure risks.

184 Advanced analytical techniques are required for a comprehensive characterization of the
185 aquatic environment, emphasizing the hyphenation of chromatography with low and high
186 resolution MS, including multidimensional separations, recent chromatographic separation
187 systems and new mass analyzers. High resolution MS (HRMS) presents strong potential for
188 identification of a large number of organic contaminants and elucidation of unknowns derived
189 from accurate-mass full-spectrum data acquired. Many papers have been published in the last
190 years illustrating the excellent performance and tremendous possibilities of HRMS coupled to
191 GC and/or LC to identify a great diversity of pollutants. A detailed review is out of the scope of
192 the present manuscript, but some references can be given as illustrative examples (Andres-
193 Costa et al., 2017; Aceña et al., 2015; Bade et al., 2015a; Bletsou et al., 2015; Díaz et al., 2012;
194 Gosetti et al., 2016; Hernández et al., 2012, 2014, 2018; Hollender et al., 2017; Letzel et al.,
195 2015; Lorenzo et al., 2018; Nurmi et al., 2012; Vergeynst et al., 2015; Wode et al., 2015).

196 People are exposed to many chemicals present in homes, the workplace, and other
197 surroundings through multiple pathways via air, water, food and soil (Wambaugh et al., 2014).
198 Risk assessment depends on two basic elements, exposure and hazardous effects, which are
199 directly related to the presence and concentrations of hazardous compounds in the

200 environment. This is the key of the prioritization schemes that provide the scientific basis for
201 regulatory procedures (Guillén et al., 2012).

202 A strategy based on the maximum environmental concentrations (MEC) has been used
203 to prioritize 500 classical and emerging contaminants in four European river basins, using two
204 indicators, the frequency of exceedance and the extent of exceedance of Predicted No-Effect
205 Concentration (PNEC) (Von der Ohe et al., 2011). Toxicity estimation is an indispensable source
206 for prioritization (e.g. DSSTox database, from EPA), along with exposure (e.g. ExpoCast) and
207 bioactivity predictions (Tox21) (Rager et al., 2016; Tice et al., 2013). However, testing every
208 chemical for every possible health and environmental effect is unrealistic and does not take
209 into account the combined effect of complex mixtures, and therefore prioritization is again
210 necessary.

211 Often, the lack of information, such as sufficient data on the occurrence or effects of a
212 compound, may exclude it from current prioritization procedures (Dulio and Slobodnik, 2015).
213 Thus, many compounds are ignored from the assessment, without any evidence of 'no harm'.
214 The last review of the list of priority pollutants revealed that about 50% of the candidate
215 substances were discarded because of the lack of data (Brack et al., 2017). This illustrates the
216 growing need for monitoring programs to screen for a large number of contaminants including
217 relevant TPs. Many efforts are being made at this challenging task, with substantial
218 improvements in the last few years thanks to the impressive progress in analytical
219 instrumentation and data processing techniques.

220 The **objective of this paper** is to present and discuss the main strategies and tools that
221 modern analytical chemistry can apply in the field of exposure science, with the main focus on
222 the identification of emerging chemical risks in the aquatic environment making use of
223 chromatography hyphenated to (high resolution) mass spectrometry. This article is not
224 conceived as a classical review, and it is directed to scientists from other disciplines related to
225 exposure science. Thus, we do not pursue a detailed/in depth discussion on analytical
226 methods, but to focus on upcoming trends and the tremendous possibilities that make
227 analytical chemistry an indispensable science in this field. Although the work is European
228 oriented, it is applicable on a global scale. We emphasize the relevance of advanced analytical
229 chemistry to provide information on occurrence and behaviour of a huge number of organic
230 contaminants and their TPs in the environment, which is essential in toxicology, environmental
231 risk assessment, exposure science, and in updating regulation.

232

233 **2. Emerging contaminants of recent concern in the aquatic environment**

234 In the last years, many efforts have been made to provide reliable data on occurrence of
235 emerging contaminants in the aquatic environment. A recent review on water analysis
236 (Richardson and Kimura, 2016) mentioned several groups of concern, including artificial
237 sweeteners, nanomaterials, hormones, disinfection by-products, benzotriazoles, siloxanes and
238 microplastics. With the impressive improvement in analytical techniques and analytical
239 strategies, the list of contaminants of emerging concern is notably increasing.

240 In addition to the above mentioned compounds of recent concern, some other families
241 are, or will likely become, a priority in the near future due to its occurrence in the aquatic
242 environment. Among these, can be emphasized pharmaceuticals and personal care products
243 (PPCPs), per- and polyfluorinated alkyl substances (PFAS) and flame retardants (FRs). PPCPs are
244 frequently found in urban wastewater and in receiving surface waters. Pharmaceuticals are
245 often called pseudo persistent substances due to their continuous use and presence in waters.
246 PCPs can contain preservatives, pigments, solvents/oils, nanoparticles, surfactants,
247 pharmaceuticals, UV filters, fragrances, minerals and polymers, and their physico-chemical
248 properties vary widely. The primary pathway of pharmaceuticals, PCPs and their metabolites
249 entering the water cycle is through human usage in or on the body, and excretion or washing
250 off Use of pharmaceuticals in veterinary practice and aquaculture can also lead to emissions
251 (Boxall et al., 2004; Yao et al., 2015).

252 From the first papers reporting pharmaceuticals as environmental contaminants in
253 waters (Daughton, 2004; Daughton and Ternes, 1999; Kolpin et al., 2002), there has been an
254 impressive increase in the number of publications on occurrence of pharmaceuticals in waters
255 around the world (e.g. Alder et al., 2010; Bartelt-Hunt et al., 2009; Campanha et al., 2015;
256 Ghoshdastidar et al., 2015; ter Laak et al., 2014; van Nuijs et al., 2015; Wu et al., 2015). The
257 development of sophisticated analytical techniques has undoubtedly played a key role in their
258 emergence (Richardson and Kimura, 2016). Most of research performed until now is focused
259 on parent pharmaceuticals, and only a limited number of metabolites/TPs is commonly
260 included in analyses (Bletsou et al., 2015; Fatta-Kassinos et al., 2011; Ibáñez et al., 2017; Kern
261 et al., 2010), and their impact on ecology or human health is often lacking (Escher and Fenner,
262 2011). Since some classes of pharmaceuticals have indirect effects such as the development of
263 antibiotic resistance, it is a challenge to determine these effects and the associated risks
264 (Boxall et al., 2012; Richardson, 2017; Singer et al., 2016). The evaluation of the hazards of
265 PCPs is a current priority in regulatory water quality monitoring (Brack et al., 2017, 2012; Loos
266 et al., 2009).

267 The occurrence of pharmaceuticals and PCPs in the aquatic environment is mainly a
268 consequence of the poor removal efficiency of WWTPs for these organic micro-pollutants

269 Therefore, improvements of such efficiency are necessary using additional treatment systems,
270 such as advanced oxidation processes, among others. The comprehensive control of these
271 processes requires monitoring not only the elimination of the parent compound under study,
272 but also the identification of potential degradation/transformation products that can be
273 formed during the process. To this aim, advanced analytical techniques (e.g. LC-HRMS) are
274 required.

275 Per- and polyfluorinated alkyl substances are a prominent group of emerging
276 contaminants. They are persistent, resist degradation and have been linked to adverse health
277 effects (de Voogt and Sáez, 2006). Sources of PFAS in the environment include a variety of
278 industries, firefighting foams, landfills and WWTPs among others (Buck et al., 2011; Busch et
279 al., 2010; Dimzon et al., 2017; Hu et al., 2016; Lau et al., 2007; Lindstrom et al., 2011;
280 Eschauzier et al., 2011). Drinking water and drinking-water based beverages may also play a
281 substantial role in the human exposure to these compounds (Sharma et al., 2016; Sun et al.,
282 2016; Vestergren and Cousins, 2009)

283 Most of the research and analysis of PFAS have focused on the perfluorinated sulfonic
284 acids (PFASs) and perfluorinated carboxylic acids (PFCAs). Several studies (Miyake et al., 2007;
285 Willach et al., 2016) have shown that identified and quantified PFAS in water usually account
286 for less than 50% of the total organic fluorine content of the sample. This can be partly
287 explained by the limited target lists investigated in low resolution MS methods, and lack of
288 standards. Hence, a major part of organic fluorine remains unknown suggesting the occurrence
289 of other fluorinated acids in addition to the known precursors. More than 3000 PFASs have
290 been marketed (Wang et al. 2017). Although in recent years many new anthropogenic PFASs
291 have been detected (Xiao, 2017; Gebbink et al. 2017; Barzen-Hanson et al. 2017) a clear
292 picture of the full range of individual substances is still lacking and reveals the need to
293 investigate in depth the occurrence of fluorinated compounds in the environment Kotthoff
294 and Bücking (2018).

295 The replacements of banned or regulated substances, such as perfluoro-octanesulfonic
296 acid (PFOS) and perfluoro-octanoic acid (PFOA), by per- and polyfluorinated alternatives, such
297 as short chain PFCAs and PFASs and perfluorinated ethers, has led to an increase in their levels
298 in environment (Arp and Slinde, 2018; Schaidler et al. 2017). For example, heptafluoropropoxy
299 propanoic acid (HFPO-DA), a replacement of PFOA in the production process of
300 polytetrafluoroethylene (PTFE), has been reported to occur in rivers close to manufacturing
301 sites where PFOA was previously found, e.g. in the river Rhine (Heydebreck et al., 2015) and
302 Cape Fear river (Sun et al., 2016).

303 While PFAS manufacturers have shifted from the production of long-chained to short-
304 chain (CF<7) PFASs, the shorter chain PFAS are equally persistent in the environment (Ritter,
305 2015) and have higher aqueous solubility than their long-chain homologues. As a result, their
306 removal from water is less efficient and breakthrough in treatment processes used for drinking
307 water production has been demonstrated to occur (Eschauzier et al., 2012; McCleaf et al.,
308 2017). Recently, a suite of persistent halogenated methanesulfonic acids including
309 trifluoromethanesulfonic acid has also been shown to occur in groundwater, surface water and
310 drinking waters (Zahn et al., 2016). These findings highlighted the need for further
311 development of advanced analytical methodologies to unravel the full spectrum of PFAS
312 present in the environment (D'Agostino and Mabury, 2014; Y. Liu et al., 2015; Ruan and Jiang,
313 2017; Strynar et al., 2015, Dimzon et al, 2017).

314 Flame retardants are another group of great interest from the point of view of exposure
315 science. These chemicals are widely applied to materials such as textiles, plastics and
316 electronic products, to delay combustion in case of ignition. Several FRs have been detected in
317 various environmental and food samples (Covaci et al., 2011; Law et al., 2014; Reemtsma et
318 al., 2008; van der Veen and de Boer, 2012). Polybrominated diphenyl ethers (PBDEs) have
319 been extensively used until two PBDEs mixtures (Penta and Octa-mix) were banned, because
320 of their persistence, toxicity and bioaccumulation in the environment and biota (EC, 2003;
321 Environment Canada, 2015, accessed 14.01.19 ; [https://www.epa.gov/assessing-and-
322 managing-chemicals-under-tsca/polybrominated-diphenyl-ethers-pbdes](https://www.epa.gov/assessing-and-managing-chemicals-under-tsca/polybrominated-diphenyl-ethers-pbdes), accessed 14.01.19).
323 As a result of the halt in manufacture of these PBDEs in many countries, other FRs, such as
324 deca-bromodiphenyl ethane (DBDPE), and other brominated and phosphorus FRs (PFRs) have
325 replaced PBDEs (Covaci et al., 2011; Law et al., 2014; van der Veen and de Boer, 2012). This
326 highlights that the continuous modification in law and use of many chemicals around the world
327 forces analytical chemists to address these new families of contaminants.

328 A number of studies have been performed on these "new" FRs and identified them in
329 the environment (Richardson and Kimura, 2016). Tris(chloropropyl)phosphate (TCPP) has been
330 reported as the PFR most frequently found in surface waters (van der Veen and de Boer,
331 2012). In drinking water from eight different Chinese cities, TCPP was also found dominant
332 together with tris(2-butoxyethyl)phosphate (TBEP) and triphenylphosphate (TPP) (Li et al.,
333 2014).

334 Despite some PBDEs have been banned, they will remain in the environment for
335 decades to come, and should therefore be monitored. For example, BDE209 (the major
336 component of the deca-mix PBDEs product) was found dominantly present in sewage sludge
337 samples from several countries (Law et al., 2014; Richardson and Kimura, 2016). BDE209 can

338 also degrade down to lower brominated diphenylethers (Gerecke et al., 2005), which, in
339 contrast to BDE209 can bioaccumulate.

340

341 3. Analytical strategies and tools

342 The analysis of environmental samples involves dealing with high complexity, low
343 concentrations, unknown and sometimes even transient substances, with a broad range of
344 physico-chemical properties. Mass spectrometry, coupled to GC or LC, is commonly the
345 technique of choice to investigate the presence of OMs due to its strong identification
346 potential and because it covers a wide selection of substances compared with other analytical
347 methods. Analytical investigation of OMs can follow different strategies: target, suspect and
348 non-target analysis (Figure 2).

349 In target analysis, the main objective is reliable identification and quantification of
350 known substances for which reference standards are available in house. The analytical
351 methods are optimized to allow highly sensitive and selective quantification of a limited
352 number of substances typically present at sub-ng to µg/L levels. Modern targeted methods are
353 mostly based on GC and LC coupled to (low resolution) tandem mass spectrometry (MS/MS)
354 with ion trap (IT) or triple quadrupole (QqQ) analyzers. GC-MS/MS and LC-MS/MS
355 methodologies have excellent performance in terms of sensitivity and selectivity (and less
356 sample treatment as a consequence of the better sensitivity), and are the most commonly
357 used in recent quantitative target analysis. Analytes of medium/high polarity are more
358 compatible with LC-MS/MS analysis, for example many pesticides, pharmaceuticals, illicit
359 drugs, veterinary drugs (Botitsi et al., 2011; García-Galán et al., 2016; Hernández et al., 2012,
360 2014; van Nuijs et al., 2011). In contrast, both GC-MS and GC-MS/MS, are the techniques of
361 choice for less polar and/or highly volatile contaminants, such as classical priority pollutants
362 (polychlorinated biphenyls (PCBs), organochlorine pesticides or polycyclic aromatic
363 hydrocarbons (PAHs)), as well as siloxanes, PBDEs, and certain pesticides (Barco-Bonilla et al.,
364 2010; Hernández et al., 2013; Law et al., 2014; Pitarch et al., 2010; Portolés et al., 2015).

365 An enrichment step (e.g. solid phase extraction (SPE), solid phase micro extraction
366 (SPME), or solvent extraction) is typically applied prior to MS analysis, and can also result in
367 some sample clean-up. However, the excellent sensitivity of new LC-MS/MS systems allows
368 quantification at low ng/L levels with direct injection of water samples, even prior dilution with
369 ultrapure water (Boix et al., 2015; Causanilles et al., 2016). Identification is achieved through
370 acquisition of at least two MS/MS transitions in the Selected Reaction Monitoring (SRM) mode,
371 and matching of the retention time (RT) and ion-intensity ratios between reference standards
372 and samples (SANTE/11945, 2015). Looking into the (near) future, new technological

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373 developments are expected to allow the expansion of targeted methods to detect, identify and
374 quantify several hundreds of targets accurately, with sensitivity down to pg/L (Hernández et
375 al., 2014; Pinhancos et al., 2011; Vergeynst et al., 2015).

376 Suspect analysis involves looking for compounds expected in the sample on the basis of
377 existing lists of (emerging) substances e.g. NORMAN suspect list exchange (Norman network,
378 2018a) , but for which standards are not available in-house. HRMS is the technique preferred
379 when using this approach, as it allows searching for a large number of suspects based on the
380 accurate-mass full-spectrum acquisition. Detected “suspects” can be tentatively identified with
381 the valuable information provided by HRMS(/MS), RT and even spectral library matching, and
382 can be confirmed retrospectively via reference standard purchase, which is a cost-effective
383 strategy that has led to the increasing popularity of this approach (Hernández et al., 2015a;
384 Moschet et al., 2013; Schymanski et al., 2015; Sjerps et al., 2016). Ion mobility Spectrometry
385 (IMS) adds a new dimension to the chromatographic-TOF MS separations, which notably
386 facilitates the identification process. The collisional cross-section (CCS), measured in IMS,
387 depends on the geometry/shape of the ionized molecule, and is a valuable parameter that
388 may help to solve problematic identifications (e.g. isomers, some analytes in highly complex
389 matrices). Prediction of CCS has been successfully applied to improve identification of suspects
390 and will surely be an efficient tool in suspect and non-target screening in the near future
391 (Bijlsma et al., 2017; Mollerup et al., 2018; Zhou et al., 2016).

392 Suspect screening has huge potential for the identification of emerging contaminants
393 and regulated substances (discussed further below), because it can speed up the evaluation of
394 unknown peaks. Of great value is the use of databases of relevant compounds to be searched
395 in the environmental samples. Considering the huge number of chemicals that are compiled in
396 databases (e.g. the CompTox Chemistry Dashboard from the US EPA (Williams et al., 2017) or
397 in the NORMAN Suspect List Exchange (Norman network, 2018a), a prioritization is needed to
398 facilitate the detection and (tentative) identification of these compounds. Such prioritization
399 can be based on key parameters, such as known or predicted toxicity, knowledge on uses and
400 consumption, persistence in the environment, bioaccumulation, as well as previous data on
401 occurrence (Howard and Muir, 2010).

402 Even after extensive target and suspect screening, many thousands detected
403 components remain unknown (Fuhrer and Zamboni, 2015; Peisl et al., 2018). As identification
404 of unknowns is very time-consuming, these are often prioritized for further elucidation using
405 various methods depending on the research question, e.g. high intensity, specific isotope
406 pattern, (Hoh et al., 2012; Schymanski et al., 2014b), mass defect (Chiaia-Hernandez et al.,
407 2014; Merel et al., 2017; Thurman et al., 2014), linkage analysis for TPs (Schollée et al., 2015)

408 and various statistical measures (Schollée et al., 2016). Experimental design, such as sampling
409 before and after a treatment, along a spatial gradient or over time can prioritize further
410 masses for identification (Hollender et al., 2017). Spectral library searches are also used for
411 non-target identification, especially in GC-MS where extensive standardized libraries are
412 available, but increasingly also in HRMS(/MS). The confirmation of the identity and
413 quantification of the compounds tentatively identified can be performed in a later stage if
414 reference standards are available (Ruff et al., 2015; Schymanski et al., 2014b).

415

416

Insert Figure 2

417

418 In contrast to target analysis, suspect and non-target screening are less mature forms
419 of data exploration and many different possibilities exist, from vendor software through to
420 open access/open source solutions. Non-target strategies differ between the established low
421 resolution GC-MS with electron ionization (EI), for which extensive libraries exist, and HRMS
422 techniques (Schymanski et al., 2015). The NIST library is one of the most extensive collections
423 of GC-EI-MS spectra (>200,000 substances) and retention indices (>83,000 substances)
424 (Vinaixa et al., 2016), and is a valuable resource for the identification of unknowns as two
425 orthogonal pieces of information are available (Schymanski et al., 2014a). If a substance is not
426 in the library, identification e.g. via structure generation remains possible but challenging due
427 to the low-resolution information (Brack et al., 2016; Schymanski et al., 2012). As the exact
428 mass is not generally available, compound database searching has not really established itself
429 for EI-MS.

430 Accurate-mass full-spectrum measurements are of great value for identification
431 purposes, especially when using soft-ionisation techniques, irrespective of whether GC or LC is
432 used. Recently, GC has been coupled to QTOF-MS using an atmospheric pressure chemical
433 ionization (APCI) interface, providing accurate-mass spectra rather similar to those of LC-QTOF
434 MS. The low fragmentation compared to EI, and high abundance of the protonated/molecular
435 ion molecule are of great help to determine mass and often the molecular formula, making GC
436 (APCI)-MS based methods more attractive for screening purposes of organic residues.
437 (Portolés et al., 2014). The complementary use of GC and LC, both coupled to HRMS(/MS),
438 facilitates a comprehensive analysis for known and unknown compounds within a wide range
439 of polarity and volatility. This combination allows to advance towards the desired universal
440 screening of organic contaminants (Hernández et al., 2015b).

441 Some MS/MS spectral libraries are now available and contain a wide range of
442 environmentally-relevant substances, especially NIST17 MSMS (Yang et al., 2017), MassBank

443 (Schulze et al., 2012) and mzCloud (HighChem, accessed 5.30.18); for a more extensive
444 overview see (Brack et al., 2016; Peisl et al., 2018; Vinaixa et al., 2016). If a compound
445 detected is not present in any library, a variety of non-target software approaches exist for
446 identification. The first step usually involves detecting masses of interest (termed peak picking
447 or feature finding), either using commercial (vendor) software or one of the open source
448 approaches such as XCMS (Smith et al., 2006), enviMass (Loos et al., 2016) or MZmine2
449 (Pluskal et al., 2010). Grouping of the resulting masses into components by summarizing
450 isotope and adduct signals is essential to reduce the identification effort (see e.g. (Schymanski
451 et al., 2015, 2014b). This also provides valuable information for calculation of the molecular
452 formula.

453 When fragment information is available, *in silico* approaches can be applied to first find
454 candidates (by searching compound databases on exact mass or formula), then these need to
455 be matched with the experimental spectrum using the structural information provided by the
456 fragments (Dührkop et al., 2015; Ruttkies et al., 2016; Schymanski et al., 2017). The
457 combination of suspect screening with spectral libraries, and *in silico* methods, shows huge
458 potential for rapid and comprehensive screening of emerging contaminants. While the *in silico*
459 methods are improving greatly, it is still vital that tentative structures are confirmed with
460 reference standards before a confident identification is claimed. Other powerful elucidation
461 techniques that do not have much application in the environmental field yet, such as LC-NMR,
462 can play a relevant role in the near future for confirmatory purposes.

463 The huge number of (non-overlapping) resources available, and the increasing volume
464 of data, which requires consistent and more efficient workflows, both remains challenges for
465 non-target screening with high resolution data. Exchange of information between institutes is
466 essential to facilitate and improve workflows (Schymanski et al., 2015). Some initiatives to
467 improve the European-wide exchange of information, such as a compilation of suspects lists,
468 have been recently reported (Norman network, 2018a, 2018b). Looking towards the future, a
469 degree of standardization that would come with greater maturity would help non-target
470 methods, providing a valuable contribution for identification purposes.

471

472 **4. The role of effect-directed analysis**

473 Effect-directed analysis (EDA) is a powerful approach for the identification of emerging
474 chemical risks. In EDA, EBTs are combined with sample fractionation using chromatography
475 and HRMS to isolate the compounds responsible for the effects observed in the bioassay used.
476 With this approach, an intrinsic prioritization is applied and the identification efforts are
477 focused on those fractions that showed to be biologically active. The combination of

478 biological/toxicological and chemical analytical tools for environmental quality assessment has
479 a long history and has led to complementary approaches developed in the United States and in
480 Europe (Burgess et al., 2013). Toxicity Identity Evaluation (TIE) was developed primarily for
481 land remediation sites, thus focusing on sediment, in vivo bioassays and bioavailability,
482 whereas the development of EDA was stimulated by the EU WFD and the need to investigate
483 the drivers of chemical water quality.

484 Typically, the EBTs used in EDA are rapid screening tools in multi-well plate format based
485 covering a wide range of toxicological endpoints, such as endocrine disruption, genotoxicity,
486 oxidative stress, etc. Both total extracts (prior to fractionation) and all fractions are tested in
487 the selected bioassay, which enables the assessment of eventual losses during fractionation
488 and antagonistic or synergistic effects (Weiss et al., 2009). Recently, an in-depth overview of
489 EDA supporting monitoring of aquatic environments has been published (Brack et al., 2016),
490 focusing on toxicity testing, sampling, extraction and identification strategies. The advantages
491 of the implementation of integrated tools on effect-based and chemical identification and the
492 monitoring of organic pollutants were highlighted in a European demonstration program
493 (Tousova et al., 2017).

494 Although some EDA studies reported the identification of compounds, e.g.
495 photosynthesis inhibitors in coastal waters (Booij et al., 2014) and mutagenic aromatic amines
496 in river water (Muz et al., 2017), and despite the technological advancements of HRMS,
497 significant progress with regard to the identification of chemicals responsible for observed
498 effects in EBTs has not been truly achieved. From the perspective of acceptance of EDA for e.g.
499 investigative monitoring in the WFD, the studies were too laborious and time consuming. To
500 tackle these two aspects, the EDA approach was transformed to a high throughput tool using
501 very small scale fractionation, i.e. the collection of fractions only a few seconds wide and thus
502 small volumes, ideally enabling a more straightforward coupling of the biological activity to the
503 chemical identity. For high-throughput EDA (HT-EDA), a high-resolution fractionation platform
504 was developed for fractionation into 96-well plates and used with an estrogenicity assay to
505 direct the analysis (Jonker et al., 2015). The direct fractionation of EDA extracts into 96-384-
506 1536 well plates demands the downscaling of toxicity assays, such as the transthyretin binding
507 assay using a fluorescent probe for high throughput screening of thyroid hormone disruption
508 in environmental samples (Ouyang et al., 2017). Other assays that have been miniaturized for
509 use in HT-EDA cover genotoxicity, (anti-)estrogenicity and (anti-) androgenicity and
510 arylhydrocarbon receptor (AhR) binding (Zwart et al., 2018a, 2018b).

511 Complementary to the ubiquitously used LC fractionation, strategies for GC fractionation
512 were developed in order to cover a larger part of the chemical space by focusing on volatile

513 compounds. An example focused on the AhR-binding assay and the identification of PAHs
514 (Pieke et al., 2013) in sediments. Improvement of the GC fractionation platform for bioactivity
515 screening of toxic compounds was demonstrated by the analysis of a mixture of test
516 pesticides, which after fractionation into a 384 well plate were tested in a post-column
517 acetylcholinesterase (AChE) assay (Jonker et al., 2016). Using a similar approach, (anti)
518 androgenicity testing using the AR-Ecoscreen in 384 well plate format has been reported
519 (Jonker et al., 2017).

520 Very small fractions may also be obtained by LCxLC, using two LC columns with different
521 (orthogonal) functionalities (Ouyang et al., 2016). A strategy based on LCxLC fractionation
522 resulting in microfractions of 9 s into 4x96 well plates, an AChE inhibition assay and parallel
523 identification by TOF-MS resulted in the identification of 3 psychoactive drugs in the effluent of
524 a WWTP. Apart from the high resolution fractionation that can be achieved by the
525 implementation of LCxLC, compounds were identified by two-dimensional retention alignment
526 as well as their AChE inhibition activity, which significantly contributes to the reliable
527 identification of bioactives in the aquatic environment.

528 Compound identification in EDA is carried out using non-target screening, following
529 analogous identification pipelines as described above. For a first check of reported toxicity of a
530 tentatively identified compound, toxicity databases such as the CompTox Chemistry
531 Dashboard (Williams et al., 2017), ToxCast and PubChem Bioassay may be consulted. To
532 facilitate identification, dedicated databases for e.g. a specific toxicological endpoint or a
533 specific sample matrix have been developed. Thus, large databases of compounds showing
534 thyroid hormone disrupting activity (Weiss et al., 2015) or directed towards the identification
535 of chemicals in house dust (Lucattini et al., 2017) have been reported. In addition to compound
536 libraries and toxicity databases, identification efforts are expected to greatly benefit in the
537 future from the implementation of less commonly used MS interfaces such as APCI and APPI
538 for specific compound classes.

539 The HT-EDA platforms based on LC and GC fractionation into ≥ 96 well plate for rapid
540 toxicity screening and parallel identification using HRMS/MS, comprehensive compound
541 databases (e.g. ChemSpider or PubChem, that contain properties such as accurate mass, log
542 Kow, etc), in addition to mass spectral libraries, are expected to contribute to an improved
543 framework for investigative monitoring in the environment.

544 Other approaches than EDA could be followed to reduce the complexity of the analysis,
545 such as the use of molecular imprinted polymers (Kubo & Otsuka, 2016) for very specific
546 extractions, or the use of antibodies (Li et al., 2017), among others. However, a complete
547 description would be far beyond the scope of the paper.

548

549 **5. Relevance of transformation products**

550 Once released into the environment, most organic contaminants are subjected to
551 biotic and abiotic transformations. These transformations generate a great variety of TPs that
552 differ from the parent compound in both their environmental behaviour and (eco)toxicity.
553 Unfortunately, there is a significant gap between the knowledge on the occurrence of TPs and
554 their toxicity, making risk assessment difficult (Agüera et al., 2013; Farré et al., 2008).

555 Current legislation is almost exclusively directed towards regulating parent chemicals,
556 with very little mention of TPs. Usually, information is lacking on analytical determination,
557 occurrence, and toxicological effects of TPs. Therefore, there is a clear need to reveal the
558 quantitative and qualitative presence of TPs in the environment and their potential risks for
559 human health.

560 HRMS has been used for identification of TPs (Díaz et al., 2012; Hernández et al., 2014,
561 2011; Hogenboom et al., 2009; Ibáñez et al., 2017; Kern et al., 2009; Nurmi et al., 2012;
562 Schollée et al., 2015), but discovering TPs in environmental samples is still challenging, as they
563 are generated from many possible reactions and automated identification workflows are still
564 missing. In most cases, a time-consuming manual inspection is required followed by the
565 acquisition of reference standards. The latter step used to be one of the main limitations
566 because standards are in many cases unavailable for TPs.

567 TPs occurring in the environment can be formed from biotic or abiotic processes. Biotic
568 processes include the activity of microbial flora in natural or engineered environmental
569 compartments, like soils, surface water or wastewater, or human or animal metabolism.
570 Abiotic TPs are formed in the aquatic environment by hydrolysis and photolysis, as well as in
571 water-treatment processes such as oxidation by chlorination, chloramination, ozonation and
572 other advanced oxidation processes used for disinfection and removal of chemicals (Bletsou et
573 al., 2015; Fenner et al., 2013; Vughs et al., 2018).

574 Simulating the transformation processes in laboratory experiments under controlled
575 conditions is a useful approach for identification of TPs (Boix et al., 2016b, 2013; Helbling et al.,
576 2010). Batch experiments can be performed under biotic or abiotic conditions at realistic or at
577 high concentrations of the compounds. During such experiments, spiked and blank samples
578 are run in parallel, together with sterilized blank samples as well as ultrapure water for
579 correcting abiotic processes (Wick et al., 2011).

580 Even under laboratory-controlled conditions, the identification of TPs is a time-
581 consuming process as commonly many products are generated; therefore, prioritization of
582 relevant TPs is recommended. This can be performed using time trends (the TP is observed to

583 form, or form and then transform further), abundance (the most intense peaks are identified),
584 prediction (transformation processes are predicted using *in silico* methods) or using observed
585 toxic effects. The latter assumes that if increased toxicity is observed, toxic TPs are likely to be
586 formed (Escher and Fenner, 2011) and it is worth to invest time and effort to identify them
587 (see above section on EDA).

588 Once the laboratory experiments are completed and the main TPs identified, the next
589 step is analysing real-world environmental samples to test their possible presence in the
590 samples (Bijlsma et al., 2013; Boix et al., 2016a, 2014; Duester et al., 2017; Kolkman et al.,
591 2015; Vughs et al., 2018). LC-HRMS(/MS) is the most common technique for investigation of
592 TPs as these are generally more polar than parent compounds. In addition, HRMS(/MS) allows
593 the application of different workflows for efficient identification: target analysis for already
594 known TPs with standards available; suspect screening for potential TPs reported in literature
595 or predicted by *in silico* models; and non-target analysis for discovering unknown, non-
596 reported nor previously identified TPs.

597 The formation of TPs by bacteria degradation can be predicted using different *in silico*
598 tools, such as the Eawag-Pathway prediction system (enviPath, formerly UM-PPS) (Wicker et
599 al., 2015) or MINE (Jeffryes et al., 2015), whereas other comprehensive knowledge-based
600 software programmes, like Meteor Nexus-Lhasa, predict the metabolic fate of chemicals,
601 among other processes. It is, however, noteworthy that the latter tools are often not freeware.
602 Obviously, these predictors have limitations yet they have also shown its value in different
603 fields of research. More applications can be expected for environmental toxicology in the near
604 future (Miller et al., 2018).

605 After prediction, suspect screening of expected exact masses of TPs is conducted in
606 HRMS data, usually comparing versus a control sample. The plausibility of RT, isotopic pattern
607 and ionization mode can be used to reduce the candidate peaks, as well as MS/MS fragment
608 peaks if available. Another interesting possibility to investigate the presence of TPs in waters is
609 a directed non-target analysis, which implies the search of unknown compounds within a
610 limited chemical space. This approach relies on the assumption that many TPs maintain
611 similarity in their chemical structure with the parent compound, and therefore present several
612 common fragment ions (Hollender et al., 2017; Ibáñez et al., 2017; Zonja et al., 2015). Thus,
613 the investigation of TPs for a given compound is not a truly non-target analysis, because the
614 searching is limited to those compounds that share a given chemical structure or fragment.
615 Searching for common fragments is a powerful strategy, able to detect and tentatively identify
616 TPs/metabolites in *the* samples, although it is time-consuming and requires a notable
617 knowledge of mass fragmentation rules. This approach benefits when all-ion-fragmentation

618 (AIF) acquisitions (also known as MS^E, all ion or broad-band CID) are performed during HRMS
619 screening (Castillo et al., 2016; Gómez Ramos et al., 2019; Hu et al., 2017; Kinyua et al., 2015;
620 Telving et al., 2016). This allows to obtain simultaneous information on intact molecules (low
621 collision energy, LE) as well as their fragments (high collision energy, HE) without precursor ion
622 selection. After a detailed study on mass fragmentation, it is possible the tentative
623 identification of the TP.

624 The common fragments strategy can be extended to the metabolites/TPs identified in
625 the samples, which means that some TPs can be discovered in an iterative process because
626 they share fragments, not with the parent molecule but with other TPs previously identified in
627 the samples (Boix et al., 2016a). Interesting extensions to this approach in metabolomics
628 includes the application of text-mining to interpreting fragmentation data (van der Hooft et al.,
629 2016) as well as molecular networking (Wang et al., 2016).

630 Unfortunately, the information available on TPs in databases is still very limited and
631 the success rate in the identification is low in non-target screening. Scientists performing such
632 studies should ensure that their data can be reused by making this accessible (Schymanski and
633 Williams, 2017). Additional techniques, as NMR, are often critical for an unambiguous
634 identification of TPs where positional isomers are likely to occur, but it is rare that sufficient
635 amount of analyte can be isolated for NMR measurement in environmental samples.
636 Unfortunately, reference standards for TPs are not always available; therefore, identification
637 of TPs in water samples mostly relies on (accurate) mass data interpretation and
638 communicating the confidence of the identification is critical (Schymanski et al., 2014a).

639

640 **6. Early warning systems**

641 Monitoring long-term trends and changes in water quality is the first step towards a
642 deeper understanding of water resources. Beyond event monitoring, establishing baseline
643 water quality has become increasingly important for protecting source water as it is affected
644 by urbanization, effluents from industry and agricultural activities. Continuous online
645 monitoring is a key initiative and it can lead to smart water management and informed
646 decision making by providing early warnings about changes in water quality.

647 Early Warning Systems (EWS) are characterized as being early in time to take measures
648 (Early) against water quality exceeding defined standards (Warning) with a well-defined
649 system (System). They can provide accurate and clear information, necessary to facilitate
650 adequate decision-making, and have been used in a wide range of applications (Towhata and
651 Uchimura, 2013), such as cyanobacterial blooms (Shi et al., 2013), water levels (Kuantama and

652 Saraswati, 2015), specific chemical compounds (Imen et al., 2015) and emerging contaminants
653 (Alygizakis et al., 2018).

654 The real time monitoring can be based on), physical (temperature, redox potential,
655 conductivity, turbidity, and spectroscopic techniques, such as fluorescence, HPLC-UV, LC-MS),
656 chemical (pH, ammonium, nitrate, chloride, fluoride, phosphate, oxygen) and biological
657 parameters (measuring live responses in Daphnia, fish, mussels and algae to water quality)
658 (Carstea et al., 2016). Chemical and biological monitoring are both valuable approaches for
659 water quality monitoring, but in most cases of an alarm, chemical analysis with identification
660 techniques (e.g., HRMS) is necessary for tracking the cause of the alarm and elucidating the
661 source of the pollution (de Hoogh et al., 2006; Hollender et al., 2017; van Leerdam et al.,
662 2014).

663 EWS present advantages, such as the continuous availability of the on-line information,
664 the combined information of the different systems and the low costs for running the system in
665 many cases. However, some drawbacks limit their application, as for example often relative
666 high detection limits compared to specific analysis in a laboratory. Improvements are required
667 for an unambiguous interpretation of data, and standardization is needed to increase the
668 comparability of results. Minimizing false positive and false negative responses and valid
669 quality assurance according to protocols also need to be improved in the near future. The
670 identification of specific (organic) compounds responsible of pollution events is surely the
671 most complex issue at present. The implementation of analytical measurements based on
672 HRMS for wide-scope screening and reliable identification of the compounds is among the
673 most urgent needs in the application of EWS and has already been showcased in the Rhine
674 River (Hollender et al., 2017) and for emerging contaminants in NormaNEWS (Alygizakis et al.,
675 2018).

676 EWS should be able to detect and identify signals of concern due to the presence of
677 chemicals. To this aim, several methodologies have been reported and the strengths and
678 challenges of different prioritization strategies for emerging pollutants in Europe have been
679 discussed (Brack, 2015; Dulio et al., 2013; Dulio and Slobodnik, 2015; Faust and Backhaus,
680 2015; Heiss and Küster, 2015). As an illustrative example, a prioritization approach based on
681 occurrence data and PNEC (derived from experimental data, QSAR or read-across predictions),
682 has been recently developed classifying chemicals into 6 categories depending on the
683 information available (Dulio and Slobodnik, 2015). These categories define the need for action,
684 including: 1) priority regular monitoring, 2) watch list monitoring, 3) extension of the
685 (eco)toxicological data set, 4) improvement of analytical methods, 5) extension of both
686 monitoring and (eco)toxicological data, and 6) compounds classified as low priority for regular

687 monitoring due to estimated low risks (Brack et al., 2017). The approach starts from a list of
688 compounds identified by experts as frequently discussed emerging substances. Monitoring
689 data of known hazardous compounds in water using multi-residue methods is the main source
690 of exposure information, and thus new and/or unknown hazardous substances that are out of
691 the scope of the analytical methods and monitoring programs cannot be traced. However,
692 most recent scientific data are included and risk is predicted despite data gaps to bring
693 potential emerging chemical risks to the attention of regulators. Using this approach several
694 compounds identified have been included into the Watch list to obtain more widespread
695 monitoring information for improved risk assessment.

696 Another example comes from the Dutch National Institute of Public Health and
697 Environment (RIVM) that developed a general methodology to identify, prioritize and suggest
698 follow-up regulatory measures for new and/or emerging risk chemicals (NERCs) for protection
699 of three vulnerable groups (workers, consumers and the environment), and to support
700 strategic and policy decision-making. The general strategy is based on several steps, involving
701 internet search systems, scientific literature, monitoring studies, and a multidisciplinary group
702 of experts that perform data evaluation (for more details, see (Bakker et al., 2014)). The first
703 crucial step is to pick-up early signals on possible NERCs for each vulnerable group, followed by
704 collecting relevant information on the potential NERCs identified. In the case of workers and
705 consumers, data collection is focused on adverse health outcomes related to exposure from
706 various sources, 'exposure first method'. For the environment, this process is severely
707 hampered by the complexity of samples, their unknown composition, the presence of
708 numerous other compounds/contaminants and their highly fluctuating concentrations.
709 Monitoring studies are of great relevance to get information on potential NERCs, as they help
710 to identify contaminants actually occurring in the environment that may be of concern (ter
711 Laak et al., 2015). Unfortunately, monitoring data are commonly limited and focused on well-
712 known and/or priority contaminants, and thus many potential hazardous compounds remain
713 ignored due to difficulties/limitations of the analytical methods. Therefore, it is rather difficult
714 to relate observed effects to a specific chemical or combination of chemicals. EDA is a
715 promising, but still challenging method, to deal with this complexity and improved non-target
716 screening approaches are likely to assist greatly in the next years (Hollender et al., 2017). It is
717 also likely that "big data" sciences may revolutionize the way we deal with non-target data in
718 the next decades (Aksenov et al., 2017).

719 Using the general strategy proposed by RIVM, NERCs have been identified for the three
720 vulnerable groups, including the recommendation of follow-up steps to reduce or eliminate
721 the risks. The necessity to have more monitoring data available has been recognized to get a

722 better picture of occurrence of NERCs in the environment (Bakker et al., 2014). Also, more
723 hazard information is needed, especially on chronic toxicity. Extensive lists with estimated
724 values(e.g. Norman network, 2018a) are a good start, yet more work is needed to incorporate
725 these in environmental investigations.

726

727 **7. The protection of drinking water: a priority issue**

728 Providing high quality drinking water is an exigency of the society at present, and a
729 priority issue for exposure science (some examples are EU funded research projects, such as
730 EU project PROMOTE, and current initiatives by the German UBA who attempt to include
731 Mobility (of organic contaminants) as a criterion in REACH legislation). Producing safe drinking
732 water starts with knowledge about source quality and on potential contamination related to
733 activities within the catchment area that may affect water quality constantly or during a
734 calamity (e.g. industrial activities, agricultural activities, infrastructure and transport activities,
735 presence of residential areas and the activities within these areas, historical contamination).
736 Furthermore, the removal rates and robustness of the treatment process are essential for
737 knowing whether suitable drinking water can be produced from a certain source, and whether
738 the system is resilient against future changes of water quality and calamities.

739 Determine the chemical water quality of sources, the quality of the produced water
740 and the performance and robustness of treatment systems are key issues. The Drinking Water
741 Directive (Council Directive 98/83/EC) concerns the quality of water intended for human
742 consumption with the objective to protect human health from adverse effects of any
743 contamination. On February 2018, the European Commission adopted a proposal for a revised
744 Directive to improve the quality of drinking water and provide greater access and information
745 to citizens. In the Annex of this proposal, quality standards for chemicals are listed
746 ([http://www.europarl.europa.eu/RegData/etudes/BRIE/2018/625179/EPRS_BRI\(2018\)625179](http://www.europarl.europa.eu/RegData/etudes/BRIE/2018/625179/EPRS_BRI(2018)625179_EN.pdf)
747 [_EN.pdf](http://www.europarl.europa.eu/RegData/etudes/BRIE/2018/625179/EPRS_BRI(2018)625179_EN.pdf)). The fast detection of relevant changes in (source) water quality is of great
748 importance to take timely measures, when necessary. The analytical tools range from fast
749 generic biological and chemical sensors and automated chemical analysis for fast detection of
750 (large) changes in water quality to very sensitive chemical analysis of a wide array of
751 compounds, as well as bioassays to assess (adverse) effects of substances (Altenburger et al.,
752 2015; Bäuerlein and Kolkman, 2016; Storey et al., 2011). Regular monitoring of sources and
753 produced drinking water is mostly done with target analysis of priority/relevant contaminants
754 (e.g. pesticides is one of the groups of highest concern). However, it is questionable whether
755 this is sufficient for appropriate quality control with the seemingly endless numbers of

756 substances that remain unmeasured or unidentified. Biological and chemical non-target
757 screening approaches have been applied to evaluate technical processes e.g. chlorination,
758 ozonation or UV-peroxide during water treatment, as it offers the possibility of assessing the
759 effectiveness without knowing the identity of the substances that are eliminated or newly
760 formed during a particular process (ter Laak et al., 2012). Even if identification is not achieved,
761 the non-target analysis is still a valuable tool as it allows comparing different samples (Bader et
762 al., 2016; Müller et al., 2011; Nürenberg et al., 2015).

763 The identities and concentrations of chemical contaminants in source waters
764 determine the type of treatment required for their removal. In particular, persistent
765 compounds that are also mobile (PMOCs), as a result of their high aqueous solubility, can
766 constitute a threat to the quality of our water resources (Reemtsma et al., 2016). Highly polar
767 or ionic substances can pass through WWTPs as well as natural barriers (river banks, sand
768 dunes) and thus become present in source waters. Treatment processes applied in drinking
769 water production may not be able to completely remove such substances, and some processes
770 may even result in the generation of new, even more recalcitrant polar compounds. Examples
771 are TPs generated in e.g. disinfection (chlorination, ozonation, UV-peroxide) treatment, some
772 of which have been shown to be particularly hazardous (Kolkman et al., 2015; Richardson et
773 al., 2007; Richardson and Kimura, 2016; Schmidt and Brauch, 2008). The control of such
774 processes requires sophisticated LC-HRMS methodologies to identify the possible products
775 generated.

776 The lack of appropriate analytical methodologies is one of the major gaps in
777 knowledge on occurrence of highly polar PMOCs in water samples (Reemtsma et al., 2016).
778 This is so because their polarity range - characterized by negative values of log D - makes them
779 hardly amenable to either GC or reversed-phase LC separation. Possible solutions may be
780 found in hydrophilic interaction liquid chromatography (HILIC) (Zahn et al., 2016), mixed-mode
781 LC (Montes et al., 2017; Ordoñez et al., 2012), combining HILIC and reversed phase LC (Bieber
782 et al., 2017), ion-pairing LC or supercritical fluid chromatography (SFC).

783 Protection of drinking water requires often advanced methods of treatment for
784 removal of emerging contaminants. While advanced oxidation methods have been shown to
785 efficiently remove traces of non polar organic compounds, they may not sufficiently protect
786 against the persistent mobile organics. Further refinement of such methods is underway and
787 includes combinations of ozonation and/or UV-peroxide treatment with granular active carbon
788 filtration (Bourgin et al., 2017). Even such combinations may not necessarily suffice, as
789 demonstrated by the presence of polar compounds in drinking water (see text above).
790 Membrane processes, such as nanofiltration and reverse osmosis (Farré et al., 2011; Fujioka et

791 al., 2012; Hajibabania et al., 2011; Verliefde et al., 2009) can remove polar and ionogenic
792 compounds more efficiently than advanced oxidation processes, but still require more testing
793 and understanding to guarantee their efficacy towards these compounds.

794

795 **8. Wastewater: a valuable source of information on lifestyle and exposure to** 796 **chemicals**

797 Wastewater can provide anonymised but comprehensive and objective information on
798 community-wide health status and lifestyle in real time, as urban water (sewerage system and
799 receiving aqueous environment) pools anonymous urine, wastewater and runoff samples from
800 thousands of households that are served by individual WWTPs. The monitoring of sewage for
801 chemicals used as indicators of the collective status of human health (stress/disease) or any
802 other facet relevant to determine trends in community-wide health is the basis of wastewater-
803 based epidemiology (WBE), which is being used to monitor community-wide use of lifestyle
804 substances such as illicit drugs, new psychoactive substances (NPS), alcohol, tobacco, doping
805 substances and counterfeit medicines. WBE enables the retrieval of epidemiological
806 information from wastewater via analysis of specific human (metabolic) excretion products
807 (called biomarkers). This approach is currently used to report on world-wide illicit drug use
808 trends e.g. (Ort et al., 2014; Thomas et al., 2012; Tschärke et al., 2016) and feeds into the
809 Europe-wide evidence based EWS managed by the European Monitoring Centre for Drugs and
810 Drug Addiction (EMCDDA, accessed 11.7.18 ; SCORE, accessed 11.7.18).

811 Human excretion products of external or internal bodily origin resulting from intentional
812 or unintentional exposure to foreign 'xenobiotic' agents and internal processes (e.g.
813 metabolism of xenobiotics, xenobiotic triggered DNA damage leading to excretion of
814 characteristic metabolites, proteins and DNA biomarkers) are pooled by the sewerage system.
815 Analysis of appropriate biomarkers in wastewater provides valuable evidence of the quantity
816 and type of xenobiotic substances to which a population is exposed by e.g., deliberate
817 administration of illicit drugs, or accidental chemicals exposure to food, environmental
818 toxicants or infectious agents (Daughton, 2018; Kasprzyk-Hordern et al., 2014). More
819 information can be obtained for guiding investigation needed for preventing, avoiding,
820 controlling or reducing human exposure risks, as well as for maintaining or promoting health.

821 The analysis of wastewater opens new perspectives within exposure science thanks to
822 the vast amount of information contained in this type of samples. This is because wastewater
823 analysis can provide information about the presence of stressors in the studied system as well
824 as community-wide biological effects (e.g. prevalence of certain diseases). To this aim, the
825 identification and quantification of appropriate urinary biomarkers of exposure (metabolic

826 residues of stressors) and effects (e.g. biomarkers of inflammation or oxidative stress) is
827 required. Human biomarkers of exposure are typically present at very low concentrations in a
828 very complex matrix, such as wastewater. LC-MS/MS QqQ is the technique most commonly
829 used, although LC-HRMS is increasingly being used in this field too (Hernández et al., 2018).
830 The concentrations of biomarkers can be used to back-calculate their daily mass loads in
831 wastewater and to estimate daily exposure after taking into consideration knowledge (if exists)
832 on human disposition, including metabolism (Castiglioni et al., 2013; Thomas et al., 2012).

833 Analysis of wastewater can be applied to assessment of spatial and temporal trends in
834 (1) lifestyle and substance use (e.g. illicit drugs, alcohol, tobacco or NPS); (2) health status (e.g.
835 stress) and (3) exposure to food and environmental toxicants (e.g. pesticides (Gracia-Lor et al.,
836 2018; Roussis et al., 2016), chemicals in personal care products (antimicrobials and UV filters)
837 (Lopardo et al, 2017, 2018), phosphorus flame retardants (Been et al., 2017), and phthalate
838 plasticizers (Gonzalez-Mariño et al, 2017))

839 An interesting analytical approach is the use of chiral chromatography, which allows for
840 understanding of origins of biomarkers in wastewater (i.e. differentiation between
841 consumption/direct exposure and other disposal routes, e.g. direct disposal of unused drug
842 (Castrignanò et al., 2018; Emke et al., 2014; Kasprzyk-Hordern and Baker, 2012; Petrie et al.,
843 2016; Vazquez-Roig et al., 2014). Sensors have also been developed for cheaper and faster
844 quantification of biomarkers (such as cocaine and prostate specific antigen (PSA)) (Yang et al.,
845 2016, 2015a, 2015b). Tracking of infectious disease and linking exposure to air pollutants with
846 respiratory diseases has been proposed with the ultimate goal of a comprehensive public
847 health assessment, since urban water can be considered as a 'diagnostic medium for the
848 health status of a city' (Daughton, 2018; Thomas and Reid, 2011; Yang et al., 2015a).

849

850 **9. Regulatory issues**

851 The regulation of chemicals on the market ensures consumer and environmental
852 safety, yet the information required in the regulatory processes often remains confidential to
853 some extent to protect commercial/trade secrets. Furthermore, many of these chemicals are
854 not listed in public compound databases often used for screening. With an estimated 140,000
855 chemicals regulated under REACH, including confidential information, market use (tonnage,
856 intended use) and hazard classifications for a subset of these (approx. 15-30,000 substances),
857 the information held by regulatory authorities could provide valuable assistance for non-target
858 and suspect screening. However, the majority of these are industrial chemicals and some level
859 of curation is needed to apply this information for screening purposes. Strengthening the

860 collaboration between regulatory authorities and environmental laboratories would lead to
861 mutual benefits – an example workflow of how to protect confidentiality yet yield vital
862 information for screening of emerging chemical risks is given in **Figure 3**. Regulatory bodies
863 could provide for instance accurate mass data instead of confidential substance structures as
864 well as exposure indexes instead of confidential consumption or application data to
865 laboratories for suspect screening purposes. In case of potential environmental or human risk,
866 reported findings could result in additional confirmatory and regulatory actions. Beyond
867 exchanging information with local regulatory authorities, for instance in the context of
868 granting permits, there is also an increasing need to exchange information internationally, as
869 the increasing use of imported products may also result in adverse human and environmental
870 impacts. Initiatives such as the NORMAN Suspect Exchange (Norman network, 2018a) and the
871 chemical lists on the CompTox Chemistry Dashboard (Williams et al., 2017) are important steps
872 to improve the exchange of information. Both resources now contain chemical lists with
873 exposure estimates. The ExpoCast list (Wambaugh et al., 2014) contains estimated data on
874 7968 chemicals, also available from the batch search to download. The KEMI Market List on
875 the NORMAN Suspect Exchange contains information and associated exposure indexes and
876 hazard scores on 30,748 chemicals (version provided 3 July 2017; further details about these
877 scores provided on the Suspect Exchange). The Chemicals and Products Database contains
878 reported and predicted information on >75,000 chemicals contained in >15,000 consumer
879 products (Dionisio et al., 2018) and is also available through the CompTox Chemistry
880 Dashboard.

881 Widening/improving suspect lists with the inputs from regulatory bodies is extremely
882 beneficial for screening purposes, and the tasks of analytical laboratories would be notably
883 facilitated, resulting in a more efficient screening, providing more realistic data on the
884 presence of contaminants of concern in monitoring programs (Gago-Ferrero et al., 2018;
885 Schulze et al., 2018). At the same time, a close collaboration would make easier the updating
886 of regulation based on occurrence on harmful compounds in the environment. As an example,
887 an exchange of information on PMOCs would be beneficial, as these contaminants have the
888 greatest chances of appearing in drinking water, because they are mobile enough in the
889 aquatic environment to enter drinking water sources and persistent enough to survive
890 treatment processes. This identification and ranking procedure for PMOCs can be part of a
891 strategy to better identify contaminants that pose a threat to drinking water sources (Arp et
892 al., 2017).

893 However, exact mass or suspect screening, especially using large lists of compounds, is
894 not sufficient. All “hits” detected in such a manner must be confirmed using additional

895 analytical information to avoid false positives. RT and fragmentation information can be used
896 to provide additional evidence for a formula or exact mass hit and library spectra or reference
897 standards should be used (as described further above) for additional confirmation. Progressive
898 efforts are underway to integrate suspect lists containing exposure data into *in silico*
899 fragmentation methods such as MetFrag (Ruttkies et al., 2016) to facilitate the connection
900 between research and regulation. Efforts are also underway to improve the assessment of
901 materials of Unknown and Variable composition, Complex reaction products and Biological
902 substances (UVCBs), which make up a significant percentage of chemical substances in
903 regulatory lists (Schymanski and Williams, 2017; Williams et al., 2017). For instance, 16,923 of
904 67,951 entries (25 %) in the April 2018 Toxic Substances Control Act (TSCA) are UVCBs.

905 Non-targeted analytical methods are suited to discover potential emerging chemical
906 risks in the environment. With improved, rapid communication and increasing regulatory
907 acceptance of screening methods, this has great potential to assist in the monitoring of this
908 type of compounds

909

910

Insert Figure 3

911

912 Chemicals identified as substances of very high concern (SVHC) in accordance with the
913 REACH Regulation are added to the so-called Candidate List, and then become subject to
914 authorization. To date, all chemicals included in the Candidate List that are classified as either
915 persistent, bioaccumulative and toxic (PBT), or very persistent and very bioaccumulative, are in
916 principle considered to be hazardous to the environment. However, really polar, mobile
917 chemicals have not been identified as SVHC. Thus, neither in the protection of surface water
918 and groundwater nor in REACH regulation PMOCs have been considered specifically, yet.
919 Although REACH does not specifically address PMOCs they can be covered under article 57 (f)
920 by identifying them as substances of equivalent concern. Thus, there is a strong need to
921 establish criteria for identifying PMOCs in order to prevent that this becomes a regulatory gap.
922 Currently, the lack of analytical methods, monitoring, and modeling data hampers an
923 evaluation of the magnitude of this gap (Reemtsma et al., 2016).

924 Despite limitations in the regulation, many countries, water authorities and
925 waterworks are taking advantage of the improved analytical capabilities and are increasing
926 their efforts to cover a broad range of emerging chemical risks in their monitoring following
927 the precautionary principle and due to public concern. Accordingly, most regulatory
928 laboratories already have GC-MS and/or GC-MS/MS, as well as LC-MS/MS, at their disposal,
929 and many have acquired HRMS instruments to broaden their analytical capabilities further.

930 This trend will most likely continue and result overall in better surveillance of water resources.
931 Improved data processing techniques and acceptance will support these efforts further. The
932 field is currently developing very rapidly and there is much reason to be positive for the future
933 of non-target screening methods in regulatory contexts (Hollender et al., 2017).

934

935 **10. Trends and perspectives**

936 To conclude the information presented in this article, firstly several key issues are
937 emphasized, then upcoming trends and perspectives are anticipated for the next few years in
938 relation to the role of analytical chemistry in the field of exposure science.

- 939 • Early warning systems show great promise as both a pro-active and retrospective
940 method to enable the timely capture of insufficiently characterized chemical threats in
941 waters. To prevent potential risks from unregulated aspects of chemicals, emerging
942 chemical risks must be traced and identified at the earliest possible stage (Bakker et
943 al., 2014; Fahrenkamp-Uppenbrink, 2018). The lack of data on occurrence of many
944 chemicals and their transformation products in the environment is a limitation at
945 present. To fill this gap, the complementary use of gas and liquid chromatography
946 hyphenated with high resolution mass spectrometry appears as one of the most
947 powerful approaches for wide-scope screening. Environmental analytical chemistry will
948 surely be a priority research area in exposure science in the next years.
- 949 • Working with large databases of compounds (e.g. in suspect screening) requires
950 careful prioritization and confirmation efforts to focus on the most relevant
951 compounds. Prioritization should be given to those chemicals where high exposure or
952 potential risk to human and environmental health is expected, and relevant
953 occurrence or risk data should be included where possible.
- 954 • Metabolites/TPs of organic contaminants are commonly present in waters at
955 concentrations sometimes exceeding those of the parent chemicals, and therefore
956 should be also considered as major environmental contaminants, e.g. those
957 compounds that may remain biologically active. Further research is needed to identify
958 and quantify these compounds in the aquatic environment, and to establish their
959 potential hazards and eco-toxicological effects. Linking contaminant levels (exposure)
960 obtained with mass spectrometry tools with biological responses (hazard) obtained
961 with rapidly developing bioassay tools provides a promise for better and faster
962 identification of those chemicals that represent a real threat to the environment and
963 public health.

964 • The lack of multi-residue/ multi-class analytical methods for highly polar, persistent
965 and mobile organic compounds implies gaps in terms of analysis, and consequently in
966 monitoring data, water treatment measures and in regulation. New methods are
967 expected in the new few years for the determination of these compounds, e.g. making
968 use of chromatographic separations based on HILIC, Mix-Mode, ion-pairing or SFC.

969 • Wastewater analysis has allowed exposure science to gain a powerful tool to verify
970 public and environmental health trends in near real time. Future developments are
971 expected in this area aimed to identify (bio)markers of exposure in wastewater.
972 Development of novel real-time, low cost and easy to operate sensors will surely be a
973 relevant issue in the near future.

974 • The increasing awareness of potential risks of organic contaminants for aquatic
975 ecosystems and human health fosters the scientific and public discussion and political
976 action for further reduction of the input of these compounds to the aquatic
977 environment. The information given by modern analytical chemistry will help in
978 making choices on the best removal process of compounds from the environment,
979 because it is possible to monitor the efficiency removal of the process applied and, at
980 the same time, the possible formation of transformation/degradation products in the
981 environment.

982 • Non-target analysis now makes possible the monitoring of both known and unknown
983 chemicals in waters in near real time and trends of these chemicals over time. It allows
984 comparing different samples and monitoring of unknown masses of interest. In this
985 way, episodes of high contamination can be detected even if the unknowns remain
986 unidentified. Especially for the evaluation of technical processes (i.e. ozonation during
987 water treatment), non-target analysis offers the possibility of assessing the
988 effectiveness without knowing the identity of the substances which are eliminated or
989 newly formed during a particular process.

990 • The most recent data reported in the literature reveal that screening by LC-HRMS now
991 leads to results similar to those from the regular target monitoring using specific (LC-
992 MS/MS) analytical methods. Still, some challenges have to be faced in the coming
993 years, such as (i) developing a further optimized quality assurance strategy, (ii)
994 optimization of the LC, prediction of retention times (Aalizadeh et al., 2016; Bade et
995 al., 2015a; McEachran et al., 2018; Miller et al., 2013; Stanstrup et al., 2015), and
996 prediction of CCS in Ion Mobility MS (Bijlsma et al., 2017; Mollerup et al., 2018; Zhou et
997 al., 2016) (iii) storage and exchange of measurement data e.g. (Wang et al., 2016) and

998 (iv) the identification of unknown compounds. The outlook is to incorporate non-target
999 LC-HRMS(/MS) screening into regulatory monitoring in the near future and embedding
1000 this in a regulatory framework. As a result, a more complete overview of the
1001 occurrence of organic compounds in the water cycle will be generated and more
1002 realistic data will be available for appropriate risk assessment.

- 1003 • Further strengthening and embedding of analytical chemistry within the field of
1004 exposure science will enhance the identification and prioritization of potential threats
1005 to health and the environment. The existing first steps outlined above with the
1006 NORMAN Suspect Exchange and the CompTox Chemistry Dashboard could be
1007 extended by setting up a European programme generating, assessing, exchanging, and
1008 communicating experimental and model-based exposure data in support of the
1009 European strategy for exposure science with a roadmap 2020-2025-2030 ("The
1010 European branch of the International Society of Exposure Science (ISES-Europe). Such
1011 activities will also be applicable on a global scale. Also on a global scale, and
1012 particularly in developing countries, UN Environment is investing since more than 10
1013 years already in capacity building for the analysis of persistent organic contaminants
1014 (Van Leeuwen et al., 2013)
- 1015 • New hazards are becoming hot topics in exposure science not related to specific
1016 chemical families but to new classes of contaminants like nanoparticles or
1017 microplastics (Pico et al., 2017, 2018; Wagner and Lambert, 2018). Nano-based
1018 technology has made enormous progress over the last decade with applications in
1019 cosmetic, pharmaceutical and medical fields using engineered nanoparticles consisting
1020 of carbon-based (eg. Fullerene) and inorganic (eg. TiO₂) forms, partly with
1021 functionalized surfaces (Bundschuh et al., 2018). The environmental concern on
1022 microplastics (i.e. fragments smaller than 1 mm) is related to their slow degradability
1023 and capability as carriers to concentrate and transport synthetic organic pollutants.
1024 Primary sources are related to cleaning and cosmetic products, while secondary
1025 sources arise as fibres coming from washing clothes, both entering aquatic system
1026 through household sewage discharge (Jiang, 2018). These have a whole new set of
1027 analytical challenges associated with them that have not been covered in this article
- 1028 • A new trend is directed towards the study of the exposome, *i.e.* the totality of
1029 environmental exposures from conception onwards, where two broad complementary
1030 interpretations can be considered. One, called "top-down", is mainly interested in
1031 identifying new causes of disease based on *omics* technologies (internal exposome)
1032 (Turner et al., 2018). This approach utilizes methods such as metabolomics to generate

1033 new hypotheses on disease causes. The second, called “bottom-up”, starts with a set
1034 of exposures or environmental compartments to determine the pathways by which
1035 such exposures lead to disease. Although the present article deals with the external
1036 exposome, focused on the aquatic environment, one should not forget that a
1037 comprehensive exposome concept also involves the internal exposome, in order to
1038 integrate the totality of environmental exposures. Analysis of the exposome (so-called
1039 exposomics) will require powerful analytical techniques, such as HRMS coupled to both
1040 GC and LC, combined with robust statistical tools and quick and trusty annotation
1041 applications, as discussed here.

1042
1043

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1055 **12. References**

- 1056 Aalizadeh, R., Thomaidis, N.S., Bletsou, A.A., Gago-Ferrero, P., 2016. Quantitative Structure-
1057 Retention Relationship Models to Support Nontarget High-Resolution Mass
1058 Spectrometric Screening of Emerging Contaminants in Environmental Samples. *J. Chem.*
1059 *Inf. Model.* 56, 1384–1398. <https://doi.org/10.1021/acs.jcim.5b00752>
- 1060 Aceña, J., Stampachiachiere, S., Pérez, S., Barceló, D., 2015. Advances in liquid
1061 chromatography–high-resolution mass spectrometry for quantitative and qualitative
1062 environmental analysis. *Anal. Bioanal. Chem.* 407, 6289–6299.
- 1063 Agüera, A., Martínez Bueno, M.J., Fernández-Alba, A.R., 2013. New trends in the analytical
1064 determination of emerging contaminants and their transformation products in
1065 environmental waters. *Environ. Sci. Pollut. Res. Int.* 20, 3496–515.
1066 <https://doi.org/10.1007/s11356-013-1586-0>
- 1067 Aksenov, A.A., Da Silva, R., Knight, R., Lopes, N.P., Dorrestein, P.C., 2017. Global chemical
1068 analysis of biology by mass spectrometry. *Nat. Rev. Chem.* 1, 1–20.
1069 <https://doi.org/10.1038/s41570-017-0054>
- 1070 Alder, A.C., Schaffner, C., Majewsky, M., Klasmeier, J., Fenner, K., 2010. Fate of β -blocker
1071 human pharmaceuticals in surface water: Comparison of measured and simulated
1072 concentrations in the Glatt Valley Watershed, Switzerland. *Water Res.* 44, 936–948.
1073 <https://doi.org/10.1016/j.watres.2009.10.002>
- 1074 Altenburger, R., Ait-Aissa, S., Antczak, P., Backhaus, T., Barceló, D., Seiler, T.-B., Brion, F., Busch,
1075 W., Chipman, K., López de Alda, M., de Aragão Umbuzeiro, G., Escher, B.I., Falciani, F.,
1076 Faust, M., Focks, A., Hilscherova, K., Hollender, J., Hollert, H., Jäger, F., Jahnke, A.,
1077 Kortenkamp, A., Krauss, M., Lemkine, G.F., Munthe, J., Neumann, S., Schymanski, E.L.,
1078 Scrimshaw, M., Segner, H., Slobodnik, J., Smedes, F., Kughathas, S., Teodorovic, I., Tindall,
1079 A.J., Tollefsen, K.E., Walz, K.-H., Williams, T.D., Van den Brink, P.J., van Gils, J., Vrana, B.,
1080 Zhang, X., Brack, W., 2015. Future water quality monitoring — Adapting tools to deal with
1081 mixtures of pollutants in water resource management. *Sci. Total Environ.*
1082 <https://doi.org/10.1016/j.scitotenv.2014.12.057>
- 1083 Alygizakis, N.A., Samanipour, S., Hollender, J., Ibáñez, M., Kaserzon, S., Kokkali, V., Van
1084 Leerdam, J.A., Mueller, J.F., Pijnappels, M., Reid, M.J., Schymanski, E.L., Slobodnik, J.,
1085 Thomaidis, N.S., Thomas, K. V., 2018. Exploring the Potential of a Global Emerging
1086 Contaminant Early Warning Network through the Use of Retrospective Suspect Screening
1087 with High-Resolution Mass Spectrometry. *Environ. Sci. Technol.* 52, 5135–5144.
1088 <https://doi.org/10.1021/acs.est.8b00365>
- 1089 Andrés-Costa, M.J., Andreu, V., Picó, Y., 2017. Liquid chromatography–mass spectrometry as a
1090 tool for wastewater-based epidemiology: Assessing new psychoactive substances and
1091 other human biomarkers. *TRAC Trends Anal. Chem.* 94, 21–38.
- 1092 Arp, H.P.H., Brown, T.N., Berger, U., Hale, S.E., 2017. Ranking REACH registered neutral,
1093 ionizable and ionic organic chemicals based on their aquatic persistency and mobility.
1094 *Environ. Sci. Process. Impacts* 19, 939–955. <https://doi.org/10.1039/C7EM00158D>
- 1095 Arp, H.P., Slinde, G.A., 2018. PFBS in the Environment: Monitoring and Physical-Chemical Data
1096 Related to the Environmental Distribution of Perfluorobutanesulfonic Acid; Norwegian
1097 Environmental Agency Report M-1122|2018, NGI DOC.NO. 20180533-01-R. Oslo
- 1098 Aus der Beek, T., Weber, F.A., Bergmann, A., Hickmann, S., Ebert, I., Hein, A., Küster, A., 2016.
1099 Pharmaceuticals in the environment-Global occurrences and perspectives. *Environ.*
1100 *Toxicol. Chem.* 35, 823–835. <https://doi.org/10.1002/etc.3339>

- 1101 Bade, R., Bijlsma, L., Miller, T.H., Barron, L.P., Sancho, J.V., Hernández, F., 2015a. Suspect
1102 screening of large numbers of emerging contaminants in environmental waters using
1103 artificial neural networks for chromatographic retention time prediction and high
1104 resolution mass spectrometry data analysis. *Sci. Total Environ.* 538, 934–941.
- 1105 Bade, R., Bijlsma, L., Sancho, J. V., Hernández, F., 2015b. Critical evaluation of a simple
1106 retention time predictor based on LogKow as a complementary tool in the identification
1107 of emerging contaminants in water. *Talanta* 139, 143–149.
- 1108 Bader, T., Schulz, W., Lucke, T., Seitz, W., Winzenbacher, R., 2016. Application of Non-Target
1109 Analysis with LC-HRMS for the Monitoring of Raw and Potable Water: Strategy and
1110 Results, in: *Assessing Transformation Products of Chemicals by Non-Target and Suspect
1111 Screening – Strategies and Workflows Volume 2.* American Chemical Society, pp. 49–70.
1112 <https://doi.org/10.1021/bk-2016-1242.ch003>
- 1113 Bakker, J., Bruinen de Bruin, Y. Hogendoorn, E.A., Kooi, M., Palmen, N., Salverda, J., Traas, T.,
1114 Sijm, D., 2014. Progress report on New or Emerging Risks of Chemicals (NERCs) RIVM
1115 Letter report 2014-0040, RIVM Letter report 2014-0040. Bilthoven, the Netherlands.
1116 <https://doi.org/10.1002/fee.1450>
- 1117 Barco-Bonilla, N., Romero-González, R., Plaza-Bolaños, P., Garrido Frenich, A., Martínez Vidal,
1118 J.L., 2010. Analysis and study of the distribution of polar and non-polar pesticides in
1119 wastewater effluents from modern and conventional treatments. *J. Chromatogr. A* 1217,
1120 7817–7825. <https://doi.org/10.1016/j.chroma.2010.10.011>
- 1121 Bartelt-Hunt, S.L., Snow, D.D., Damon, T., Shockley, J., Hoagland, K., 2009. The occurrence of
1122 illicit and therapeutic pharmaceuticals in wastewater effluent and surface waters in
1123 Nebraska. *Environ. Pollut.* 157, 786–791. <https://doi.org/10.1016/j.envpol.2008.11.025>
- 1124 Barzen-Hanson, K.A., Roberts, S.C., Oetjen, S.K., McAlees, A., Riddell, N., McCrindle, R.,
1125 Ferguson, P.L., Higgins, C.P., Field, J.A., 2017. Discovery of 40 Classes of Per- and
1126 Polyfluoroalkyl Substances in Historical Aqueous Film-Forming Foams (AFFFs) and AFFF-
1127 Impacted Groundwater. *Xiao. Environ. Sci. Technol.* 51, 2047-2057
- 1128 Bäuerlein, P.S., Kolkman, A., 2016. HPLC UV screening 2015, KWR 2016.31. Nieuwegein, the
1129 Netherlands.
- 1130 Been, F., Bastiaensen, M., Lai, F.Y., Van Nuijs, A.L.N., Covaci, A., 2017. Liquid Chromatography-
1131 Tandem Mass Spectrometry Analysis of Biomarkers of Exposure to Phosphorus Flame
1132 Retardants in Wastewater to Monitor Community-Wide Exposure. *Anal. Chem.* 89,
1133 10045–10053. <https://doi.org/10.1021/acs.analchem.7b02705>
- 1134 Bieber, S., Greco, G., Grosse, S., Letzel, T., 2017. RPLC-HILIC and SFC with Mass Spectrometry:
1135 Polarity-Extended Organic Molecule Screening in Environmental (Water) Samples. *Anal.
1136 Chem.* 89, 7907–7914. <https://doi.org/10.1021/acs.analchem.7b00859>
- 1137 Bijlsma, L., Bade, R., Celma, A., Mullin, L., Cleland, G., Stead, S., Hernandez, F., Sancho, J. V.,
1138 2017. Prediction of Collision Cross-Section Values for Small Molecules: Application to
1139 Pesticide Residue Analysis. *Anal. Chem.* 89, 6583–6589.
1140 <https://doi.org/10.1021/acs.analchem.7b00741>
- 1141 Bijlsma, L., Boix, C., Niessen, W.M.A., Ibáñez, M., Sancho, J. V., Hernández, F., 2013.
1142 Investigation of degradation products of cocaine and benzoylecgonine in the aquatic
1143 environment. *Sci. Total Environ.* 443, 200–208.
1144 <https://doi.org/10.1016/j.scitotenv.2012.11.006>
- 1145 Bijlsma, L., Serrano, R., Ferrer, C., Tormos, I., Hernández, F., 2014. Occurrence and behavior of
1146 illicit drugs and metabolites in sewage water from the Spanish Mediterranean coast

- 1147 (Valencia region). *Sci. Total Environ.* 487, 703–709.
1148 <https://doi.org/10.1016/j.scitotenv.2013.11.131>
- 1149 Bletsou, A.A., Jeon, J., Hollender, J., Archontaki, E., Thomaidis, N.S., 2015. Targeted and non-
1150 targeted liquid chromatography-mass spectrometric workflows for identification of
1151 transformation products of emerging pollutants in the aquatic environment. *TrAC -*
1152 *Trends Anal. Chem.* 66, 32–44.
- 1153 Bobeldijk, I., Vissers, J.P., Kearney, G., Major, H., van Leerdam, J., 2001. Screening and
1154 identification of unknown contaminants in water with liquid chromatography and
1155 quadrupole-orthogonal acceleration-time-of-flight tandem mass spectrometry. *J.*
1156 *Chromatogr. A* 929, 63–74. [https://doi.org/10.1016/S0021-9673\(01\)01156-6](https://doi.org/10.1016/S0021-9673(01)01156-6)
- 1157 Boecking, W., Klamar, A., Kitzmann, F., Kirch, W., 2012. Pharmaco-economic impact of
1158 demographic change on pharmaceutical expenses in Germany and France. *BMC Public*
1159 *Health* 12, 894–902. <https://doi.org/10.1186/1471-2458-12-894>
- 1160 Boix, C., Ibáñez, M., Bagnati, R., Zuccato, E., Sancho, J. V., Hernández, F., Castiglioni, S., 2016a.
1161 High resolution mass spectrometry to investigate omeprazole and venlafaxine
1162 metabolites in wastewater. *J. Hazard. Mater.* 302, 332–340.
1163 <https://doi.org/10.1016/j.jhazmat.2015.09.059>
- 1164 Boix, C., Ibáñez, M., Bijlsma, L., Sancho, J. V., Hernández, F., 2014. Investigation of cannabis
1165 biomarkers and transformation products in waters by liquid chromatography coupled to
1166 time of flight and triple quadrupole mass spectrometry. *Chemosphere* 99, 64–71.
1167 <https://doi.org/10.1016/j.chemosphere.2013.10.007>
- 1168 Boix, C., Ibáñez, M., Sancho, J. V., Niessen, W.M.A., Hernández, F., 2013. Investigating the
1169 presence of omeprazole in waters by liquid chromatography coupled to low and high
1170 resolution mass spectrometry: Degradation experiments. *J. Mass Spectrom.* 48, 1091–
1171 1100. <https://doi.org/10.1002/jms.3260>
- 1172 Boix, C., Ibáñez, M., Sancho, J. V., Parsons, J.R., Voogt, P. de, Hernández, F., 2016b.
1173 Biotransformation of pharmaceuticals in surface water and during waste water
1174 treatment: Identification and occurrence of transformation products. *J. Hazard. Mater.*
1175 302, 175–187. <https://doi.org/10.1016/j.jhazmat.2015.09.053>
- 1176 Boix, C., Ibáñez, M., Sancho, J. V., Rambla, J., Aranda, J.L., Ballester, S., Hernández, F., 2015. Fast
1177 determination of 40 drugs in water using large volume direct injection liquid
1178 chromatography – tandem mass spectrometry. *Talanta* 131, 719–727.
- 1179 Booiij, P., Vethaak, A.D., Leonards, P.E.G., Sjollema, S.B., Kool, J., De Voogt, P., Lamoree, M.H.,
1180 2014. Identification of photosynthesis inhibitors of pelagic marine algae using 96-well
1181 plate microfractionation for enhanced throughput in effect-directed analysis. *Environ. Sci.*
1182 *Technol.* 48, 8003–8011. <https://doi.org/10.1021/es405428t>
- 1183 Botitsi, H. V., Garbis, S.D., Economou, A., Tsipi, D.F., 2011. Current mass spectrometry
1184 strategies for the analysis of pesticides and their metabolites in food and water matrices.
1185 *Mass Spectrom. Rev.* 907–939. <https://doi.org/10.1002/mas>
- 1186 Bourgin, M., Borowska, E., Helbing, J., Hollender, J., Kaiser, H.-P., Kienle, C., Mc Ardell, C.S.,
1187 Simon, E., von Gunten, U., 2017. Effect of operational and water quality parameters on
1188 conventional ozonation and the advanced oxidation process O₃/H₂O₂: Kinetics of
1189 micropollutant abatement, transformation product and bromate formation in a surface
1190 water. *Water Res.* 122, 234–245. <https://doi.org/10.1016/j.watres.2017.05.018>
- 1191 Boxall, A.B.A., Fogg, L.A., Blackwell, P.A., Blackwell, P., Kay, P., Pemberton, E.J., Croxford, A.,
1192 2004. Veterinary Medicines in the Environment, in: *Reviews of Environmental*

- 1193 Contamination and Toxicology. Springer New York, New York, NY, pp. 1–91.
1194 https://doi.org/10.1007/0-387-21729-0_1
- 1195 Boxall, A.B.A., Rudd, M.A., Brooks, B.W., Caldwell, D.J., Choi, K., Hickmann, S., Innes, E.,
1196 Ostapyk, K., Staveley, J.P., Verslycke, T., Ankley, G.T., Beazley, K.F., Belanger, S.E.,
1197 Berninger, J.P., Carriquiriborde, P., Coors, A., DeLeo, P.C., Dyer, S.D., Ericson, J.F., Gagné,
1198 F., Giesy, J.P., Gouin, T., Hallstrom, L., Karlsson, M. V, Larsson, D.G.J., Lazorchak, J.M.,
1199 Mastrocco, F., McLaughlin, A., McMaster, M.E., Meyerhoff, R.D., Moore, R., Parrott, J.L.,
1200 Snape, J.R., Murray-Smith, R., Servos, M.R., Sibley, P.K., Straub, J.O., Szabo, N.D., Topp, E.,
1201 Tetreault, G.R., Trudeau, V.L., Van Der Kraak, G., 2012. Pharmaceuticals and personal care
1202 products in the environment: what are the big questions? *Environ. Heal. Perspect.* 120,
1203 1221–1229. <https://doi.org/http://dx.doi.org/10.1016/j.envint.2013.06.012>
- 1204 Brack, W., 2015. The Challenge: Prioritization of emerging pollutants. *Environ. Toxicol. Chem.*
1205 34, 2181–2182. <https://doi.org/10.1002/etc.3046>
- 1206 Brack, W., Ait-Aissa, S., Burgess, R.M., Busch, W., Creusot, N., Di Paolo, C., Escher, B.I., Mark
1207 Hewitt, L., Hilscherova, K., Hollender, J., Hollert, H., Jonker, W., Kool, J., Lamoree, M.,
1208 Muschket, M., Neumann, S., Rostkowski, P., Ruttkies, C., Schollee, J., Schymanski, E.L.,
1209 Schulze, T., Seiler, T.B., Tindall, A.J., De Aragão Umbuzeiro, G., Vrana, B., Krauss, M., 2016.
1210 Effect-directed analysis supporting monitoring of aquatic environments - An in-depth
1211 overview. *Sci. Total Environ.* 544, 1073–1118.
- 1212 Brack, W., Dulio, V., Agerstrand, M., Allan, I., Altenburger, R., Brinkmann, M., Bunke, D.,
1213 Burgess, R.M., Cousins, I., Escher, B.I., Hernández, F.J., Hewitt, L.M., Hilscherová, K.,
1214 Hollender, J., Hollert, H., Kase, R., Klauer, B., Lindim, C., López Herráez, D., Miège, C.,
1215 Munthe, J., O’Toole, S., Posthuma, L., Rüdél, H., Schäfer, R.B., Sengl, M., Smedes, F., van
1216 de Meent, D., van den Brink, P.J., van Gils, J., van Wezel, A.P., Vethaak, A.D., Vermeirssen,
1217 E., von der Ohe, P.C., Vrana, B., 2017. Towards the review of the European Union Water
1218 Framework Directive: Recommendations for more efficient management of chemical
1219 contamination in European surface water resources. *Sci. Total Environ.* 576, 720–737.
1220 <https://doi.org/10.1016/j.scitotenv.2016.10.104>
- 1221 Brack, W., Dulio, V., Slobodnik, J., 2012. The NORMAN Network and its activities on emerging
1222 environmental substances with a focus on effect-directed analysis of complex
1223 environmental contamination. *Environ. Sci. Eur.* 24, 1–5. <https://doi.org/10.1186/2190-4715-24-29>
- 1225 Bruinen de Bruin, Y., Bessems, J., Fantke, P., van Goetz, N., Schlüter, U., Connolly, A., 2018.
1226 Anchoring Exposure Science in Europe, in: Heinemeijer, G., Jantunen, M., Hakkinen, P.
1227 (Eds.), *The Practice of Consumer Exposure Assessment*. Springer International Publishing
1228 AG., ISBN 987-3-319-96147-7.
- 1229 Buck, R.C., Franklin, J., Berger, U., Conder, J.M., Cousins, I.T., Voogt, P. De, Jensen, A.A.,
1230 Kannan, K., Mabury, S.A., van Leeuwen, S.P.J., 2011. Perfluoroalkyl and polyfluoroalkyl
1231 substances in the environment: Terminology, classification, and origins. *Integr. Environ.*
1232 *Assess. Manag.* 7, 513–541. <https://doi.org/10.1002/ieam.258>
- 1233 Bundschuh, M., Filser, J., Lüderwald, S., McKee, M.S., Metreveli, G., Schaumann, G.E., Schulz,
1234 R., Wagner, S., 2018. Nanoparticles in the environment: where do we come from, where
1235 do we go to? *Environ. Sci. Eur.* 30. <https://doi.org/10.1186/s12302-018-0132-6>
- 1236 Burgess, R.M., Ho, K.T., Brack, W., Lamoree, M., 2013. Effects-directed analysis (EDA) and
1237 toxicity identification evaluation (TIE): Complementary but different approaches for
1238 diagnosing causes of environmental toxicity. *Environ. Toxicol. Chem.* 32, 1935–1945.
1239 <https://doi.org/10.1002/etc.2299>

- 1240 Busch, J., Ahrens, L., Sturm, R., Ebinghaus, R., 2010. Polyfluoroalkyl compounds in landfill
1241 leachates. *Environ. Pollut.* 158, 1467–1471. <https://doi.org/10.1016/j.envpol.2009.12.031>
- 1242 Campanha, M.B., Awan, A.T., de Sousa, D.N.R., Grosseli, G.M., Mozeto, A.A., Fadini, P.S., 2015.
1243 A 3-year study on occurrence of emerging contaminants in an urban stream of São Paulo
1244 State of Southeast Brazil. *Environ. Sci. Pollut. Res.* 22, 7936–7947.
1245 <https://doi.org/10.1007/s11356-014-3929-x>
- 1246 Carstea, E.M., Bridgeman, J., Baker, A., Reynolds, D.M., 2016. Fluorescence spectroscopy for
1247 wastewater monitoring: A review. *Water Res.* 95, 205–219.
1248 <https://doi.org/10.1016/j.watres.2016.03.021>
- 1249 Castiglioni, S., Bijlsma, L., Covaci, A., Emke, E., Hernández, F., Reid, M., Ort, C., Thomas, K. V.,
1250 Van Nuijs, A.L.N., De Voogt, P., Zuccato, E., 2013. Evaluation of uncertainties associated
1251 with the determination of community drug use through the measurement of sewage
1252 drug biomarkers. *Environ. Sci. Technol.* 47, 1452–1460.
1253 <https://doi.org/10.1021/es302722f>
- 1254 Castillo, N.I., Ibáñez, M., Beltrán, E., Rivera-Monroy, J., Ochoa, J.C., Páez-Castillo, M., Posada-
1255 Buitrago, M.L., Sulyok, M., Hernández, F., 2016. Identification of mycotoxins by UHPLC-
1256 QTOF MS in airborne fungi and fungi isolated from industrial paper and antique
1257 documents from the Archive of Bogotá. *Environ. Res.* 144, 130-138.
- 1258 Castrignanò, E., Yang, Z., Bade, R., Baz-Lomba, J.A., Castiglioni, S., Causanilles, A., Covaci, A.,
1259 Gracia-Lor, E., Hernandez, F., Kinyua, J., McCall, A.K., van Nuijs, A.L.N., Ort, C., Plósz, B.G.,
1260 Ramin, P., Rousis, N.I., Ryu, Y., Thomas, K. V., de Voogt, P., Zuccato, E., Kasprzyk-Hordern,
1261 B., 2018. Enantiomeric profiling of chiral illicit drugs in a pan-European study. *Water Res.*
1262 130, 151–160. <https://doi.org/10.1016/j.watres.2017.11.051>
- 1263 Causanilles, A., Emke, E., de Voogt, P., 2016. Determination of phosphodiesterase type V
1264 inhibitors in wastewater by direct injection followed by liquid chromatography coupled to
1265 tandem mass spectrometry. *Sci. Total Environ.* 565, 140–147.
1266 <https://doi.org/10.1016/j.scitotenv.2016.04.158>
- 1267 Chiaia-Hernandez, A.C., Schymanski, E.L., Kumar, P., Singer, H.P., Hollender, J., 2014. Suspect
1268 and nontarget screening approaches to identify organic contaminant records in lake
1269 sediments. *Anal. Bioanal. Chem.* 406, 7323–7335. <https://doi.org/10.1007/s00216-014-8166-0>
1270
- 1271 Covaci, A., Harrad, S., Abdallah, M.A.E., Ali, N., Law, R.J., Herzke, D., de Wit, C.A., 2011. Novel
1272 brominated flame retardants: A review of their analysis, environmental fate and
1273 behaviour. *Environ. Int.* 37, 532–556. <https://doi.org/10.1016/j.envint.2010.11.007>
- 1274 D'Agostino, L.A., Mabury, S.A., 2014. Identification of Novel Fluorinated Surfactants in
1275 Aqueous Film Forming Foams and Commercial Surfactant Concentrates. *Environ. Sci.*
1276 *Technol.* 48, 121–129. <https://doi.org/10.1021/es403729e>
- 1277 Daughton, C.G., 2018. Monitoring wastewater for assessing community health: Sewage
1278 Chemical-Information Mining (SCIM). *Sci. Total Environ.* 619–620, 748–764.
1279 <https://doi.org/10.1016/j.scitotenv.2017.11.102>
- 1280 Daughton, C.G., 2004. Non-regulated water contaminants: emerging research. *Environ. Impact*
1281 *Assess. Rev.* 24, 711–732. <https://doi.org/10.1016/j.eiar.2004.06.003>
- 1282 Daughton, C.G., Ternes, T.A., 1999. Pharmaceuticals and Personal Care Products in the
1283 Environment: Agents of Subtle Change? *Environ. Health Perspect.* 107, 907–938.
1284 <https://doi.org/10.1021/bk-2001-0791>
- 1285 de Hoogh, C.J., Wagenvoort, A.J., Jonker, F., van Leerdam, J.A., Hogenboom, A.C., 2006. HPLC-

1286 DAD and Q-TOF MS Techniques Identify Cause of Daphnia Biomonitor Alarms in the River
1287 Meuse. *Environ. Sci. Technol.* 40, 2678–2685. <https://doi.org/10.1021/es052035a>

1288 de Voogt, P., Sáez, M., 2006. Analytical chemistry of perfluoroalkylated substances. *TrAC -*
1289 *Trends Anal. Chem.* 25, 326–342. <https://doi.org/10.1016/j.trac.2005.10.008>

1290 de Zwart, D., Adams, W., Galay Burgos, M., Hollender, J., Junghans, M., Merrington, G., Muir,
1291 D., Parkerton, T., De Schampelaere, K.A.C., Whale, G., Williams, R., 2018. Aquatic
1292 exposures of chemical mixtures in urban environments: Approaches to impact
1293 assessment. *Environ. Toxicol. Chem.* 37, 703–714. <https://doi.org/10.1002/etc.3975>

1294 Díaz, R., Ibañez, M., Sancho, J.V., Hernández, F., 2012. Target and non-target screening
1295 strategies for organic contaminants, residues and illicit substances in food, environmental
1296 and human biological samples by UHPLC-QTOF-MS. *Anal. Methods* 4, 196–209.

1297 Dimzon, I.K., Westerveld, J., Gremmel, C., Frömel, T., Knepper, T.P., de Voogt, P., 2017.
1298 Sampling and simultaneous determination of volatile per- and polyfluoroalkyl substances
1299 in wastewater treatment plant air and water. *Anal. Bioanal. Chem.* 409, 1395–1404.
1300 <https://doi.org/10.1007/s00216-016-0072-1>

1301 Dionisio, K.L., Phillips, K., Price, P.S., Grulke, C., Williams, A., Biryol, D., Hong, T., Isaacs, K.K.,
1302 2018. The Chemical and Products Database, a resource for exposure-relevant data on
1303 chemicals in consumer products. *Nat. Sci. Data* 5, 180125.
1304 <https://doi.org/10.1038/sdata.2018.125>

1305 Duester, L., Wahrendorf, D.S., Brinkmann, C., Fabricius, A.L., Meermann, B., Pelzer, J., Ecker,
1306 D., Renner, M., Schmid, H., Ternes, T.A., Heininger, P., 2017. A framework to evaluate the
1307 impact of armourstones on the chemical quality of surface water. *PLoS One* 12, 1–12.
1308 <https://doi.org/10.1371/journal.pone.0168926>

1309 Dührkop, K., Shen, H., Meusel, M., Rousu, J., Böcker, S., 2015. Searching molecular structure
1310 databases with tandem mass spectra using CSI:FingerID. *Proc. Natl. Acad. Sci.* 112,
1311 12580–12585. <https://doi.org/10.1073/pnas.1509788112>

1312 Dulio, V., Ohe, P.C. Von Der, James-Casas, A., Roose, P., 2013. Working Group on Prioritisation
1313 of Emerging Substances NORMAN Prioritisation framework for emerging substances.

1314 Dulio, V., Slobodnik, J., 2015. In Response: The NORMAN perspectives on prioritization of
1315 emerging pollutants. *Environ. Toxicol. Chem.* 34, 2183–2184.

1316 EC, 2018. Commission implementing decision(EU) 2018/840 of 5 June 2018 2018, 5–8.

1317 EC, 2013. Directive 2013/39/EU of 12 August 2013 amending Directives 2000/60/EC and
1318 2008/105/EC as regards priority substances in the field of water policy. *Off. J. Eur. Union*
1319 L226, 1–17. [https://doi.org/http://eur-lex.europa.eu/legal-](https://doi.org/http://eur-lex.europa.eu/legal-content/EN/TXT/?uri=celex:32013L0039)
1320 [content/EN/TXT/?uri=celex:32013L0039](https://doi.org/http://eur-lex.europa.eu/legal-content/EN/TXT/?uri=celex:32013L0039)

1321 EC, 2008. Directive 2008/105/EC of 16 December 2008 on environmental quality standards in
1322 the field of water policy, amending and subsequently repealing Council Directives
1323 82/176/EEC, 83/513/EEC, 84/156/EEC, 84/491/ECC,. *Off. J. Eur. Union* L348, 84–97.
1324 <https://doi.org/http://eur-lex.europa.eu/legal-content/EN/TXT/?uri=celex:32008L0105>

1325 EC, 2003. Directive 2002/95/EC of the European Parliament and of the Council of 27 January
1326 2003 on the restriction of the use of certain hazardous substances in electrical and
1327 electronic equipment. *Off. J. Eur. Communities* 4, 42–46.
1328 <https://doi.org/10.1016/j.jclepro.2010.02.014>

1329 EC, 2000. Directive 2000/60/EC of 23 October 2000 establishing a framework for Community
1330 action in the field of water policy. *Off. J. Eur. Parliam.* L327, 1–82.

- 1331 <https://doi.org/10.1039/ap9842100196>
- 1332 EEA, 2013. European Environment Agency. Late lessons from early warnings: science,
1333 precaution, innovation, Report No 1/2013. <https://doi.org/10.2800/73322>
- 1334 Eggen, R.I.L., Hollender, J., Joss, A., Scha, M., 2014. Reducing the Discharge of Micropollutants
1335 in the Aquatic Environment: The Benefits of Upgrading Wastewater Treatment Plants.
1336 *Environ. Sci. Technol.* 48, 7683–7689.
- 1337 EMCDDA, n.d. <http://www.emcdda.europa.eu/wastewater-analysis> [WWW Document]. URL
1338 <http://www.emcdda.europa.eu/wastewater-analysis> (accessed 7.18.18).
- 1339 Emke, E., Evans, S., Kasprzyk-Hordern, B., de Voogt, P., 2014. Enantiomer profiling of high loads
1340 of amphetamine and MDMA in communal sewage: a Dutch perspective. *Sci. Total*
1341 *Environ.* 487, 666–672. <https://doi.org/10.1016/j.scitotenv.2013.11.043>
- 1342 Environment Canada, 2015. Polybrominated diphenyl ethers regulations(SOR/SOR/2008-218),
1343 Online: <http://www.ec.gc.ca/lcpe-cepa/eng/regulations/detailReg.cfm?intReg=108>.
1344 (accessed 14.01.19)
- 1345 Eschauzier, C., Beerendonk, E., Scholte-Veenendaal, P., De Voogt, P., 2012. Impact of
1346 Treatment Processes on the Removal of Perfluoroalkyl Acids from the Drinking Water
1347 Production Chain. *Environ. Sci. Technol.* 46, 1708–1715.
1348 <https://doi.org/10.1021/es201662b>
- 1349 Eschauzier, C., de Voogt, P., Brauch, H., Lange, F., 2011. Perfluorinated compounds in surface
1350 waters, groundwater and drinking water, in: Knepper, T.P. (Ed.), *Handbook of*
1351 *Environmental Chemistry*. Springer, Heidelberg, pp. 73–102.
- 1352 Escher, B.I., Fenner, K., 2011. Recent Advances in Environmental Risk Assessment of
1353 Transformation Products. *Environ. Sci. Technol.* 45, 3835–3847.
1354 <https://doi.org/10.1021/es1030799>
- 1355 European Chemicals Agency (ECHA), 2017. [https://echa.europa.eu/information-on-](https://echa.europa.eu/information-on-chemicals/ec-inventory)
1356 [chemicals/ec-inventory](https://echa.europa.eu/information-on-chemicals/ec-inventory) [WWW Document]. URL [https://echa.europa.eu/information-on-](https://echa.europa.eu/information-on-chemicals/ec-inventory)
1357 [chemicals/ec-inventory](https://echa.europa.eu/information-on-chemicals/ec-inventory) (accessed 5.24.18).
- 1358 Fahrenkamp-Uppenbrink, 2018. Early warning about emerging contaminants. *Science* 11. vol.
1359 360, issue 6389, p. 617. <https://doi.org/10.1111/ajd.12242>
- 1360 Farré, M., Pérez, S., Kantiani, L., 2008. Fate and toxicity of emerging pollutants, their
1361 metabolites and transformation products in the aquatic environment. *TrAC Trends Anal.*
1362 *Chem.* 27, 991–1007. <https://doi.org/10.1016/j.trac.2008.09.010>
- 1363 Farré, M.J., Keller, J., Holling, N., Poussade, Y., Gernjak, W., 2011. Occurrence of N-
1364 nitrosodimethylamine precursors in wastewater treatment plant effluent and their fate
1365 during ultrafiltration-reverse osmosis membrane treatment. *Water Sci. Technol.* 63, 605–
1366 612.
- 1367 Fatta-Kassinos, D., Vasquez, M.I., Kümmerer, K., 2011. Transformation products of
1368 pharmaceuticals in surface waters and wastewater formed during photolysis and
1369 advanced oxidation processes – Degradation, elucidation of byproducts and assessment
1370 of their biological potency. *Chemosphere* 85, 693–709.
1371 <https://doi.org/10.1016/j.chemosphere.2011.06.082>
- 1372 Faust, M., Backhaus, T., 2015. In response: Prioritization and standard setting for pollutant
1373 mixtures in the aquatic environment: A business consultant’s perspective. *Environ.*
1374 *Toxicol. Chem.* 34, 2185–2187. <https://doi.org/10.1002/etc.3046>
- 1375 Fenner, K., Canonica, S., Wackett, L.P., Elsner, M., 2013. Evaluating Pesticide Degradation in

- 1376 the Environment: Blind Spots and Emerging Opportunities. *Science*, 341, 752–758.
1377 <https://doi.org/10.1126/science.1236281>
- 1378 Fuhrer, T., Zamboni, N., 2015. High-throughput discovery metabolomics. *Curr. Opin.*
1379 *Biotechnol.* 31, 73–78. <https://doi.org/10.1016/j.copbio.2014.08.006>
- 1380 Fujioka, T., Khan, S.J., Poussade, Y., Drewes, J.E., Nghiem, L.D., 2012. N-nitrosamine removal by
1381 reverse osmosis for indirect potable water reuse – A critical review based on
1382 observations from laboratory-, pilot- and full-scale studies. *Sep. Purif. Technol.* 98, 503–
1383 515. <https://doi.org/10.1016/j.seppur.2012.07.025>
- 1384 Gago-Ferrero, P., Krettek, A., Fischer, S., Wiberg, K., Ahrens, L., 2018. Suspect screening and
1385 regulatory databases: A powerful combination to identify emerging micropollutants.
1386 *Environ. Sci. Technol.* 52, 6881–6894. <https://doi.org/10.1021/acs.est.7b06598>
- 1387 García-Galán, M.J., Petrovic, M., Rodríguez-Mozaz, S., Barceló, D., 2016. Multiresidue trace
1388 analysis of pharmaceuticals, their human metabolites and transformation products by
1389 fully automated on-line solid-phase extraction-liquid chromatography-tandem mass
1390 spectrometry. *Talanta* 158, 330–341. <https://doi.org/10.1016/j.talanta.2016.05.061>
- 1391 Gebbink, W. A., van Asseldonk, L., van Leeuwen, S. P., 2017. Presence of emerging per-and
1392 polyfluoroalkyl substances (PFASs) in river and drinking water near a fluorochemical
1393 production plant in the Netherlands. *Environ. Sci. Technol.* 51, 11057-11065.
- 1394 Gerecke, A.C., Hartmann, P.C., Heeb, N. V., Kohler, H.P.E., Giger, W., Schmid, P., Zennegg, M.,
1395 Kohler, M., 2005. Anaerobic degradation of decabromodiphenyl ether. *Environ. Sci.*
1396 *Technol.* 39, 1078–1083. <https://doi.org/10.1021/es048634j>
- 1397 Ghoshdastidar, A.J., Fox, S., Tong, A.Z., 2015. The presence of the top prescribed
1398 pharmaceuticals in treated sewage effluents and receiving waters in Southwest Nova
1399 Scotia, Canada. *Environ. Sci. Pollut. Res.* 22, 689–700. [https://doi.org/10.1007/s11356-](https://doi.org/10.1007/s11356-014-3400-z)
1400 [014-3400-z](https://doi.org/10.1007/s11356-014-3400-z)
- 1401 Gómez Ramos, M.J., Lozano, A., Fernández-Alba, A.R., 2019. High-resolution mass
1402 spectrometry with data independent acquisition for the comprehensive non-targeted
1403 analysis of migrating chemicals coming from multilayer plastic packaging materials used
1404 for fruit purée and juice. *Talanta* 191, 180-192.
- 1405 González-Mariño, I., Rodil, R., Barrio, I., Cela, R., Quintana, J.B., 2017. Wastewater-Based
1406 Epidemiology as a New Tool for Estimating Population Exposure to Phthalate Plasticizers.
1407 *Environ. Sci. Technol.* 51, 3902–3910. <https://doi.org/10.1021/acs.est.6b05612>
- 1408 Gosetti, F., Mazzucco, E., Gennaro, M.C., Marengo, E., 2016. Contaminants in water : non-
1409 target UHPLC / MS analysis. *Environ. Chem. Lett.* 14, 51–65.
- 1410 Gracia-Lor, E., Rousis, N.I., Hernández, F., Zuccato, E., Castiglioni, S., 2018. Wastewater-Based
1411 Epidemiology as a Novel Biomonitoring Tool to Evaluate Human Exposure To Pollutants.
1412 *Environ. Sci. Technol.* 52, 10224-10226.
- 1413 Gracia-Lor, E., Sancho, J. V., Serrano, R., Hernández, F., 2012. Occurrence and removal of
1414 pharmaceuticals in wastewater treatment plants at the Spanish Mediterranean area of
1415 Valencia. *Chemosphere* 87, 453–462.
1416 <https://doi.org/10.1016/j.chemosphere.2011.12.025>
- 1417 Gros, M., Petrović, M., Ginebreda, A., Barceló, D., 2010. Removal of pharmaceuticals during
1418 wastewater treatment and environmental risk assessment using hazard indexes. *Environ.*
1419 *Int.* 36, 15–26. <https://doi.org/10.1016/j.envint.2009.09.002>
- 1420 Guillén, D., Ginebreda, A., Farré, M., Darbra, R.M., Petrovic, M., Gros, M., Barceló, D., 2012.

- 1421 Prioritization of chemicals in the aquatic environment based on risk assessment :
1422 Analytical, modeling and regulatory perspective. *Sci. Total Environ.* 440, 236–252.
- 1423 Hajibabania, S., Verliefde, A., McDonald, J.A., Khan, S.J., Le-Clech, P., 2011. Fate of trace
1424 organic compounds during treatment by nanofiltration. *J. Memb. Sci.* 373, 130–139.
1425 <https://doi.org/10.1016/j.memsci.2011.02.040>
- 1426 Heiss, C., Küster, A., 2015. In Response: A regulatory perspective on prioritization of emerging
1427 pollutants in the context of the Water Framework Directive. *Environ. Toxicol. Chem.* 34,
1428 2181–2182.
- 1429 Helbling, D.E., Hollender, J., Kohler, H.-P.E., Singer, H., Fenner, K., 2010. High-Throughput
1430 Identification of Microbial Transformation Products of Organic Micropollutants. *Environ.*
1431 *Sci. Technol.* 44, 6621–6627. <https://doi.org/10.1021/es100970m>
- 1432 Hernández, F., Castiglioni, S., Covaci, A., De Voogt, P., Emke, E., Kasprzyk-Hordern, B., Ort, C.,
1433 Reid, M., Sancho, J.V., Thomas, K.V., van Nuijs, A.L.N., Zuccato, E., Bijlsma, L., 2018. Mass
1434 Spectrometric strategies for the investigation of biomarkers of illicit drug use in
1435 wastewater. *Mass Spectrom. Rev.* 37, 258–280. <https://doi.org/10.1002/mas.21525>
- 1436 Hernández, F., Cervera, M.I., Portolés, T., Beltrán, J., Pitarch, E., 2013. The role of GC-MS/MS
1437 with triple quadrupole in pesticide residue analysis in food and the environment. *Anal.*
1438 *Methods* 5, 5875–5894. <https://doi.org/10.1039/c3ay41104d>
- 1439 Hernández, F., Ibáñez, M., Bade, R., Bijlsma, L., Sancho, J.V., 2014. Investigation of
1440 pharmaceuticals and illicit drugs in waters by liquid chromatography-high-resolution
1441 mass spectrometry. *TrAC Trends Anal. Chem.* 63, 140–157.
- 1442 Hernández, F., Ibáñez, M., Botero-Coy, A.M., Bade, R., Bustos-Lopez, M.C., Rincon, J.,
1443 Moncayo, A., Bijlsma, L., 2015a. LC-QTOF MS screening of more than 1,000 licit and illicit
1444 drugs and their metabolites in wastewater and surface waters from the area of Bogotá,
1445 Colombia. *Anal. Bioanal. Chem.* 407, 6405–6416. <https://doi.org/10.1007/s00216-015-8796-x>
1446
- 1447 Hernández, F., Ibáñez, M., Gracia-Lor, E., Sancho, J.V., 2011. Retrospective LC-QTOF-MS
1448 analysis searching for pharmaceutical metabolites in urban wastewater. *J. Sep. Sci.* 34,
1449 3517–3526.
- 1450 Hernández, F., Ibáñez, M., Portolés, T., Cervera, M.I., Sancho, J. V, López, F.J., 2015b.
1451 Advancing towards universal screening for organic pollutants in waters. *J. Hazard. Mater.*
1452 282, 86–95.
- 1453 Hernández, F., Sancho, J.V., Ibáñez, M., Abad, E., Portolés, T., Mattioli, L., 2012. Current use of
1454 high-resolution mass spectrometry in the environmental sciences. *Anal. Bioanal. Chem.*
1455 403, 1251–1264.
- 1456 Heydebreck, F., Tang, J., Xie, Z., Ebinghaus, R., 2015. Alternative and Legacy Perfluoroalkyl
1457 Substances: Differences between European and Chinese River/Estuary Systems. *Environ.*
1458 *Sci. Technol.* 49, 8386–8395. doi:10.1021/acs.est.5b01648
- 1459 HighChem, 2016. mzCloud Mass Spectral Library [WWW Document]. URL
1460 <https://www.mzcloud.org/> (accessed 5.30.18).
- 1461 Hogenboom, A.C., van Leerdam, J.A., de Voogt, P., 2009. Accurate mass screening and
1462 identification of emerging contaminants in environmental samples by liquid
1463 chromatography-hybrid linear ion trap Orbitrap mass spectrometry. *J. Chromatogr. A*
1464 1216, 510–519. <https://doi.org/10.1016/j.chroma.2008.08.053>
- 1465 Hoh, E., Dodder, N.G., Lehotay, S.J., Pangallo, K.C., Reddy, C.M., Maruya, K.A., 2012.

1466 Nontargeted Comprehensive Two-Dimensional Gas Chromatography/Time-of-Flight Mass
1467 Spectrometry Method and Software for Inventorying Persistent and Bioaccumulative
1468 Contaminants in Marine Environments. *Environ. Sci. Technol.* 46, 8001–8008.
1469 <https://doi.org/10.1021/es301139q>

1470 Hollender, J., Schymanski, E.L., Singer, H., Ferguson, P.L., 2017. Non-target screening with high
1471 resolution mass spectrometry in the environment: Ready to go? *Environ. Sci. Technol.*
1472 *acs.est.7b02184*. <https://doi.org/10.1021/acs.est.7b02184>

1473 Howard, P.H., Muir, D.C.G., 2010. Identifying New Persistent and Bioaccumulative Organics
1474 Among Chemicals in Commerce. *Environ. Sci. Technol.* 44, 2277–2285.
1475 <https://doi.org/10.1021/es903383a>

1476 Hu, S., Zhao, M., Xi, Y., Mao, Q., Zhou, X., Chen, D., Yan, P., 2017. Nontargeted Screening and
1477 Determination of Sulfonamides: A Dispersive Micro Solid-Phase Extraction Approach to
1478 the Analysis of Milk and Honey Samples Using Liquid Chromatography-High-Resolution
1479 Mass Spectrometry. *J. Agric. Food. Chem.* 65, 1984–1991

1480 Hu, X.C., Andrews, D.Q., Lindstrom, A.B., Bruton, T.A., Schaidler, L.A., Grandjean, P., Lohmann,
1481 R., Carignan, C.C., Blum, A., Balan, S.A., Higgins, C.P., Sunderland, E.M., 2016. Detection of
1482 Poly- and Perfluoroalkyl Substances (PFASs) in U.S. Drinking Water Linked to Industrial
1483 Sites, Military Fire Training Areas, and Wastewater Treatment Plants. *Environ. Sci.*
1484 *Technol. Lett.* 3, 344–350. <https://doi.org/10.1021/acs.estlett.6b00260>

1485 Ibáñez, M., Borova, V., Boix, C., Aalizadeh, R., Bade, R., Thomaidis, N.S., Hernández, F., 2017.
1486 UHPLC-QTOF MS screening of pharmaceuticals and their metabolites in treated
1487 wastewater samples from Athens. *J. Hazard. Mater.* 323, 26–35.
1488 <https://doi.org/10.1016/j.jhazmat.2016.03.078>

1489 Imen, S., Chang, N.-B., Yang, Y.J., 2015. Developing the remote sensing-based early warning
1490 system for monitoring TSS concentrations in Lake Mead. *J. Environ. Manage.* 160, 73–89.
1491 <https://doi.org/10.1016/j.jenvman.2015.06.003>

1492 ISES-Europe: the European branch of the International Society of Exposure Science [WWW
1493 Document], n.d. . <https://ises-europe.org>. URL <https://ises-europe.org>. (accessed
1494 7.18.18).

1495 Jeffryes, J.G., Colastani, R.L., Elbadawi-Sidhu, M., Kind, T., Niehaus, T.D., Broadbelt, L.J.,
1496 Hanson, A.D., Fiehn, O., Tyo, K.E.J., Henry, C.S., 2015. MINEs: Open access databases of
1497 computationally predicted enzyme promiscuity products for untargeted metabolomics. *J.*
1498 *Cheminform.* 7, 1–8. <https://doi.org/10.1186/s13321-015-0087-1>

1499 Jelic, A., Gros, M., Ginebreda, A., Cespedes-Sánchez, R., Ventura, F., Petrovic, M., Barcelo, D.,
1500 2011. Occurrence, partition and removal of pharmaceuticals in sewage water and sludge
1501 during wastewater treatment. *Water Res.* 45, 1165–1176.
1502 <https://doi.org/10.1016/j.watres.2010.11.010>

1503 Jiang, J.Q., 2018. Occurrence of microplastics and its pollution in the environment: A review.
1504 *Sustain. Prod. Consum.* 13, 16–23. <https://doi.org/10.1016/j.spc.2017.11.003>

1505 Jonker, W., Clarijs, B., de Witte, S.L., van Velzen, M., de Koning, S., Schaap, J., Somsen, G.W.,
1506 Kool, J., 2016. Gas chromatography fractionation platform featuring parallel flame-
1507 ionization detection and continuous high-resolution analyte collection in 384-well plates.
1508 *J. Chromatogr. A* 1462, 100–106. <https://doi.org/10.1016/j.chroma.2016.07.068>

1509 Jonker, W., Lamoree, M.H., Houtman, C.J., Hamers, T., Somsen, G.W., Kool, J., 2015. Rapid
1510 activity-directed screening of estrogens by parallel coupling of liquid chromatography
1511 with a functional gene reporter assay and mass spectrometry. *J. Chromatogr. A* 1406,

- 1512 165–174. <https://doi.org/10.1016/j.chroma.2015.06.012>
- 1513 Jonker, W., Zwart, N., Stöckl, J.B., de Koning, S., Schaap, J., Lamoree, M.H., Somsen, G.W.,
1514 Hamers, T., Kool, J., 2017. Continuous fraction collection of gas chromatographic
1515 separations with parallel mass spectrometric detection applied to cell-based bioactivity
1516 analysis. *Talanta* 168, 162–167. <https://doi.org/10.1016/j.talanta.2017.02.067>
- 1517 Kasprzyk-Hordern, B., Baker, D.R., 2012. Estimation of community-wide drugs use via
1518 stereoselective profiling of sewage. *Sci. Total Environ.* 423, 142–150.
1519 <https://doi.org/10.1016/j.scitotenv.2012.02.019>
- 1520 Kasprzyk-Hordern, B., Bijlsma, L., Castiglioni, S., Covaci, A., de Voogt, P., Emke, E., Hernandez,
1521 F., Ort, C., Reid, M.J., van Nuijs, A.L.N., Thomas, K. V., 2014. Wastewater-based
1522 epidemiology - the concept and historical perspectives. *Environ. Ind. Mag.* 33, 145–151.
- 1523 Kern, S., Baumgartner, R., Helbling, D.E., Hollender, J., Singer, H., Loos, M.J., Schwarzenbach,
1524 R.P., Fenner, K., 2010. A tiered procedure for assessing the formation of
1525 biotransformation products of pharmaceuticals and biocides during activated sludge
1526 treatment. *J. Environ. Monit.* 12, 2100–2111. <https://doi.org/10.1039/C0EM00238K>
- 1527 Kern, S., Fenner, K., Singer, H.P., Schwarzenbach, R.P., Hollender, J., 2009. Identification of
1528 transformation products of organic contaminants in natural waters by computer-aided
1529 prediction and high-resolution mass spectrometry. *Environ. Sci Technol* 43, 7039–7046.
1530 <https://doi.org/10.1021/es901979h>
- 1531 Kinyua J., Negreira, N., Ibáñez, M., Bijlsma, L., Hernández, F., Covaci, A., Van Nuijs, A.L.N., 2015.
1532 A data-independent acquisition workflow for qualitative screening of new psychoactive
1533 substances in biological samples. *Anal. Bioanal. Chem.* 407, 8773–8785.
- 1534 Kolkman, A., Martijn, B.J., Vughs, D., Baken, K.A., van Wezel, A.P., 2015. Tracing Nitrogenous
1535 Disinfection Byproducts after Medium Pressure UV Water Treatment by Stable Isotope
1536 Labeling and High Resolution Mass Spectrometry. *Environ. Sci. Technol.* 49, 4458–4465.
1537 <https://doi.org/10.1021/es506063h>
- 1538 Kolpin, D.W., Furlong, E.T., Meyer, M.T., Thurman, E.M., Zaugg, S.D., Barber, L.B., Buxton, H.T.,
1539 2002. Pharmaceuticals, Hormones, and Other Organic Wastewater Contaminants in U.S.
1540 Streams, 1999–2000: A National Reconnaissance. *Environ. Sci. Technol.* 36, 1202–1211.
1541 <https://doi.org/10.1021/es011055j>
- 1542 Kotthoff, M., Bücking, M., 2018. Four Chemical Trends Will Shape the Next Decade's
1543 Directions in Perfluoroalkyl and Polyfluoroalkyl Substances. *Research Front. Chem.* 6,
1544 103.
- 1545 Kuantama, E., Saraswati, M., 2015. Water level measurement and pre-flood warning system
1546 with SMS method. *Internetworking Indones. J.* 7, 1–7.
- 1547 Kubo, T., Otsuka, K., 2016. Recent progress in molecularly imprinted media by new preparation
1548 concepts and methodological approaches for selective separation of targeting
1549 compounds, *Trends Anal. Chem.* 81, 102–109
- 1550 Lau, C., Anitole, K., Hodes, C., Lai, D., Pfahles-Hutchens, A., Seed, J., 2007. Perfluoroalkyl acids:
1551 A review of monitoring and toxicological findings. *Toxicol. Sci.* 99, 366–394.
1552 <https://doi.org/10.1093/toxsci/kfm128>
- 1553 Law, R.J., Covaci, A., Harrad, S., Herzke, D., Abdallah, M.A., Fernie, K., Toms, L.L., Takigami, H.,
1554 2014. Levels and trends of PBDEs and HBCDs in the global environment : Status at the
1555 end of 2012. *Environ. Int.* 65, 147–158. <https://doi.org/10.1016/j.envint.2014.01.006>
- 1556 Letzel, T., Bayer, A., Schulz, W., Heermann, A., Lucke, T., Greco, G., Grosse, S., Schüssler, W.,

Con formato: Español

Con formato: Español

- 1557 Sengl, M., Letzel, M., 2015. LC – MS screening techniques for wastewater analysis and
1558 analytical data handling strategies : Sartans and their transformation products as an
1559 example. *Chemosphere* 137, 198–206.
- 1560 Li, Y.F., Sun, Y.M., Beier R.C., Lei, H.T., Gee, S., Hammock, B.D., Wang, H., Wang, Z., Sun, X.,
1561 Shen, Y.D., Yang, J.Y., Xu, Z.L., 2017. Immunochemical techniques for multianalyte
1562 analysis of chemical residues in food and the environment: A review. *Trends Anal. Chem.*
1563 88, 25-40
- 1564 Li, J., Yu, N., Zhang, B., Jin, L., Li, M., Hu, M., Zhang, X., Wei, S., Yu, H., 2014. Occurrence of
1565 organophosphate flame retardants in drinking water from China. *Water Res.* 54, 53–61.
1566 <https://doi.org/10.1016/j.watres.2014.01.031>
- 1567 Lindstrom, A.B., Strynar, M.J., Libelo, E.L., 2011. Polyfluorinated Compounds: Past, Present,
1568 and Future. *Environ. Sci. Technol.* 45, 7954–7961. <https://doi.org/10.1021/es2011622>
- 1569 Liu, J., Lu, G., Xie, Z., Zhang, Z., Li, S., Yan, Z., 2015. Occurrence, bioaccumulation and risk
1570 assessment of lipophilic pharmaceutically active compounds in the downstream rivers of
1571 sewage treatment plants. *Sci. Total Environ.* 511, 54–62.
1572 <https://doi.org/10.1016/j.scitotenv.2014.12.033>
- 1573 Liu, Y., Pereira, A.D.S., Martin, J.W., 2015. Discovery of C5–C17 Poly- and Perfluoroalkyl
1574 Substances in Water by In-Line SPE-HPLC-Orbitrap with In-Source Fragmentation
1575 Flagging. *Anal. Chem.* 87, 4260–4268. <https://doi.org/10.1021/acs.analchem.5b00039>
- 1576 Loos, M.J., Ruff, M., Singer, H.P., 2016. enviMass version 3.0 screening software [WWW
1577 Document]. URL <https://github.com/blosloos/enviMass> (accessed 5.30.17).
- 1578 Loos, R., Carvalho, R., António, D.C., Comero, S., Locoro, G., Tavazzi, S., Paracchini, B., Ghiani,
1579 M., Lettieri, T., Blaha, L., Jarosova, B., Voorspoels, S., Servaes, K., Haglund, P., Fick, J.,
1580 Lindberg, R.H., Schwesig, D., Gawlik, B.M., 2013. EU-wide monitoring survey on emerging
1581 polar organic contaminants in wastewater treatment plant effluents. *Water Res.* 47,
1582 6475–6487.
- 1583 Loos, R., Gawlik, B.M., Locoro, G., Rimaviciute, E., Contini, S., Bidoglio, G., 2009. EU-wide
1584 survey of polar organic persistent pollutants in European river waters. *Environ. Pollut.*
1585 157, 561–568.
- 1586 Lopardo, L., Cummins, A., Rydevik, A., Kasprzyk-Hordern, B., 2017. New Analytical Framework
1587 for Verification of Biomarkers of Exposure to Chemicals Combining Human Biomonitoring
1588 and Water Fingerprinting. *Anal. Chem.* 89, 7232–7239.
1589 <https://doi.org/10.1021/acs.analchem.7b01527>
- 1590 Lopardo, L., Smith, A., Rydevik, A., Kasprzyk-Hordern, B., 2018. Verifying community-wide
1591 exposure to antimicrobial agents in personal care products – in quest for metabolic
1592 biomarkers of exposure via in-vitro studies and wastewater-based epidemiology. *Water*
1593 *Res.* 143, 117–126. <https://doi.org/10.1016/j.watres.2018.06.028>
- 1594 Lorenzo, M., Campo, J., Picó, Y., 2018. Analytical challenges to determine emerging persistent
1595 organic pollutants in aquatic ecosystems. *TRAC Trends Anal. Chem.* 103, 137-155.
- 1596 Lucattini, P., Lamoree, M.H., Leonards, P.E.G., de Boer, J., 2017. Non-target and suspect
1597 screening of flame retardants and other compounds in indoor dust from five countries,
1598 in: 8th International Symposium on Flame Retardants. York, UK.
- 1599 Malaj, E., Ohe, P.C. Von Der, Grote, M., Kühne, R., Mondy, C.P., Usseglio-polatera, P., Brack,
1600 W., Schäfer, R.B., 2014. Organic chemicals jeopardize the health of freshwater
1601 ecosystems on the continental scale. *PNAS* 111, 9549–9554.

Con formato: Inglés (americano)

- 1602 McCleaf, P., Englund, S., Östlund, A., Lindegren, K., Wiberg, K., Ahrens, L., 2017. Removal
1603 efficiency of multiple poly- and perfluoroalkyl substances (PFASs) in drinking water using
1604 granular activated carbon (GAC) and anion exchange (AE) column tests. *Water Res.* 120,
1605 77–87. <https://doi.org/10.1016/j.watres.2017.04.057>
- 1606 McEachran, A.D., Mansouri, K., Newton, S.R., Beverly, B.E.J., Sobus, J.R., Williams, A.J., 2018. A
1607 comparison of three liquid chromatography (LC) retention time prediction models.
1608 *Talanta* 182, 371–379. <https://doi.org/10.1016/j.talanta.2018.01.022>
- 1609 Merel, S., Lege, S., Yanez Heras, J.E., Zwiener, C., 2017. Assessment of N-Oxide Formation
1610 during Wastewater Ozonation. *Environ. Sci. Technol.* 51, 410–417.
1611 <https://doi.org/10.1021/acs.est.6b02373>
- 1612 Miller, T.H., Gallidabino, M.D., MacRae, J.I., Hogstrand, C., Bury, N.R., Barron, L.P., Snape, J.R.,
1613 Owen, S.F., 2018. Machine Learning for Environmental Toxicology: A call for integration
1614 and innovation. *Environ. Sci. Technol.* 52, 12953–12955.
- 1615 Miller, T.H., Musenga, A., Cowan, D. a, Barron, L.P., 2013. Prediction of Chromatographic
1616 Retention Time in HighResolution Anti-Doping Screening Data Using Artificial Neural
1617 Networks. *Anal. Chem.* 85, 10330–10337.
- 1618 Miyake, Y., Yamashita, N., Rostkowski, P., So, M.K., Taniyasu, S., Lam, P.K.S., Kannan, K., 2007.
1619 Determination of trace levels of total fluorine in water using combustion ion
1620 chromatography for fluorine: A mass balance approach to determine individual
1621 perfluorinated chemicals in water. *J. Chromatogr. A* 1143, 98–104.
1622 <https://doi.org/10.1016/j.chroma.2006.12.071>
- 1623 Mollerup, C.B., Mardal, M., Dalsgaard, P.W., Linnet, K., Barron, L.P., 2018. Prediction of
1624 collision cross section and retention time for broad scope screening in gradient reversed-
1625 phase liquid chromatography-ion mobility-high resolution accurate mass spectrometry. *J.*
1626 *Chromatogr. A* 1542, 82–88. <https://doi.org/10.1016/j.chroma.2018.02.025>
- 1627 Montes, R., Aguirre, J., Vidal, X., Rodil, R., Cela, R., Quintana, J.B., 2017. Screening for Polar
1628 Chemicals in Water by Trifunctional Mixed-Mode Liquid Chromatography-High Resolution
1629 Mass Spectrometry. *Environ. Sci. Technol.* 51, 6250–6259.
1630 <https://doi.org/10.1021/acs.est.6b05135>
- 1631 Moschet, C., Piazzoli, A., Singer, H., Hollender, J., 2013. Alleviating the reference standard
1632 dilemma using a systematic exact mass suspect screening approach with liquid
1633 chromatography-high resolution mass spectrometry. *Anal. Chem.* 85, 10312–10320.
- 1634 Moschet, C., Wittmer, I., Simovic, J., Junghans, M., Piazzoli, A., Singer, H., Stamm, C., Leu, C.,
1635 Hollender, J., 2014. How a Complete Pesticide Screening Changes the Assessment of
1636 Surface Water Quality. *Environ. Sci. Technol.* 48, 5423–5432.
1637 <https://doi.org/10.1021/es500371t>
- 1638 Müller, A., Schulz, W., Ruck, W.K.L., Weber, W.H., 2011. A new approach to data evaluation in
1639 the non-target screening of organic trace substances in water analysis. *Chemosphere* 85,
1640 1211–1219. <https://doi.org/10.1016/j.chemosphere.2011.07.009>
- 1641 Muz, M., Dann, J.P., Jäger, F., Brack, W., Krauss, M., 2017. Identification of Mutagenic Aromatic
1642 Amines in River Samples with Industrial Wastewater Impact. *Environ. Sci. Technol.* 4681–
1643 4688. <https://doi.org/10.1021/acs.est.7b00426>
- 1644 Norman network, 2018a. Norman Suspect List Exchange [WWW Document]. URL
1645 <http://www.norman-network.com/?q=node/236> (accessed 5.30.18).
- 1646 Norman network, 2018b. The Norman Early Warning System (NormaNEWS) [WWW
1647 Document]. URL <http://www.normandata.eu/?q=node/244> (accessed 5.30.18).

- 1648 Nürenberg, G., Schulz, M., Kunkel, U., Ternes, T.A., 2015. Development and validation of a
1649 generic nontarget method based on liquid chromatography – high resolution mass
1650 spectrometry analysis for the evaluation of different wastewater treatment options. *J.*
1651 *Chromatogr. A* 1426, 77–90.
- 1652 Nurmi, J., Pellinen, J., Rantalainen, A., 2012. Critical evaluation of screening techniques for
1653 emerging environmental contaminants based on accurate mass measurements with
1654 time-of-flight mass spectrometry. *J. Mass Spectrom.* 47, 303–312.
- 1655 Ordoñez, E.Y., Quintana, J.B., Rodil, R., Cela, R., 2012. Computer assisted optimization of liquid
1656 chromatographic separations of small molecules using mixed-mode stationary phases. *J.*
1657 *Chromatogr. A* 1238, 91–104. <https://doi.org/10.1016/j.chroma.2012.03.055>
- 1658 Ort, C., van Nuijs, A.L.N., Berset, J.D., Bijlsma, L., Castiglioni, S., Covaci, A., de Voogt, P., Emke,
1659 E., Fatta-Kassinos, D., Griffiths, P., Hernández, F., González-Mariño, I., Grabic, R.,
1660 Kasprzyk-Hordern, B., Mastroianni, N., Meierjohann, A., Nefau, T., Östman, M., Pico, Y.,
1661 Racamonde, I., Reid, M., Slobodnik, J., Terzic, S., Thomaidis, N., Thomas, K. V., 2014.
1662 Spatial differences and temporal changes in illicit drug use in Europe quantified by
1663 wastewater analysis. *Addiction* 109, 1338–1352. <https://doi.org/10.1111/add.12570>
- 1664 Ouyang, X., Froment, J., Leonards, P.E.G., Christensen, G., Tollefsen, K.-E., de Boer, J., Thomas,
1665 K. V., Lamoree, M.H., 2017. Miniaturization of a transthyretin binding assay using a
1666 fluorescent probe for high throughput screening of thyroid hormone disruption in
1667 environmental samples. *Chemosphere* 171, 722–728.
1668 <https://doi.org/10.1016/j.chemosphere.2016.12.119>
- 1669 Ouyang, X., Leonards, P.E.G., Touseva, Z., Slobodnik, J., De Boer, J., Lamoree, M.H., 2016. Rapid
1670 Screening of Acetylcholinesterase Inhibitors by Effect-Directed Analysis Using LC x LC
1671 Fractionation, a High Throughput in Vitro Assay, and Parallel Identification by Time of
1672 Flight Mass Spectrometry. *Anal. Chem.* 88, 2353–2360.
1673 <https://doi.org/10.1021/acs.analchem.5b04311>
- 1674 Peisl, B.Y.L., Schymanski, E.L., Wilmes, P., 2018. Dark matter in host-microbiome
1675 metabolomics: Tackling the unknowns-A review. *Anal. Chim. Acta* in press.
1676 <https://doi.org/10.1016/j.aca.2017.12.034>
- 1677 Petrie, B., Youdan, J., Barden, R., Kasprzyk-Hordern, B., 2016. New Framework To Diagnose the
1678 Direct Disposal of Prescribed Drugs in Wastewater – A Case Study of the Antidepressant
1679 Fluoxetine. *Environ. Sci. Technol.* 50, 3781–3789.
1680 <https://doi.org/10.1021/acs.est.6b00291>
- 1681 Picó, Y., Alfarham, A., Barceló, D., 2017. Analysis of emerging contaminants and nanomaterials
1682 in plant materials following uptake from soils. *TRAC Trends Anal. Chem.* 94, 173-189.
- 1683 Picó, Y., Alfarham, A., Barceló, D., 2018. Nano- and microplastic analysis: Focus on their
1684 occurrence in freshwater ecosystems and remediation technologies. *TRAC Trends Anal.*
1685 *Chem.* in press. <https://doi.org/10.1016/j.trac.2018.08.022>
- 1686 Pieke, E., Heus, F., Kamstra, J.H., Mladic, M., Velzen, M. Van, Kamminga, D., Lamoree, M.H.,
1687 Hamers, T., Leonards, P., Niessen, W.M.A., Kool, J., 2013. High-resolution fractionation
1688 after gas chromatography for effect-directed analysis. *Anal. Chem.* 85, 8204–8211.
1689 <https://doi.org/10.1021/ac401384q>
- 1690 Pinhancos, R., Maass, S., Ramanathan, D.M., 2011. High-resolution mass spectrometry method
1691 for the detection, characterization and quantitation of pharmaceuticals in water. *J. Mass*
1692 *Spectrom.* 46, 1175–1181. <https://doi.org/10.1002/jms.2005>
- 1693 Pitarch, E., Portoles, T., Marin, J.M., Ibañez, M., Albarran, F., Hernandez, F., 2010. Analytical

1694 strategy based on the use of liquid chromatography and gas chromatography with triple-
1695 quadrupole and time-of-flight MS analyzers for investigating organic contaminants in
1696 wastewater. *Anal. Bioanal. Chem.* 397, 2763–2776. [https://doi.org/10.1007/s00216-010-](https://doi.org/10.1007/s00216-010-3692-x)
1697 3692-x

1698 Pluskal, T., Castillo, S., Villar-Briones, A., Orešič, M., 2010. MZmine 2: Modular framework for
1699 processing, visualizing, and analyzing mass spectrometry-based molecular profile data.
1700 *BMC Bioinformatics* 11, 395. <https://doi.org/10.1186/1471-2105-11-395>

1701 Portolés, T., Mol, J.G.J., Sancho, J. V., Hernández, F., 2014. Use of electron ionization and
1702 atmospheric pressure chemical ionization in gas chromatography coupled to time-of-
1703 flight mass spectrometry for screening and identification of organic pollutants in waters.
1704 *J. Chromatogr. A* 1339, 145–153.

1705 Portolés, T., Sales, C., Gómara, B., Sancho, J.V., Beltran, J., Herrero, L., González, M.J.,
1706 Hernandez, F., 2015. Novel Analytical Approach for Brominated Flame Retardants Based
1707 on the Use of Gas Chromatography-Atmospheric Pressure Chemical Ionization-Tandem
1708 Mass Spectrometry with Emphasis in Highly Brominated Congeners. *Anal. Chem.* 87,
1709 9892–9899. <https://doi.org/10.1021/acs.analchem.5b02378>

1710 Rager, J.E., Strynar, M.J., Liang, S., McMahan, R.L., Richard, A.M., Grulke, C.M., Wambaugh,
1711 J.F., Isaacs, K.K., Judson, R., Williams, A.J., Sobus, J.R., 2016. Linking high resolution mass
1712 spectrometry data with exposure and toxicity forecasts to advance high-throughput
1713 environmental monitoring. *Environ. Int.* 88, 269–280.

1714 Reemtsma, T., Berger, U., Arp, H.P.H., Gallard, H., Knepper, T.P., Neumann, M., Quintana, J.B.,
1715 Voogt, P. de, 2016. Mind the Gap: Persistent and Mobile Organic Compounds—Water
1716 Contaminants That Slip Through. *Environ. Sci. Technol.* 50, 10308–10315.
1717 <https://doi.org/10.1021/acs.est.6b03338>

1718 Reemtsma, T., García-López, M., Rodríguez, I., Quintana, J.B., Rodil, R., 2008.
1719 Organophosphorus flame retardants and plasticizers in water and air I. Occurrence and
1720 fate. *TRAC - Trends Anal. Chem.* 27, 727–737. <https://doi.org/10.1016/j.trac.2008.07.002>

1721 Richardson, L.A., 2017. Understanding and overcoming antibiotic resistance. *PLoS Biol.* 15, 1–5.
1722 <https://doi.org/10.1371/journal.pbio.2003775>

1723 Richardson, M.L., Bowron, J.M., 1985. The fate of pharmaceutical chemicals in the aquatic
1724 environment. *J. Pharm. Pharmacol.* 37, 1–12. [https://doi.org/10.1111/j.2042-](https://doi.org/10.1111/j.2042-7158.1985.tb04922.x)
1725 7158.1985.tb04922.x

1726 Richardson, S.D., Kimura, S.Y., 2016. Water Analysis: Emerging Contaminants and Current
1727 Issues. *Anal. Chem.* 88, 546–582. <https://doi.org/10.1021/acs.analchem.5b04493>

1728 Richardson, S.D., Plewa, M.J., Wagner, E.D., Schoeny, R., DeMarini, D.M., 2007. Occurrence,
1729 genotoxicity, and carcinogenicity of regulated and emerging disinfection by-products in
1730 drinking water: A review and roadmap for research. *Mutat. Res. Mutat. Res.* 636, 178–
1731 242. <https://doi.org/10.1016/j.mrrev.2007.09.001>

1732 Ritter, S.K., 2015. The Shrinking Case For Fluorochemicals. *Chem. Eng. News Arch.* 93, 27–29.
1733 <https://doi.org/10.1021/cen-09328-scitech1>

1734 Rockström, J., Steffen, W., Noone, K., Persson, Å., Chapin, F.S.I., Lambin, E., Lenton, T.M.,
1735 Scheffer, M., Folke, C., Schellnhuber, H.J., Nykvist, B., Wit, C.A. De, Hughes, T., Leeuw, S.
1736 Van Der, Rodhe, H., Sörlin, S., Snyder, P.K., Costanza, R., Svedin, U., Falkenmark, M.,
1737 Karlberg, L., Corell, R.W., Fabry, V.J., Hansen, J., Walker, B., Liverman, D., Richardson, K.,
1738 Crutzen, P., Foley, J., 2009a. Planetary Boundaries : Exploring the Safe Operating Space
1739 for Humanity. *Ecol. Soc.* 14(2).

- 1740 Rockström, J., Steffen, W., Noone, K., Persson, Å., Chapin, F.S.I., Lambin, E.F., Lenton, T.M.,
1741 Scheffer, M., Folke, C., Schellnhuber, H.J., Nykvist, B., Wit, C.A. de, Hughes, T., Leeuw, S.
1742 van der, Rodhe, H., Sörlin, S., Snyder, P.K., Costanza, R., Svedin, U., Falkenmark, M.,
1743 Karlberg, L., Corell, R.W., Fabry, V.J., Hansen, J., Walker, B., Liverman, D., Richardson, K.,
1744 Crutzen, P., Foley, J.A., 2009b. A safe operating space for humanity. *Nature* 461, 472–475.
- 1745 Rousis, N.I., Zuccato, E., Castiglioni, S., 2016. Monitoring population exposure to pesticides
1746 based on liquid chromatography-tandem mass spectrometry measurement of their
1747 urinary metabolites in urban wastewater: A novel biomonitoring approach. *Sci. Total*
1748 *Environ.* 571, 1349–1357. <https://doi.org/10.1016/j.scitotenv.2016.07.036>
- 1749 Ruan, T., Jiang, G., 2017. Analytical methodology for identification of novel per- and
1750 polyfluoroalkyl substances in the environment. *TrAC Trends Anal. Chem.* 95, 122–131.
1751 <https://doi.org/10.1016/j.TRAC.2017.07.024>
- 1752 Ruff, M., Mueller, M.S., Loos, M., Singer, H.P., 2015. Quantitative target and systematic non-
1753 target analysis of polar organic micro-pollutants along the river Rhine using high-
1754 resolution mass- spectrometry - Identification of unknown sources and compounds.
1755 *Water Res.* 87, 145–154.
- 1756 Ruttkies, C., Schymanski, E.L., Wolf, S., Hollender, J., Neumann, S., 2016. MetFrag relaunched:
1757 incorporating strategies beyond in silico fragmentation. *J. Cheminform.* 8, 3.
1758 <https://doi.org/10.1186/s13321-016-0115-9>
- 1759 SANTE/11945, 2015. Guidance document on analytical quality control and method validation
1760 procedures for pesticide residues analysis in food and feed.
- 1761 Schaidler, L.A., Balan, S.A., Blum, A., Andrews, D.Q., Strynar, M.J., Dickinson, M.E.,
1762 Lunderberg, D.M., Lang, J.R., Peaslee, G.F., 2017. Fluorinated Compounds in U.S. Fast
1763 Food Packaging. *Environ. Sci. Technol. Lett.* 4, 105–111
- 1764 Schmidt, C.K., Brauch, H.-J., 2008. N,N-Dimethylsulfamide as Precursor for N-
1765 Nitrosodimethylamine (NDMA) Formation upon Ozonation and its Fate During Drinking
1766 Water Treatment. *Environ. Sci. Technol.* 42, 6340–6346.
1767 <https://doi.org/10.1021/es7030467>
- 1768 Schollée, J.E., Schymanski, E.L., Avak, S.E., Loos, M., Hollender, J., 2015. Prioritizing Unknown
1769 Transformation Products from Biologically- Treated Wastewater Using High-Resolution
1770 Mass Spectrometry, Multivariate Statistics, and Metabolic Logic. *Anal. Chem.* 87, 12121–
1771 12129.
- 1772 Schollée, J.E., Schymanski, E.L., Hollender, J., 2016. Statistical Approaches for LC-HRMS Data To
1773 Characterize, Prioritize, and Identify Transformation Products from Water Treatment
1774 Processes, in: *Assessing Transformation Products of Chemicals by Non-Target and*
1775 *Suspect Screening – Strategies and Workflows Volume 1.* pp. 45–65.
1776 <https://doi.org/10.1021/bk-2016-1241.ch004>
- 1777 Schulze, S., Sättler, D., Neumann, M., Arp, H.P.H., Reemtsma, T., Berger, U., 2018. Using REACH
1778 registration data to rank the environmental emission potential of persistent and mobile
1779 organic chemicals. *Sci. Total Environ.* 625, 1122–1128.
1780 <https://doi.org/10.1016/j.scitotenv.2017.12.305>
- 1781 Schulze, T., Schymanski, E.L., Stravs, M.A., Neumann, S., Krauss, M., Singer, H., Hug, C.,
1782 Gallampois, C.M.J., Hollender, J., Slobodnik, J., Brack, W., 2012. NORMAN MassBank
1783 Towards a community-driven, open-access accurate mass spectral database for the
1784 identification of emerging pollutants. *Norman Bull.* 9–11.
- 1785 Schymanski, E.L., Gallampois, C.M.J., Krauss, M., Meringer, M., Neumann, S., Schulze, T., Wolf,

1786 S., Brack, W., 2012. Consensus structure elucidation combining GC/EI-MS, structure
1787 generation, and calculated properties. *Anal Chem* 84. <https://doi.org/10.1021/ac203471y>

1788 Schymanski, E.L., Jeon, J., Gulde, R., Fenner, K., Ruff, M., Singer, H.P., Hollender, J., 2014a.
1789 Identifying Small Molecules via High Resolution Mass Spectrometry: Communicating
1790 Confidence. *Environ. Sci. Technol.* 48, 2097–2098.

1791 Schymanski, E.L., Ruttkies, C., Krauss, M., Brouard, C., Kind, T., Dührkop, K., Allen, F., Vaniya, A.,
1792 Verdegem, D., Böcker, S., Rousu, J., Shen, H., Tsugawa, H., Sajed, T., Fiehn, O.,
1793 Ghesquière, B., Neumann, S., 2017. Critical Assessment of Small Molecule Identification
1794 2016: automated methods. *J. Cheminform.* 9, 1–21. [https://doi.org/10.1186/s13321-017-](https://doi.org/10.1186/s13321-017-0207-1)
1795 [0207-1](https://doi.org/10.1186/s13321-017-0207-1)

1796 Schymanski, E.L., Singer, H.P., Longrée, P., Loos, M., Ruff, M., Stravs, M.A., Ripollés Vidal, C.,
1797 Hollender, J., 2014b. Strategies to Characterize Polar Organic Contamination in
1798 Wastewater: Exploring the Capability of High Resolution Mass Spectrometry. *Environ. Sci.*
1799 *Technol.* 48, 1811–1818.

1800 Schymanski, E.L., Singer, H.P., Slobodnik, J., Ipolyi, I.M., Oswald, P., Krauss, M., Schulze, T.,
1801 Haglund, P., Letzel, T., Grosse, S., Thomaidis, N.S., Bletsou, A., Zwiener, C., Ibáñez, M.,
1802 Portolés, T., Boer, R. De, Reid, M.J., Onghena, M., Kunkel, U., Schulz, W., Guillon, A.,
1803 Noyon, N., Leroy, G., Bados, P., Bogialli, S., Stipančič, D., Rostkowski, P., Hollender, J.,
1804 2015. Non-target screening with high-resolution mass spectrometry : critical review using
1805 a collaborative trial on water analysis. *Anal. Bioanal. Chem.* 407, 6237–6255.

1806 Schymanski, E.L., Williams, A.J., 2017. Open Science for Identifying “Known Unknown”
1807 Chemicals. *Environ. Sci. Technol.* 51, 5357–5359.
1808 <https://doi.org/10.1021/acs.est.7b01908>

1809 SCORE (2010) Sewage analysis CORE group Europe [WWW Document], n.d. .
1810 <http://www.webcitation.org/6tIO1NrbC>. URL <http://score-cost.eu/> (accessed 5.28.18).

1811 Sharma, B.M., Bharat, G.K., Tayal, S., Larssen, T., Bečanová, J., Karásková, P., Whitehead, P.G.,
1812 Futter, M.N., Butterfield, D., Nizzetto, L., 2016. Perfluoroalkyl substances (PFAS) in river
1813 and ground/drinking water of the Ganges River basin: Emissions and implications for
1814 human exposure. *Environ. Pollut.* 208, 704–713.
1815 <https://doi.org/10.1016/j.envpol.2015.10.050>

1816 Shi, H.-C., Song, B.-D., Long, F., Zhou, X.-H., He, M., Lv, Q., Yang, H.-Y., 2013. Automated Online
1817 Optical Biosensing System for Continuous Real-Time Determination of Microcystin-LR
1818 with High Sensitivity and Specificity: Early Warning for Cyanotoxin Risk in Drinking Water
1819 Sources. *Environ. Sci. Technol.* 47, 4434–4441. <https://doi.org/10.1021/es305196f>

1820 Singer, A.C., Shaw, H., Rhodes, V., Hart, A., 2016. Review of antimicrobial resistance in the
1821 environment and its relevance to environmental regulators. *Front. Microbiol.* 7, 1–22.
1822 <https://doi.org/10.3389/fmicb.2016.01728>

1823 Sjerps, R.M.A., Vughs, D., van Leerdam, J.A., ter Laak, T.L., van Wezel, A.P., 2016. Data-driven
1824 prioritization of chemicals for various water types using suspect screening LC-HRMS.
1825 *Water Res.* 93, 254–264. <https://doi.org/10.1016/j.watres.2016.02.034>

1826 Smith, C.A., Want, E.J., O'Maille, G., Abagyan, R., Siuzdak, G., 2006. XCMS: processing mass
1827 spectrometry data for metabolite profiling using nonlinear peak alignment, matching,
1828 and identification. *Anal Chem* 78, 779–787. <https://doi.org/10.1021/ac051437y>

1829 Stanstrup, J., Neumann, S., Vrhovšek, U., 2015. PredRet: Prediction of Retention Time by Direct
1830 Mapping between Multiple Chromatographic Systems. *Anal. Chem.* 87, 9421–9428.
1831 <https://doi.org/10.1021/acs.analchem.5b02287>

- 1832 Steffen, W., Richardson, K., Rockström, J., Cornell, S.E., Fetzer, I., Bennett, E.M., Biggs, R.,
 1833 Carpenter, S.R., de Vries, W., de Wit, C.A., Folke, C., Gerten, D., Heinke, J., Mace, G.M.,
 1834 Persson, L.M., Ramanathan, V., Reyers, B., Sörlin, S., 2015. Planetary boundaries: Guiding
 1835 human development on a changing planet. *Science* (80-.). 347, 1259855.
 1836 <https://doi.org/10.1126/science.1259855>
- 1837 Storey, M. V., van der Gaag, B., Burns, B.P., 2011. Advances in on-line drinking water quality
 1838 monitoring and early warning systems. *Water Res.* 45, 741–747.
 1839 <https://doi.org/10.1016/j.watres.2010.08.049>
- 1840 Strynar, M., Dagnino, S., McMahan, R., Liang, S., Lindstrom, A., Andersen, E., McMillan, L.,
 1841 Thurman, M., Ferrer, I., Ball, C., 2015. Identification of Novel Perfluoroalkyl Ether
 1842 Carboxylic Acids (PFECAs) and Sulfonic Acids (PFESAs) in Natural Waters Using Accurate
 1843 Mass Time-of-Flight Mass Spectrometry (TOFMS). *Environ. Sci. Technol.* 49, 11622–
 1844 11630. <https://doi.org/10.1021/acs.est.5b01215>
- 1845 Sun, M., Arevalo, E., Strynar, M., Lindstrom, A., Richardson, M., Kearns, B., Pickett, A., Smith,
 1846 C., Knappe, D.R.U., 2016. Legacy and Emerging Perfluoroalkyl Substances Are Important
 1847 Drinking Water Contaminants in the Cape Fear River Watershed of North Carolina.
 1848 *Environ. Sci. Technol. Lett.* 3, 415–419. <https://doi.org/10.1021/acs.estlett.6b00398>
- 1849 Telving, R., Hasselström, J.B., Andreassen, M.F., 2016. Targeted toxicological screening for
 1850 acidic, neutral and basic substances in postmortem and antemortem whole blood using
 1851 simple protein precipitation and UPLC-HR-TOF-MS. *Forensic Sci. Int.* 266, 453-461.
- 1852 ter Laak, T.L., Kooij, P.J.F., Tolkamp, H., Hofman, J., 2014. Different compositions of
 1853 pharmaceuticals in Dutch and Belgian rivers explained by consumption patterns and
 1854 treatment efficiency. *Environ. Sci. Pollut. Res.* 21, 12843–12855.
 1855 <https://doi.org/10.1007/s11356-014-3233-9>
- 1856 ter Laak, T.L., Puijker, L.M., van Leerdam, J.A., Raat, K.J., Kolkman, A., de Voogt, P., van Wezel,
 1857 A.P., 2012. Broad target chemical screening approach used as tool for rapid assessment
 1858 of groundwater quality. *Sci. Total Environ.* 427, 308–313.
 1859 <https://doi.org/10.1016/j.scitotenv.2012.04.013>
- 1860 ter Laak, T.L., van Leerdam, J.A., Sjerps, R., Vughs, D., 2015. Dialog between environmental
 1861 occurrence data and REACH II; Report number KWR 400134. Nieuwegein, the
 1862 Netherlands.
- 1863 Thomas, K. V., Bijlsma, L., Castiglioni, S., Covaci, A., Emke, E., Grabic, R., Hernández, F., Karolak,
 1864 S., Kasprzyk-Hordern, B., Lindberg, R.H., Lopez de Alda, M., Meierjohann, A., Ort, C., Pico,
 1865 Y., Quintana, J.B., Reid, M., Rieckermann, J., Terzic, S., van Nuijs, A.L.N., de Voogt, P.,
 1866 2012. Comparing illicit drug use in 19 European cities through sewage analysis. *Sci. Total*
 1867 *Environ.* 432, 432–439. <https://doi.org/10.1016/j.scitotenv.2012.06.069>
- 1868 Thomas, K. V., Reid, M.J., 2011. What else can the analysis of sewage for urinary biomarkers
 1869 reveal about communities? *Environ. Sci. Technol.* 45, 7611–7612.
 1870 <https://doi.org/10.1021/es202522d>
- 1871 Thurman, E.M., Ferrer, I., Blotvogel, J., Borch, T., 2014. Analysis of Hydraulic Fracturing
 1872 Flowback and Produced Waters Using Accurate Mass: Identification of Ethoxylated
 1873 Surfactants. *Anal. Chem.* 86, 9653–9661. <https://doi.org/10.1021/ac502163k>
- 1874 Tice, R.R., Austin, C.P., Kavlock, R.J., Bucher, J.R., 2013. Improving the Human Hazard
 1875 Characterization of Chemicals: A Tox21 Update. *Environ. Heal. Perspect* 121, 756–765.
- 1876 Touseva, Z., Oswald, P., Slobodnik, J., Blaha, L., Muz, M., Hu, M., Brack, W., Krauss, M., Di
 1877 Paolo, C., Tarcai, Z., Seiler, T.-B., Hollert, H., Koprivica, S., Ahel, M., Schollée, J.E.,

Con formato: Español

- 1878 Hollender, J., Suter, M.J.-F., Hidasi, A.O., Schirmer, K., Sonavane, M., Ait-Aissa, S., Creusot,
1879 N., Brion, F., Froment, J., Almeida, A.C., Thomas, K., Tollefsen, K.E., Tufi, S., Ouyang, X.,
1880 Leonards, P., Lamoree, M., Torrens, V.O., Kolkman, A., Schriks, M., Spirhanzlova, P.,
1881 Tindall, A., Schulze, T., 2017. European demonstration program on the effect-based and
1882 chemical identification and monitoring of organic pollutants in European surface waters.
1883 *Sci. Total Environ.* 601–602, 1849–1868. <https://doi.org/10.1016/j.scitotenv.2017.06.032>
- 1884 Towhata, I., Uchimura, T., 2013. Low-cost and Simple Early Warning Systems of Slope
1885 Instability, in: Sassa, K., Rouhban, B., Briceño, S., McSaveney, M., He, B. (Eds.), *Landslides:
1886 Global Risk Preparedness*. Springer Berlin Heidelberg, Berlin, Heidelberg, pp. 213–225.
1887 https://doi.org/10.1007/978-3-642-22087-6_14
- 1888 Tränckner, J., Koegst, T., 2010. Demographic effects on domestic pharmaceutical emissions in
1889 Germany, in: NOVATECH 2010, 7th International Conference on Sustainable Techniques
1890 and Strategies for Urban Water Management. pp. 1–9.
- 1891 Tscharke, Chen, C., Gerber, J.P., White, J.M., 2016. Temporal trends in drug use in Adelaide,
1892 South Australia by wastewater analysis. *Sci. Total Environ.* 565, 384–391.
1893 <https://doi.org/10.1016/j.scitotenv.2016.04.183>
- 1894 Turner, M.C., Vineis, P., Seleiro, E., Dijmarescu, M., Balshaw, D., Bertollini, R., Chadeau-Hyam,
1895 M., Gant, T., Gulliver, J., Jeong, A., Kyrtopoulos, S., Martuzzi, M., Miller, G.W., Nawrot, T.,
1896 Nieuwenhuijsen, M., Phillips, D.H., Probst-Hensch, N., Samet, J., Vermeulen, R.,
1897 Vlaanderen, J., Vrijheid, M., Wild, C., Kogevinas, M., 2018. EXPOsOMICS: final policy
1898 workshop and stakeholder consultation. *BMC Public Health* 18, 260.
1899 <https://doi.org/10.1186/s12889-018-5160-z>
- 1900 van der Hooft, J.J.J., Wandy, J., Barrett, M.P., Burgess, K.E. V., Rogers, S., 2016. Topic modeling
1901 for untargeted substructure exploration in metabolomics. *Proc. Natl. Acad. Sci.* 113,
1902 13738–13743. <https://doi.org/10.1073/pnas.1608041113>
- 1903 van der Veen, I., de Boer, J., 2012. Phosphorus flame retardants: Properties, production,
1904 environmental occurrence, toxicity and analysis. *Chemosphere* 88, 1119–1153.
1905 <https://doi.org/10.1016/j.chemosphere.2012.03.067>
- 1906 van Leerdam, J.A., Vervoort, J., Stroomberg, G., de Voogt, P., 2014. Identification of unknown
1907 microcontaminants in Dutch river water by liquid chromatography-high resolution mass
1908 spectrometry and nuclear magnetic resonance spectroscopy. *Environ. Sci. Technol.* 48,
1909 12791–12799.
- 1910 van Leeuwen, S.P.J., van Bavel, B., Abad, E., Leslie, H.A., Fiedler, H., de Boer, J. 2013. POPs
1911 analysis reveals issues in bringing laboratories in developing countries to a higher quality
1912 level *TrAC Trends Anal. Chem.* 46, 198–206.
- 1913 van Nuijs, A.L.N., Castiglioni, S., Tarcomnicu, I., Postigo, C., Lopez de Alda, M., Neels, H.,
1914 Zuccato, E., Barcelo, D., Covaci, A., 2011. Illicit drug consumption estimations derived
1915 from wastewater analysis: A critical review. *Sci. Total Environ.* 409, 3564–3577.
- 1916 van Nuijs, A.L.N., Covaci, A., Beyers, H., Bervoets, L., Blust, R., Verpooten, G., Neels, H., Jorens,
1917 P.G., 2015. Do concentrations of pharmaceuticals in sewage reflect prescription figures?
1918 *Environ. Sci. Pollut. Res.* 22, 9110–9118. <https://doi.org/10.1007/s11356-014-4066-2>
- 1919 Vazquez-Roig, P., Kasprzyk-Hordern, B., Blasco, C., Picó, Y., 2014. Stereoisomeric profiling of
1920 drugs of abuse and pharmaceuticals in wastewaters of Valencia (Spain). *Sci. Total
1921 Environ.* 494, 49–57. <https://doi.org/10.1016/j.scitotenv.2014.06.098>
- 1922 Vergeynst, L., Van Langenhove, H., Demeestere, K., 2015. Trends in liquid chromatography
1923 coupled to high-resolution mass spectrometry for multi-residue analysis of organic

- 1924 micropollutants in aquatic environments. *TrAC - Trends Anal. Chem.* 67, 192–208.
- 1925 Verliefe, A.R.D., Cornelissen, E.R., Heijman, S.G.J., Verberk, J.Q.J.C., Amy, G.L., Van der
1926 Bruggen, B., van Dijk, J.C., 2009. Construction and validation of a full-scale model for
1927 rejection of organic micropollutants by NF membranes. *J. Memb. Sci.* 339, 10–20.
1928 <https://doi.org/10.1016/j.memsci.2009.03.038>
- 1929 Vestergren, R., Cousins, I.T., 2009. Tracking the pathways of human exposure to
1930 perfluorocarboxylates. *Environ. Sci. Technol.* 43, 5565–5575.
1931 <https://doi.org/10.1021/es900228k>
- 1932 Vinaixa, M., Schymanski, E.L., Neumann, S., Navarro, M., Salek, R.M., Yanes, O., 2016. Mass
1933 spectral databases for LC/MS- and GC/MS-based metabolomics: State of the field and
1934 future prospects. *TrAC Trends Anal. Chem.* 78, 23–35.
1935 <https://doi.org/10.1016/j.trac.2015.09.005>
- 1936 Von der Ohe, P.C., Dulio, V., Slobodnik, J., De Deckere, E., Kühne, R., Ebert, R.-U., Ginebreda,
1937 A., De Cooman, W., Schüürmann, G., Brack, W., 2011. A new risk assessment approach
1938 for the prioritization of 500 classical and emerging organic microcontaminants as
1939 potential river basin specific pollutants under the European Water Framework Directive.
1940 *Sci. Total Environ.* 409, 2064–2077.
- 1941 Vughs, D., Baken, K.A., Kolkman, A., Martijn, A.J., de Voogt, P., 2018. Application of effect-
1942 directed analysis to identify mutagenic nitrogenous disinfection by-products of advanced
1943 oxidation drinking water treatment. *Environ. Sci. Pollut. Res.* 25, 3951–3964.
1944 <https://doi.org/10.1007/s11356-016-7252-6>
- 1945 Wagner, M., Lambert, S., 2018. Freshwater microplastics: emerging environmental
1946 contaminants? In: *The handbook of environmental chemistry*, Vol. 58. Springer.
1947 ISBN:978-3-319-61614-8
- 1948 Wambaugh, J.F., Wang, A., Dionisio, K.L., Frame, A., Egeghy, P., Judson, R., Setzer, R.W., 2014.
1949 High Throughput Heuristics for Prioritizing Human Exposure to Environmental Chemicals.
1950 *Environ. Sci. Technol.* 48, 12760–12767.
- 1951 Wang, M., Carver, J.J., Phelan, V. V., Sanchez, L.M., Garg, N., Peng, Y., Nguyen, D.D., Watrous,
1952 J., Kapon, C.A., Luzzatto-Knaan, T., Porto, C., Bouslimani, A., Melnik, A. V., Meehan, M.J.,
1953 Liu, W.T., Crüsemann, M., Boudreau, P.D., Esquenazi, E., Sandoval-Calderón, M., Kersten,
1954 R.D., Pace, L.A., Quinn, R.A., Duncan, K.R., Hsu, C.C., Floros, D.J., Gavilan, R.G., Kleigrewe,
1955 K., Northen, T., Dutton, R.J., Parrot, D., Carlson, E.E., Aigle, B., Michelsen, C.F., Jelsbak, L.,
1956 Sohlenkamp, C., Pevzner, P., Edlund, A., McLean, J., Piel, J., Murphy, B.T., Gerwick, L.,
1957 Liaw, C.C., Yang, Y.L., Humpf, H.U., Maansson, M., Keyzers, R.A., Sims, A.C., Johnson, A.R.,
1958 Sidebottom, A.M., Sedio, B.E., Klitgaard, A., Larson, C.B., Boya, C.A.P., Torres-Mendoza,
1959 D., Gonzalez, D.J., Silva, D.B., Marques, L.M., Demarque, D.P., Pociute, E., O'Neill, E.C.,
1960 Briand, E., Helfrich, E.J.N., Granatosky, E.A., Glukhov, E., Ryffel, F., Houson, H., Mohimani,
1961 H., Kharbush, J.J., Zeng, Y., Vorholt, J.A., Kurita, K.L., Charusanti, P., McPhail, K.L., Nielsen,
1962 K.F., Vuong, L., Elfeki, M., Traxler, M.F., Engene, N., Koyama, N., Vining, O.B., Baric, R.,
1963 Silva, R.R., Mascuch, S.J., Tomasi, S., Jenkins, S., Macherla, V., Hoffman, T., Agarwal, V.,
1964 Williams, P.G., Dai, J., Neupane, R., Gurr, J., Rodríguez, A.M.C., Lamsa, A., Zhang, C.,
1965 Dorrestein, K., Duggan, B.M., Almaliti, J., Allard, P.M., Phapale, P., Nothias, L.F.,
1966 Alexandrov, T., Litaudon, M., Wolfender, J.L., Kyle, J.E., Metz, T.O., Peryea, T., Nguyen,
1967 D.T., VanLeer, D., Shinn, P., Jadhav, A., Müller, R., Waters, K.M., Shi, W., Liu, X., Zhang, L.,
1968 Knight, R., Jensen, P.R., Palsson, B., Pogliano, K., Lington, R.G., Gutiérrez, M., Lopes,
1969 N.P., Gerwick, W.H., Moore, B.S., Dorrestein, P.C., Bandeira, N., 2016. Sharing and
1970 community curation of mass spectrometry data with Global Natural Products Social
1971 Molecular Networking. *Nat. Biotechnol.* 34, 828–837. <https://doi.org/10.1038/nbt.3597>

- 1972 Wang, Z., DeWitt, J.C., Higgins, C.P., Cousins, I.T., 2017. A Never-Ending Story of Per- and
1973 Polyfluoroalkyl Substances (PFASs)? *Environ. Sci. Technol.* 51, 2508-2518
- 1974 Weiss, J.M., Hamers, T., Thomas, K.V., van der Linden, S., Leonards, P.E.G., Lamoree, M.H.,
1975 2009. Masking effect of anti-androgens on androgenic activity in European river sediment
1976 unveiled by effect-direct analysis. *Anal. Bioanal. Chem.* 394, 1385-1397 DOI
1977 10.1007/s00216-009-2807-8
- 1978 Weiss, J.M., Andersson, P.L., Zhang, J., Simon, E., Leonards, P.E.G., Hamers, T., Lamoree, M.H.,
1979 2015. Tracing thyroid hormone-disrupting compounds: database compilation and
1980 structure-activity evaluation for an effect-directed analysis of sediment. *Anal. Bioanal.*
1981 *Chem.* 5625-5634. <https://doi.org/10.1007/s00216-015-8736-9>
- 1982 Wick, A., Wagner, M., Ternes, T.A., 2011. Elucidation of the Transformation Pathway of the
1983 Opium Alkaloid Codeine in Biological Wastewater Treatment. *Environ. Sci. Technol.* 45,
1984 3374-3385. <https://doi.org/10.1021/es103489x>
- 1985 Wicker, J., Lorschach, T., Gütlein, M., Schmid, E., Latino, D., Kramer, S., Fenner, K., 2015.
1986 enviPath - The environmental contaminant biotransformation pathway resource. *Nucleic*
1987 *Acids Res.* 44, D502-D508. <https://doi.org/10.1093/nar/gkv1229>
- 1988 Willach, S., Brauch, H.-J., Lange, F.T., 2016. Contribution of selected perfluoroalkyl and
1989 polyfluoroalkyl substances to the adsorbable organically bound fluorine in German rivers
1990 and in a highly contaminated groundwater. *Chemosphere* 145, 342-350.
1991 <https://doi.org/10.1016/j.chemosphere.2015.11.113>
- 1992 Williams, A.J., Grulke, C.M., Edwards, J., McEachran, A.D., Mansouri, K., Baker, N.C., Patlewicz,
1993 G., Shah, I., Wambaugh, J.F., Judson, R.S., Richard, A.M., 2017. The CompTox Chemistry
1994 Dashboard: A community data resource for environmental chemistry. *J. Cheminform.* 9,
1995 1-27. <https://doi.org/10.1186/s13321-017-0247-6>
- 1996 Wode, F., Baar, P. Van, Dünnbier, U., Hecht, F., Taute, T., Jekel, M., Reemtsma, T., 2015. Search
1997 for over 2000 current and legacy micropollutants on a wastewater infiltration site with a
1998 UPLC-high resolution MS target screening method. *Water Res.* 69, 274-283.
- 1999 Wu, M., Xiang, J., Que, C., Chen, F., Xu, G., 2015. Occurrence and fate of psychiatric
2000 pharmaceuticals in the urban water system of Shanghai, China. *Chemosphere* 138, 486-
2001 493. <https://doi.org/10.1016/j.chemosphere.2015.07.002>
- 2002 Xiao, F., 2017. Emerging poly- and perfluoroalkyl substances in the aquatic environment: A
2003 review of current literature. *Water Res.* 124, 482-495.
- 2004 Yang, X., Neta, P., Stein, S.E., 2017. Extending a Tandem Mass Spectral Library to Include MS2
2005 Spectra of Fragment Ions Produced In-Source and MSn Spectra. *J. Am. Soc. Mass*
2006 *Spectrom.* 28, 2280-2287. <https://doi.org/10.1007/s13361-017-1748-2>
- 2007 Yang, Z., Castrignanò, E., Estrela, P., Frost, C.G., Kasprzyk-Hordern, B., 2016. Community
2008 Sewage Sensors towards Evaluation of Drug Use Trends: Detection of Cocaine in
2009 Wastewater with DNA-Directed Immobilization Aptamer Sensors. *Sci. Rep.* 6, 1-10.
2010 <https://doi.org/10.1038/srep21024>
- 2011 Yang, Z., Kasprzyk-Hordern, B., Frost, C.G., Estrela, P., Thomas, K. V., 2015a. Community
2012 sewage sensors for monitoring public health. *Environ. Sci. Technol.* 49, 5845-5846.
2013 <https://doi.org/10.1021/acs.est.5b01434>
- 2014 Yang, Z., Kasprzyk-Hordern, B., Goggins, S., Frost, C.G., Estrela, P., 2015b. A novel
2015 immobilization strategy for electrochemical detection of cancer biomarkers: DNA-
2016 directed immobilization of aptamer sensors for sensitive detection of prostate specific
2017 antigens. *Analyst* 140, 2628-2633. <https://doi.org/10.1039/c4an02277g>

Con formato: Inglés (americano)

2018 Yao, L., Wang, Y., Tong, L., Li, Y., Deng, Y., Guo, W., Gan, Y., 2015. Seasonal variation of
2019 antibiotics concentration in the aquatic environment: a case study at Jiangnan Plain,
2020 central China. *Sci. Total Environ.* 527, 56–64.
2021 <https://doi.org/10.1016/j.scitotenv.2015.04.091>

2022 Zahn, D., Frömel, T., Knepper, T.P., 2016. Halogenated methanesulfonic acids: A new class of
2023 organic micropollutants in the water cycle. *Water Res.* 101, 292–299.
2024 <https://doi.org/10.1016/j.watres.2016.05.082>

2025 Zhou, Z., Shen, X., Tu, J., Zhu, Z., 2016. Large-Scale Prediction of Collision Cross-Section Values
2026 for Metabolites in Ion Mobility-Mass Spectrometry. *Anal. Chem.* 88, 11084–11094.
2027 <https://doi.org/10.1021/acs.analchem.6b03091>

2028 Zonja, B., Delgado, A., Pérez, S., Barceló, D., 2015. LC-HRMS suspect screening for detection-
2029 based prioritization of iodinated contrast media photodegradates in surface waters.
2030 *Environ. Sci. Technol.* 49, 3464–3472.

2031 Zwart, N., Lamoree, M.H., Houtman, C.J., de Boer, J., Kool, J., Hamers, T., 2018a. Development
2032 of a luminescent mutagenicity test for high-throughput screening of aquatic samples.
2033 *Toxicol. Vitro.* 46, 350–360. <https://doi.org/10.1016/j.tiv.2017.09.005>

2034 Zwart, N., Nio, S.L., Houtman, C.J., De Boer, J., Kool, J., Hamers, T., Lamoree, M.H., 2018b.
2035 High-Throughput Effect-Directed Analysis Using Downscaled in Vitro Reporter Gene
2036 Assays to Identify Endocrine Disruptors in Surface Water. *Environ. Sci. Technol.* 52, 4367–
2037 4377. <https://doi.org/10.1021/acs.est.7b06604>

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